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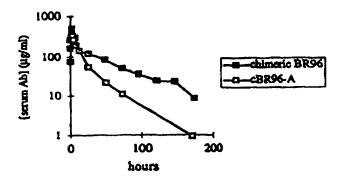
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(54) Title: A METHOD FOR INHIBITING IMMUNOGLOBULIN-INDUCED TOXICITY RESULTING FROM THE USE OF IMMUNOGLOBULINS IN THERAPY AND IN VIVO DIAGNOSIS



Plasma clearance in high LeY expressing dogs chimeric versus constant region mutant of cBR96-2.

(57) Abstract

The present invention provides a method for inhibiting immunoglobulin-induced toxicity resulting from immunotherapy in a subject comprising administering an immunoglobulin or Ig fusion protein molecule to the subject, the immunoglobulin molecule having a variable region and a constant region, the immunoglobulin molecule being modified prior to administration by inactivation of at least a portion of the constant region.

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5 A METHOD FOR INHIBITING IMMUNOGLOBULIN-INDUCED TOXICITY RESULTING FROM THE USE OF IMMUNOGLOBULINS IN THERAPY AND IN VIVO DIAGNOSIS

Throughout this application various publications are referenced. The disclosures of these publications in their entireties are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this invention pertains.

15 TECHNICAL FIELD OF THE INVENTION

The present invention relates to methods for inhibiting or reducing immunoglobulin-induced toxicity resulting from therapy or in vivo diagnosis. Specifically, in lieu of using unmodified antibodies or recombinant binding proteins for in vivo use, the invention provides the use of modified antibodies or recombinant binding proteins which have been structurally altered in the constant domain so that upon administration immunoglobulin-induced toxicity is reduced or inhibited.

BACKGROUND OF THE INVENTION

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Over the years investigators have attempted to harness the immune system for therapeutic use. Immunoglobulin (Ig) molecules which constitute an important part of the immune system are of great interest because they (1) react with a diverse family of ligands, (2) possess different effector functions and (3) are of great biological importance. Despite its potential, a persistent problem with

immunoglobulin immunotherapy has been, among other problems, the toxic effect to normal cells of using antibodies which recognize both normal and diseased cells. This problem is far-reaching because the majority of antibodies presently available recognize a target located on both normal and diseased cells (Slavin-Chiorini, et al., Int. J. Cancer 53: 97-103 (1993)).

The constant region can promote cell death through antibody dependent cell mediated cytotoxicity (ADCC) or by complement dependent cytotoxicity (CDC). Despite the deletion of portions of the constant region, particularly the CH₂ domain, the antigen binding function can be retained (D. Yelton, M. Scharf, Mutant monoclonal antibody with alterations in biological functions, J. Exp. Methods 156:1131-1148 (1982)).

Others have generated a CH₂-deleted antibody (Mueller et al., Proc. Natl. Acad. Sci. USA 87: 5702-5705 (1990)). Their findings provide that the CH₂-deleted antibody was cleared from the blood of tumor-bearing mice much faster than the corresponding intact antibody. Other in vivo findings also confirmed that a CH₂-deleted antibody, designated ch14.18DCH2, is a potentially useful reagent for radioimmunodetection of human tumors because of its reduced immunogenicity, increased target specificity, and rapid clearance from circulation (Mueller et al., Proc. Natl. Acad. Sci. USA 87: 5702-5705 (1990).

Generally, whole antibody molecules are composed of two heavy (H) and two light (L) chains which are held together by covalent bonds (disulfide) and non-covalent interactions. Each chain contains a variable region (V) and a constant region (C). The variable regions at the amino termini of the two chains form the antigen binding region. The constant region of the H chain has three components or domains. Occasionally, the first constant region domain (CH₁) interacts with the C region of the L chain through hydrophobic interactions and generally a disulfide bond,

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depending on isotype. The next C region stretch is the hinge-acting disulfide bond stably introduced between two H chains. The second constant region domain (CH₂) is adjacent to the hinge region. CH₂ contains sequences important for effector functions of the antibody, such as the sequences responsible for complement fixation, and Fc receptor binding The third constant region domain (CH₃) is located at the carboxyl terminus of the H chain, and is considered to play an important role in H chain assembly as well as some C region functions.

Today many antibodies in clinical trials are directed against tumor associated antigens. Most tumor associated antigens are not tumor specific but are also generally found on the cell surface of some normal, non-tumorigenic cells. The clinical use of some antibodies directed against tumor associated antigens are limited because of the toxicity associated with their use. Therefore, there is a need for methods for inhibiting toxicity associated with immunoglobulin use in the field of disease therapy (e.g., therapy for tumors, kidney disease, and the like) and in vivo diagnosis.

We addressed this need by discovering methods for inhibiting or reducing toxicity to normal cells generally associated with immunoglobulin immunotherapy or in vivo diagnosis, wherein the immunoglobulin recognizes both diseased and normal cells. Our discovery involves generating immunoglobulin molecules or Ig fusion proteins having structurally altered constant regions which inhibit or reduce immunoglobulin-induced toxicity.

25 SUMMARY OF THE INVENTION

The present invention provides methods for inhibiting immunoglobulin-induced toxicity by using known immunoglobulin or Ig fusion protein molecules which are structurally altered in their constant regions so that the resulting structurally altered

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immunoglobulin or Ig fusion protein molecules exhibit reduced or inhibited toxicity in vivo compared to their original unmodified counterparts.

Structural alteration of the constant region may be effected in a number of ways as long as it results in reducing or inhibiting immunoglobulin-induced toxicity.

In accordance with the practice of the invention, structural alteration of the constant region is effected by deletion of the entire constant region. In another embodiment, only the CH₂ domain is deleted. In another embodiment, only that portion of the CH₂ domain that binds the Fc receptor is deleted. In yet another embodiment, only that portion of the CH₂ domain that binds the complement component Clq is deleted. Alternatively, in another embodiment, multiple deletions in discrete Fc receptor and complement component binding domains are effected.

15 Alternatively, structural alteration is effected by single or multiple mutations in the CH₂ domain such as amino acid insertions and substitutions. The mutation or mutations must result in inhibiting immunoglobulin-induced toxicity. By way of example, the amino acids in multiple toxicity associated domains in the constant region can be altered so as to render the constant region unable to mediate a ADCC response or activate complement thereby inhibiting immunoglobulin induced toxicity resulting from immunotherapy. Alternatively, multiple amino acids in a single toxicity associated domain in the constant region can be altered.

Further alternatively, structural alteration can be effected by isotype switching resulting in an altered immunoglobulin molecule that either does not induce toxicity or induces some limited toxicity but does not cause a harmful effect. For example, isotype switching can result in the constant region being unable to mediate a CDC or ADCC response or some other activity which mediates toxicity.

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BRIEF DESCRIPTION OF THE FIGURES

Figure 1 is a line graph showing plasma clearance in high Le^Y expressing dogs using chimeric BR96 versus constant region mutant of cBR96-2.

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Figure 2 is a schematic diagram of a plasmid designated pTWD-cJVK.L1 including the chimeric (c)BR96-light chain (SEQ ID NO. 11).

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Figure 3 is a schematic diagram of a plasmid designated pD16hJ1.L1 including the

human (h)BR96-light chain (SEQ ID NO. 13).

Figure 4 is a schematic diagram of a plasmid, designated pD17-hJm14-dCH2.H1, of

hBR96-2A (i.e., human mutant BR96 having the H1, H2, and H3 mutations and the

CH₂ deletion (PCT Application No. 95/305444, published March 6, 1996)).

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Figure 5 is a schematic diagram of a plasmid, designated pD17-cJ-dCH2.H1, of cBR96-A (SEQ ID NO. 10) (i.e., chimeric BR96 having the CH₂ deletion (PCT

Application No. 95/305444, published March 6, 1996)).

20 Figure 6 is a schematic diagram of a plasmid, designated pD17-cJ.H1, of cBR96.

Figure 7 is a line graph showing the results of an ELISA assay of (1) hBR96-2A-

Dox to Ley (closed diamond), (2) hBR96-2A to Ley (96:0006A2 R/A)(closed

square), (3) hBR96-2A to Ley (96:0006B R/A)(closed triangle), and BR96-Dox to

25 Le^y (X).

Figure 8 is a line graph showing the results of an ELISA assay of (1) BR96-A-Dox to Le^y (closed diamond), (2) chiBR96 to Le^y (closed square), (3) cBR96-A to Le^y

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(96:0003 R/A)(closed triangle), and cBR96-Dox to Ley (X).

Figures 9a-c are schematic diagrams showing the steps for deleting a CH₂ domain.

Figures 10a-c are schematic diagrams showing the construction of BR96 IgG1 CH₂ domain point mutations.

Figure 11 is a schematic diagram showing the construction of the pNg1/14 vector.

Figure 12 is a schematic diagram showing the construction of pD17-hBR96-2.

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Figure 13 is a schematic diagram showing the construction of pD17-hJm14-dCH2.H1.

Figure 14 is the nucleic acid sequence of pD17-cJ-dCH2.H1, the plasmid shown in Figure 5, chimeric BR96 having the CH₂ deletion.

Figure 15 is a line graph showing the results of an ELISA assay comparing whole chiBR96 and deleted CH₂ chiBR96 on Le^y.

20 Figure 16 is a description of the seven structural alterations.

Figure 17 is a schematic diagram of a plasmid designated pD17-hG1b.

Figure 18 is the nucleic acid sequence of pD17-hJm14.H1.

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Figure 19 is the nucleic acid sequence of pD17-hG1b.

Figure 20 is a line graph showing complement dependent cytotoxicity. In the legend, the closed square is hBR96-1; closed diamond is hBR96-2B; closed circle is

hBR96-2C; closed triangle is hBR96-2D; open square is hBR96-2H; open circle is hBR96-2A and open triangle is 2B8, anti-*Pseudonomas aeruginosa* flagella type b mAb. negative control.

Figure 21 is a line graph showing antibody dependent cell-mediated cytotoxity. In the legend, the closed square is hBR96-1; closed diamond is hBR96-2B; closed circle is hBR96-2C; closed triangle is hBR96-2D; open square is hBR96-2H; open circle is hBR96-2A and open triangle is 2B8, anti-Pseudonomas aeruginosa flagella type b monoclonal antibody (mAb), negative control.

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Figure 22 is a line graph showing binding activity of hBR96-2 constant region mutants on LeY-HSA. In the legend, the solid diamond is hBR96-1; solid square is hBR96-2A (CH2 deletion); solid triangle is hBR96-2B (235, 237 mutations); open square is hBR96-2C (318, 320, 322 mutations); open circle is hBR96-2D (331 mutation); and open triangle is hBR96-2H (235, 237, 318, 320, 322, 331 mutations).

Figure 23 is a line graph showing binding activity of hBR96-2 constant region mutants on LNFPIII-BSA. LNFPIII is a lacto-N-fucopentasose, a Lewis X trisaccharide with an additional lactose spacer (V Labs, Covington, LA). In the legend, the solid diamond is hBR96-1; solid square is hBR96-2A (CH2 deletion); solid triangle is hBR96-2B (235, 237 mutations); open square is hBR96-2C (318, 320, 322 mutations); open circle is hBR96-2D (331 mutation); and open triangle is

hBR96-2H (235, 237, 318, 320, 322, 331 mutations).

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Figures 24A and 24B provide a strategy for introducing multiple mutations by RPCR. (A) Diagram of he 1.4 kpb IgG heavy chain region showing the hinge CH₂ and CH₃ domains as boxed regions. Site-specific mutations to be introduced into CH₂ positions L1, L2, and L3 are encoded by complementary sets of mutant PCR

primers (A1 and A2; B1 and B2; and C1 and C2). The asterisks (*) indicate the number of amino acid changes introduced at each L position. The two PCR primers, Rs (Recombination -sense) and Ra (Recombination-antisense), flank the Eco-47-III restriction sites and mediate homologous recombination with vector ends. The 3' ends of the oligonucleotides are represented by arrowheads. (B) A three-way homologous recombination event between fragments RsA2, A1Ra and the linearized vector produces the L1 mutant IgG. Two distally located sets of mutations (L1 and L2) are simultaneously introduced by increasing the number of recombining PCR produces as is shown in the four-way recombination of RsA2, A1B1, B1Ra with vector.

Figure 25 is a gel showing Eco-47-III restriction endonuclease analysis of DNAs prepared from colonies generated by multiple PCR fragment RPCR. Lane M: 1kb ladder DNA marker (GIBCO/BRL Life Science Technology). Lanes 1-12: Twelve randomly selected colonies resulting from quadruple homologous recombination events were used to prepare plasmid and digested with Eco47-III. Clones 1, 2, 6 and 9 contain the fully assembled 1.4 kpb insert.

Figure 26 provides the amino acid sequence for hBR96-2 heavy-chain variable region and the human IgG1 constant region.

Figure 27 provides the amino acid sequence for hBR96-2A heavy-chain variable region and the human IgG1 constant region.

Figure 28 provides the amino acid sequence for chi BR96 heavy-chain variable region and the human IgG1 constant region without the CH₂ domain.

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DETAILED DESCRIPTION OF THE INVENTION

DEFINITIONS

As used herein the term "inhibiting immunoglobulin-induced toxicity" means to reduce or alleviate symptoms generally associated with toxicity caused by immunoglobulin or Ig fusion protein therapy, e.g., toxicity mediated by effector functions of the Fc receptor. For example, BR96 antibody recognizes and binds BR96 antigen which is found at some levels in the gastrointestinal tract and at elevated levels in tumors (as compared to the gastrointestinal tract of normal tissues). The binding of BR96 antibody to BR96 antigen in vivo causes symptoms associated with gastrointestinal toxicity. These symptoms include rapid onset of vomiting, often with blood, and nausea. In humans the bleeding is limited to the fundus of the stomach, causing erosion of the superficial mucosa of the stomach.

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The pathology of the wound is limited and resolves. However, the extreme nature of the nausea and vomiting, unrelieved by anti-emetics, defines it as the dose-limiting toxicity. For highly elevated levels of other antigens found in the central nervous system (CNS), liver, and other locations, the toxicity will be characterized by symptoms other than those described above.

As used herein the term "immunoglobulin molecule" can be produced by B cells or be generated through recombinant engineering or chemical synthetic means. Examples of immunoglobulin molecules include (1) antibodies, e.g., polyclonal and monoclonal antibodies, chimeric or humanized, and (2) recombinant Ig containing binding proteins, e.g., Ig fusion proteins. Recombinant Ig containing binding proteins include cell surface proteins, e.g., CD antigens (in one embodiment, CTLA4), to which an Ig tail is joined.

As used herein the terms "structurally altered" or "structural alteration" means manipulating the constant region so that the resulting molecule or protein exhibits a diminished ability to induce toxicity. Structural alteration can be by chemical modification, proteolytic alteration, or by recombinant genetic means. Recombinant genetic means may include, but is not limited to, the deletion, insertion and substitution of amino acid moieties.

As used herein the terms "multiple toxicity associated domains" means more than one discrete toxicity associated domain. As there appear to be at least two toxicity associated domains in the immunoglobulin molecule, one roughly localized to amino acids 231-238 and another roughly localized to amino acids 310-331, an example of the structural alteration of multiple toxicity associated domains comprises the insertion, substitution or deletion of amino acid residues in both of these domains. This definition excludes structural alterations targeting a single toxicity associated domain.

Merely by way of example, the constant region of the immunoglobulin molecule can be structurally altered so that the molecule no longer mediates a CDC or ADCC response. However, the methods of the invention encompasses the use of structurally altered immunoglobulin molecules regardless of whether it mediates a CDC or ADCC response. The underlying requirement is that the altered molecule must inhibit immunoglobulin-induced toxicity.

Structural alteration can be effected in a number of ways. For example, structural alteration can be effected by deletion of the entire constant region.

Alternatively, structural alteration can be effected by deletion of the entire CH₂ domain of the constant region. In this instance, deletion of the entire CH₂ domain may render the molecule unable to (1) bind an Fc receptor thereby eliminating the

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molecule's possibility of mediating antibody-dependent cellular cytotoxicity (ADCC), (2) bind Clq, or (3) activate complement.

Alternatively, structural alteration can be effected by deletion of only that portion of the CH₂ domain that binds the Fc receptor or complement.

Further alternatively, a single mutation or multiple mutations such as substitutions and insertions in the CH₂ domain can be made. The underlying requirement of any mutation is that it must inhibit, diminish, or block immunoglobulin-induced toxicity. For example, this can be achieved by mutating the constant region such that the altered molecule is rendered unable to mediate a CDC response or an ADCC response, or to activate complement.

Alternatively, structural alteration can be effected by isotype switching (also known as class switching) so that the altered molecule does not induce toxicity in the subject. In one embodiment, the constant region of the immunoglobulin is structurally altered so that it no longer binds the Fc receptor or a complement component, e.g., switching a molecule's original IgG isotype from IgG1 to IgG4. Isotype switching can be effected regardless of species, i.e., an isotype from a non-human being can be switched with an isotype from a human being (E.D. Finkelman et al. (1990) Annu. Rev. Immunol. 8:303-333; T. Honjo et al. (1979) Cell 18: 559-568; T. Honjo et al. In "Immunoglobulin Genes" pp. 124-149 Academic Press, London)).

As used herein the term "Ig fusion protein" means any recombinantly produced antigen or ligand binding domain having a constant region which can be structurally altered.

As used herein "cytotoxic agent" includes antimetabolites, alkylating agents, anthracyclines, antibiotics, anti-mitotic agents, and chemotherapeutic agents. Specific examples within these groups include but are not limited to ricin, doxorubicin, daunorubicin, taxol, ethidium bromide, mitomycin, etoposide, tenoposide, vincristine, vinblastine, colchicine, supporin, gelonin, PE40, bryodin, dihydroxy anthracin dione, actinomycin D, and 1-dehydrotestosterone.

As used herein the term "BR96" refers to (1) the whole BR96 monoclonal antibody disclosed in PCT No. 95/305444, published March 6, 1996, (2) chimeric BR96 monoclonal antibody disclosed in PCT No. 95/305444, published March 6, 1996, or (3) BR96 mutant molecules disclosed in PCT No. 95/305444, published March 6, 1996.

As used herein, "treating" means to (1) provide tumor regression so that the tumor is not palpable for a period of time (standard tumor measurement procedures may be followed (A.B. Miller et al. "Reporting results of cancer treatment" Cancer 47:207-214 (1981)); (2) stabilize the disease; or (3) provide any clinically beneficial effects.

As used herein, an "effective amount" is an amount of the antibody, 20 immunoconjugate, or recombinant molecule which kills cells or inhibits the proliferation thereof.

As used herein, "administering" means oral administration, administration as a suppository, topical contact, intravenous, intraperitoneal, intramuscular or subcutaneous administration, or the implantation of a slow-release device such as a miniosmotic pump, to the subject.

As used herein, "pharmaceutically acceptable carrier" includes any material which when combined with the antibody retains the antibody's specificity or efficacy and is

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non-reactive with the subject's immune system. Examples include, but are not limited to, any of the standard pharmaceutical carriers such as a phosphate buffered saline solution, water, emulsions such as oil/water emulsion, and various types of wetting agents. Other carriers may also include sterile solutions, tablets including coated tablets and capsules.

Typically such carriers contain excipients such as starch, milk, sugar, certain types of clay, gelatin, stearic acid or salts thereof, magnesium or calcium stearate, talc, vegetable fats or oils, gums, glycols, or other known excipients. Such carriers may also include flavor and color additives or other ingredients. Compositions comprising such carriers are formulated by well known conventional methods.

As used herein, "mutation" means a single amino acid or nucleic acid mutation or multiple mutations by whatever means, e.g., homologous recombination, error prone PCR, or site directed mutagenesis.

In order that the invention herein described may be more fully understood, the following description is set forth.

20 METHODS OF THE PRESENT INVENTION

The present invention provides a method for inhibiting immunoglobulin-induced toxicity resulting from the use of immunoglobulin during therapy or in vivo diagnosis. For example, the methods of the invention would be useful to minimize the toxicity associated with prolonged clinical exposure to immunoglobulin use during or after tumor imaging with radiolabeled antibodies.

In accordance with the practice of this invention, the subject includes, but is not limited to, human, equine, porcine, bovine, murine, canine, feline, and avian

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subjects. Other warm blooded animals are also included in this invention.

This method comprises administering an immunoglobulin molecule to the subject.

The immunoglobulin can be IgG, IgM, or IgA. IgG is preferred.

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In one embodiment of the invention, the immunoglobulin molecule recognizes and binds Le^x. In another embodiment, the immunoglobulin recognizes and binds Le^x. In a further embodiment, the immunoglobulin is a monoclonal antibody BR96 produced by the hybridoma deposited on February 22, 1989 with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, MD 20852 and accorded ATCC Accession No.: HB 10036. In yet another embodiment, the immunoglobulin is a chimeric antibody ChiBR96 produced by the hybridoma deposited on May 23, 1990, with the ATCC, 12301 Parklawn Drive, Rockville, MD 20852 and accorded ATCC Accession No.: HB 10460.

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In accordance with the practice of the invention, the immunoglobulin can be a bispecific antibody with a binding specificity for two different antigens, one of the antigens being that with which the monoclonal antibody BR96 produced by the hybridoma having the identifying characteristics of HB 10036 as deposited with the ATCC binds. Also, in accordance with the practice of the invention, the immunoglobulin can be an anti-idiotypic antibody.

As required by the invention, at least a portion of the constant region of the immunoglobulin molecule is structurally altered. Structural alteration can be effected by a number of means. In one embodiment, the entire constant region, i.e., CH₁, CH₂, and CH₃ domains, can be deleted.

In another embodiment, only the CH₂ domain is deleted from the immunoglobulin molecule (e.g., cBR96-A (Figure 5), hBR96-2A (Figure 4). In this embodiment, the

CH₂ deletion may result in a molecule unable to bind the Fc receptor or a complement component.

In another embodiment, only that portion of the CH₂ domain which binds the complement component C1q is deleted. In yet another embodiment, mutations in specific portions of the CH₂ domain are made. For example, the immunoglobulin molecule may be modified by structurally altering multiple toxicity associated domains in the constant region so that immunoglobulin-induced toxicity is inhibited. A discussion of such mutations are further found hereinafter.

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Regardless of the means, the underlying requirement for any structural alteration of the constant region is that immunoglobulin-induced toxicity is substantially reduced or inhibited. In one embodiment, immunoglobulin-induced toxicity is inhibited by structurally altering the constant region such that the molecule's ability to mediate a CDC response or ADCC response and/or activate the complement cascade is prevented or inhibited. Methods for determining whether the molecule is able to inhibit a CDC response are well known, e.g., one method involves a ⁵¹Cr-release test (H. Garrigues et al. Int. J. Cancer 29:511 (1982); I. Hellström et al. PNAS 82:1499 (1985)). Methods for determining whether the molecule is able to inhibit an ADCC response are well known (I. Hellström et al. PNAS 82:1499 (1985)). Methods for determining whether the molecule is able to activate a complement cascade are well known.

In another embodiment of the invention, the method comprises administering to the subject an Ig fusion protein having a structurally altered constant region. Structural alteration of the constant region may include deletion of the entire C region or portions thereof, e.g., alteration of the CH₂ domain so that the altered molecule no longer binds the Fc receptor or a complement component.

The invention further provides a method for inhibiting immunoglobulin-induced toxicity resulting from immunotherapy in a subject. The method comprises administering to the subject an antibody which has been modified so that at least a portion of the constant region has been structurally altered as discussed supra. In one embodiment, the antibody recognizes and binds Le^y. In another embodiment, the antibody recognizes and binds to Le^x.

In accordance with the practice of this invention, the antibody can be monoclonal antibody BR96 produced by the hybridoma having the identifying characteristics of HB 10036 as deposited with the ATCC. Alternatively, the antibody can be chimeric antibody ChiBR96 produced by the hybridoma having the identifying characteristics of HB 10460 as deposited with the ATCC. Further, the antibody can be a bispecific antibody with a binding specificity for two different antigens, one of the antigens being that with which the monoclonal antibody BR96 produced by the hybridoma having the identifying characteristics of HB 10036 as deposited with the ATCC binds.

Additionally, the present invention provides a method for inhibiting immunoglobulin-induced toxicity resulting from immunotherapy for a disease in a subject. The disease will vary with the antigen sought to be bound. Examples of diseases include but are not limited to immunological diseases, cancer, cardiovascular diseases, neurological diseases, dermatological diseases or kidney disease.

This method comprises the following steps. Step one provides selecting an antibody for a target. Generally, the target is associated with the disease and the antibody directed to the target is known. For example, the target can be the BR96 antigen and the antibody selected is BR96.

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Step two of this method provides structurally altering the constant region of the antibody so selected so that immunoglobulin induced toxicity is inhibited. Inactivation can include any of the means discussed above. For example, inactivation can be effected by structurally altering multiple toxicity associated domains in the CH₂ domain of the constant region of the Ig protein so selected.

Step three of this method provides administering the structurally altered antibody of step two to the subject under conditions that the structurally altered antibody recognizes and binds the target and that such binding directly or indirectly alleviates symptoms associated with the disease.

In accordance with the invention, in one embodiment step one provides selecting an Ig fusion protein for a target. Further, the method provides mutating the Ig fusion protein so selected by structurally altering the CH₂ domain of the constant region of the Ig protein by the same means discussed above.

The invention further provides methods to treat human carcinoma. For example, the immunoglobulin, antibody, or Ig fusion protein discussed above can be used in combination with standard or conventional treatment methods such as chemotherapy, radiation therapy or can be conjugated or linked to a therapeutic drug, or toxin, as well as to a lymphokine or a tumor-inhibitory growth factor, for delivery of the therapeutic agent to the site of the carcinoma.

Techniques for conjugating therapeutic agents to immunoglobulins are well known (see, e.g., Arnon et al., "Monoclonal Antibodies For Immunotargeting Of Drugs In Cancer Therapy", in Monoclonal Antibodies And Cancer Therapy, Reisfeld et al. (eds.), pp. 243-56 (Alan R. Liss, Inc. 1985); Hellström et al., "Antibodies For Drug Delivery", in Controlled Drug Delivery (2nd Ed.), Robinson et al. (eds.), pp. 623-53 (Marcel Dekker, Inc. 1987); Thorpe, "Antibody Carriers Of Cytotoxic Agents In

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Cancer Therapy: A Review", in Monoclonal Antibodies '84: Biological And Clinical Applications, Pinchera et al. (eds.), pp. 475-506 (1985); and Thorpe et al., "The Preparation And Cytotoxic Properties Of Antibody-Toxin Conjugates", Immunol. Rev., 62:119-58 (1982)).

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Alternatively, the structurally altered antibody or Ig fusion protein can be coupled to high-energy radiative agents, e.g., a radioisotope such as ¹³¹I; which, when localized at the tumor site, results in a killing of several cell diameters (see, e.g., Order, "Analysis, Results, And Future Prospective Of The Therapeutic Use Of Radiolabeled Antibody In Cancer Therapy", in Monoclonal Antibodies For Cancer Detection And Therapy, Baldwin et al. (eds.), pp. 303-16 (Academic Press 1985)). According to yet another embodiment, the structurally altered BR96 antibody can be conjugated to a second antibody to form an antibody heteroconjugate for the treatment of tumor cells as described by Segal in United States Patent 4,676,980.

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Still other therapeutic applications for the structurally altered antibody or Ig fusion protein of the invention include conjugation or linkage, e.g., by recombinant DNA techniques or protein chemical techniques, to an enzyme capable of converting a prodrug into a cytotoxic drug and the use of that antibody-enzyme conjugate in combination with the prodrug to convert the prodrug to a cytotoxic agent at the tumor site (see, e.g., Senter et al., "Anti-Tumor Effects Of Antibody-alkaline Phosphatase", Proc. Natl. Acad. Sci. USA, 85:4842-46 (1988); "Enhancement of the in vitro and in vivo Antitumor Activities of Phosphorylated Mitomycin C and Etoposide Derivatives by Monoclonal Antibody-Alkaline Phosphatase Conjugates", Cancer Research 49:5789-5792 (1989); and Senter, "Activation of Prodrugs by Antibody-Enzyme Conjugates: A New Approach to Cancer Therapy," FASEB J. 4:188-193 (1990)).

It is apparent therefore that the present invention encompasses pharmaceutical compositions including immunoglobulin molecules, antibodies, and Ig fusion proteins all having structurally altered CH₂ domains, and their use in methods for treating human carcinomas. For example, the invention includes pharmaceutical compositions for use in the treatment of human carcinomas comprising a pharmaceutically effective amount of a structurally altered BR96 and a pharmaceutically acceptable carrier.

The compositions may contain the structurally altered antibody or Ig fusion protein or antibody fragments, either unmodified, conjugated to a therapeutic agent (e.g., drug, toxin, enzyme or second antibody). The compositions may additionally include other antibodies or conjugates for treating carcinomas (e.g., an antibody cocktail).

The compositions of the invention can be administered using conventional modes of administration including, but not limited to, intrathecal, intravenous, intraperitoneal, oral, intralymphatic or administration directly into the tumor. Intravenous administration is preferred.

The composition of the invention can be in a variety of dosage forms which include, but are not limited to, liquid solutions or suspensions, tablets, pills, powders, suppositories, polymeric microcapsules or microvesicles, liposomes, and injectable or infusible solutions. The preferred form depends upon the mode of administration and the therapeutic application.

The compositions of the invention also preferably include conventional pharmaceutically acceptable carriers and adjuvants known in the art such as human serum albumin, ion exchangers, alumina, lecithin, buffer substances such as phosphates, glycine, sorbic acid, potassium sorbate, and salts or electrolytes such as protamine sulfate.

In accordance with the practice of the invention, the pharmaceutical carrier can be a lipid carrier. The lipid carrier can be a phospholipid. Further, the lipid carrier can be a fatty acid. Also, the lipid carrier can be a detergent. As used herein, a detergent is any substance that alters the surface tension of a liquid, generally lowering it.

In one example of the invention, the detergent can be a nonionic detergent. Examples of nonionic detergents include, but are not limited to, polysorbate 80 (also known as Tween 80 or (polyoxyethylenesorbitan monooleate), Brij, and Triton (for example Triton WR-1339 and Triton A-20).

Alternatively, the detergent can be an ionic detergent. An example of an ionic detergent includes, but is not limited to, alkyltrimethylammonium bromide.

Additionally, in accordance with the invention, the lipid carrier can be a liposome.

As used in this application, a "liposome" is any membrane bound vesicle which contains any molecules of the invention or combinations thereof.

The most effective mode of administration and dosage regimen for the compositions of this invention depends upon the severity and course of the disease, the patient's health and response to treatment and the judgment of the treating physician.

The interrelationship of dosages for animals of various sizes and species and humans based on mg/m² of surface area is described by Freireich, E.J., et al. Cancer Chemother., Rep. 50 (4): 219-244 (1966). Adjustments in the dosage regimen can be made to optimize the tumor cell growth inhibiting and killing response, e.g., doses can be divided and administered on a daily basis or the dose reduced proportionally depending upon the situation (e.g., several divided doses can be

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administered daily or proportionally reduced depending on the specific therapeutic situation).

THE MOLECULES OF THE INVENTION

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The present invention provides structurally altered BR96 or BR96 Ig fusion proteins. Structurally altered BR96 antibodies or Ig fusion proteins have the variable region of BR96 and a modified constant region. This modification provides structurally altered BR96 antibodies or Ig fusion proteins with the ability to inhibit immunoglobulin-induced toxicity.

Various embodiments of structurally altered BR96 or BR96 lg fusion proteins have been made.

- In one embodiment, designated cBR96-A, the entire CH₂ domain of cBR96 was deleted. CBR96-A is expressed by the plasmid having the sequence shown in SEQ. ID. NO. 10. cBR96 is expressed by a plasmid having the sequence in SEQ ID NO. 9.
- In another embodiment, designated hBR96-2A, the entire CH₂ domain of hBR96 was deleted. hBR96-A is expressed by the plasmid having the sequence shown in SEQ. ID. NO. 12. hBR96 is a mutant BR96 having the H1, H2, and H3 mutations described in PCT Application No. 95/305444, published March 6, 1996.
- In yet another embodiment, designated hBR96-2B, the leucine residue located at amino acid position 235 is mutated to alanine. Additionally, the glycine residue located at amino acid position 237 is mutated to alanine. The amino acid position numbering used is described in Kabat et al. Sequences of Proteins of Immunological Interest 5th Edition (1991) United States Department of Health and Human Services.

In a further embodiment, designated hBR96-2C, the glutamic acid residue at position 318 is mutated to serine; the lysine residue located at position 320 is mutated to serine; and the lysine residue located at position 322 is mutated to serine using standard protocols (Alexander R. Duncan and Greg Winter "The binding site for C1q on IgG" Nature 332:738 (1988)).

In another embodiment, designated hBR96-2D, the proline residue at position 331 is mutated to alanine (M-H. Tao et al., "Structural features of human immunoglobulin G that determine isotype-specific differences in complement activation" J. Exp. Med. 178:661-667 (1993); Y. Xu et al., "Residue at position 331 in the IgG1 and IgG4 domains contributes to their differential ability to bind and activate complement" J. Biol. Chem. 269:3469-3474 (1994)).

In an additional embodiment, designated hBR96-2E, the leucine residue at position 235 is mutated to alanine; the glycine residue located at position 237 is mutated to alanine; the glutamic acid residue located at position 318 is mutated to serine; the lysine residue located at position 320 is mutated to serine; and the lysine residue located at position 322 is mutated to serine (A. Morgan et al., "The N-terminal end of the CH₂ domain of chimeric human IgG1 anti-HLA-DR is necessary for C1q, Fc(gamma)RI and Fc(gamma)RIII binding" Immunol. 86:319-324 (1995)).

In yet a further embodiment, designated hBR96-2F, the leucine residue located at position 235 is mutated to alanine; the glycine residue located at position 237 is mutated to alanine; and the proline residue located at position 331 is mutated to alanine.

In yet another embodiment, designated hBR96-2G, the glutamic acid residue located at position 318 is mutated to serine; the lysine residue located at position 320 is

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mutated to serine; the lysine residue located at position 322 is mutated to serine; and the proline residue located at position 331 is mutated to alanine.

In another embodiment, designated hBR96-2H, the leucine residue located at position 235 is mutated to alanine; the glycine residue located at position 237 is mutated to alanine; the glutamic acid residue at position 318 is mutated to serine; the lysine residue located at position 320 is mutated to serine; the lysine residue located at position 322 is mutated to serine; and the proline residue located at position 331 is mutated to alanine.

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Depending on its form, a structurally altered BR96 antibody or fusion protein can be a monofunctional antibody, such as a monoclonal antibody, or bifunctional antibody, such as a bispecific antibody or a heteroantibody. The uses of structurally altered BR96, i.e., as a therapeutic or diagnostic agent, will determine the different forms of structurally altered BR96 which is made.

Several options exists for antibody expression. Immunoexpression libraries can be combined with transfectoma technology, i.e., the genes for the Fab molecules derived from the immunoglobulin gene expression library can be connected to the desired constant-domain exons. These recombinant genes can then be transfected and expressed in a transfectoma that would secrete an antibody molecule.

Once produced, the polypeptides of the invention can be modified, i.e., by amino acid modifications within the molecule, so as to produce derivative molecules. Such derivative molecules would retain the functional property of the polypeptide, namely, the molecule having such substitutions will still permit the binding of the polypeptide to the BR96 antigen or portions thereof.

It is a well-established principle of protein chemistry that certain amino acid

substitutions, entitled "conservative amino acid substitutions," can frequently be made in a protein without altering either the conformation or the function of the protein.

5 Amino acid substitutions include, but are not necessarily limited to, amino acid substitutions known in the art as "conservative".

Such changes include substituting any of isoleucine (I), valine (V), and leucine (L) for any other of these hydrophobic amino acids; aspartic acid (D) for glutamic acid (E) and vice versa; glutamine (Q) for asparagine (N) and vice versa; and serine (S) for threonine (T) and vice versa.

Other substitutions can also be considered conservative, depending on the environment of the particular amino acid and its role in the three-dimensional structure of the protein. For example, glycine (G) and alanine (A) can frequently be interchangeable, as can alanine and valine (V).

Methionine (M), which is relatively hydrophobic, can frequently be interchanged with leucine and isoleucine, and sometimes with valine. Lysine (K) and arginine (R) are frequently interchangeable in locations in which the significant feature of the amino acid residue is its charge and the differing pK's of these two amino acid residues are not significant. Still other changes can be considered "conservative" in particular environments.

In one embodiment of the present invention, the polypeptide is substantially pure, i.e., free of other amino acid residues which would inhibit or diminish binding of the polypeptide to its target and would inhibit or reduce gastrointestinal toxicity which are normally exhibited during or after antibody therapy.

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NUCLEIC ACID MOLECULES ENCODING THE PRESENT INVENTION

The nucleotide sequences and the amino acid sequences of the variable and constant regions of BR96 are known. The sequence for the immunoglobulin constant region is known and provided in Figure 18. Specific mutations in the constant region of the BR96 antibody were made. Nucleic acid molecules encoding the seven mutants described above (hBR96-2B through hBR96-2H) are as follows.

In hBR96-2B, alanine at amino acid positions 235 and 237 is encoded by codons 10 GCU, GCC, GCA, or GCG.

In hBR96-2C, serine at positions 318, 320, and 322 is encoded by UCU, UCC, UCA, or UGG.

15 In hBR96-2D, alanine at position 331 is encoded by codons GCU, GCC, GCA, or GCG.

In hBR96-2E, alanine at positions 235 and 237 is encoded by codons GCU, GCC, GCA, or GCG. Serine at positions 318, 320, and 322 is encoded by UCU, UCC, UCA, or UGG.

In hBR96-2F, alanine at positions 235, 237, and 331 is encoded by codons GCU, GCC, GCA, or GCG.

In hBR96-2G, serine at positions 318, 320, 322 is encoded by UCU, UCC, UCA, or UGG. Further, the alanine at position 331 is encoded by codons GCU, GCC, GCA, or GCG.

In hBR96-2H, alanine at positions 235, 237, and 331 is encoded by codons GCU,

GCC, GCA, or GCG. Additionally, serine at positions 318, 320, 322 is encoded by UCU, UCC, UCA, or UGG.

Any of the above can be deoxyribonucleic acid (DNA), e.g., complementary DNA (cDNA), or ribonucleic acid (RNA).

IMMUNOCONJUGATES

Immunoconjugates (having whole antibody or Ig fusion proteins) may be constructed using a wide variety of chemotherapeutic agents such as folic acid and 10 anthracyclines (Peterson et al., "Transport And Storage Of Anthracyclines In Experimental Systems And Human Leukemia", in Anthracycline Antibiotics In Cancer Therapy, Muggia et al. (Eds.), p. 132 (Martinus Nijhoff Publishers (1982); Smyth et al., "Specific Targeting of Chlorambucil to Tumors With the Use of Monoclonal Antibodies", J. Natl. Cancer Inst., 76:503-510 (1986)), including 15 doxorubicin (DOX) (Yang and Reisfeld "Doxorubicin Conjugated with a Monoclonal Antibody Directed to a Human Melanoma-Associated Proteoglycan Suppresses Growth of Established Tumor xenografts in Nude Mice PNAS (USA)" 85:1189-1193 (1988)), Daunomycin (Arnon and Sela "In Vitro and in vivo Efficacy of Conjugates of Daunomycin With Anti-Tumor Antibodies" Immunol. Rev., 65:5-20 27 (1982)), and morpholinodoxorubicin (Mueller et al., "Antibody Conjugates With Morpholinodoxorubicin and Acid-Cleavable Linkers", Bioconjugate Chem., 1:325-330 (1990)).

25 BR96 has been conjugated to doxorubicin and has been shown to be effective in therapy of certain cancers or carcinomas (Trail, P.A., Willner, D., Lasch, S.J., Henderson, A.J., Casazza, A.M., Firestone, R.A., Hellström, I., and Hellström, K.E. Cure of xenografted human carcinomas by BR96-doxorubicin immunoconjugates. Science, 261:212-215, 1993).

In accordance with the practice of the invention, structurally altered BR96 can be used in forms including unreduced IgG, reduced structurally altered IgG, and fusion proteins (PCT Application No. 95/305444, published March 6, 1996).

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Suitable therapeutic agents for use in making the immunoconjugate includes Pseudomonas exotoxin A (PE) in either the native PE or LysPE40 form. LysPE40 is a truncated form containing a genetically modified amino terminus that includes a lysine residue for conjugation purposes. Doxorubicin is also a suitable therapeutic agent.

Additional examples of therapeutic agents include, but are not limited to, antimetabolites, alkylating agents, anthracyclines, antibiotics, and anti-mitotic agents.

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Antimetabolites include methotrexate, 6-mercaptopurine, 6-thioguanine, cytarabine, 5-fluorouracil decarbazine.

Alkylating agents include mechlorethamine, thiotepa chlorambucil, melphalan,
carmustine (BSNU) and lomustine (CCNU), cyclothosphamide, busulfan,
dibromomannitol, streptozotocin, mitomycin C, and cis-dichlorodiamine platinum
(II) (DDP) cisplatin.

Anthracyclines include daunorubicin (formerly daunomycin) and doxorubicin (also referred to herein as adriamycin). Additional examples include mitozantrone and bisantrene.

Antibiotics include dactinomycin (formerly actinomycin), bleomycin, mithramycin, and anthramycin (AMC).

Antimitotic agents include vincristine and vinblastine (which are commonly referred to as vinca alkaloids).

Other cytotoxic agents include procarbazine, hydroxyurea, asparaginase, corticosteroids, mytotane (O,P'-(DDD)), interferons.

Further examples of cytotoxic agents include, but are not limited to, ricin, bryodin, gelonin, supporin, doxorubicin, taxol, cytochalasin B, gramicidin D, ethidium bromide, etoposide, tenoposide, colchicine, dihydroxy anthracin dione, 1-dehydrotestosterone, and glucocorticoid.

Clearly analogs and homologs of such therapeutic and cytotoxic agents are encompassed by the present invention. For example, the chemotherapuetic agent aminopterin has a correlative improved analog namely methotrexate.

Further, the improved analog of doxorubicin is an Fe-chelate. Also, the improved analog for 1-methylnitrosourea is lomustine. Further, the improved analog of vinblastine is vincristine. Also, the improved analog of mechlorethamine is cyclophosphamide.

METHODS FOR MAKING MOLECULES OF THE INVENTION

There are multiple approaches to making site specific mutations in the CH₂ domain of an immunoglobulin molecule. One approach entails PCR amplification of the CH₂ domain with the mutations followed by homologous recombination of the mutated CH₂ into the vector containing the desired immunoglobulin, e.g., hBR96-2. For example, hBR96-2B and hBR96-2D have been made by this method.

Another approach would be to introduce mutations by site-directed mutagenesis of single-stranded DNA. For example, vector pD17-hG1b, which contains only the constant region of IgG1 and not the V domain of hBR96, has the f1 origin of replication. This gives the vector the properties of a phagemid and site-directed mutagenesis experiments can be performed according to the methods of Kunkel, et al. (Kunkel, T.A., J.D. Roberts, and R.A. Zakour, 1987 Methods Enzymol. 154:367-383) as provided in the Bio-Rad Muta-Gene® phagemid *in vitro* mutagenesis kit, version 2. For example, hBR96-2B, -C, -D, -E, -F, -G, and -H were made by this method.

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In order that the invention described herein may be more fully understood, the following examples are set forth. It should be understood that these examples are for illustrative purposes only and are not to be construed as limiting the scope of this invention in any manner.

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EXAMPLE 1

The following standard ELISA protocol was used.

Materials: Immulon2 96 well plates and Genetic Systems Specimen Diluent
Concentrate (10x); antibody conjugate was Goat Anti Human Kappa-HRP Mouse
Adsorbed, Southern Biotech. at 1:10,000 in Genetic Systems Conjugate Diluent
(1x); Genetic Systems EIA Chromogen Reagent (TMB) (1:100); Genetic Systems
EIA Buffered Substrate (1x); primary antibody or antigen were AffiniPure F(ab')₂
 Fragment Goat Anti Human IgG Fc Fragment specific (Jackson Immuno Research),
Goat Anti Human Kappa-UNLB (Southern Biotechnology Associates), Le^y-HSA
(Alberta Research Council).

Methods: Dilute primary antibody or antigen to 1.0 μ g/ml in 0.05M Carb/Bicarb buffer. Add 100 μ l of the diluted solution per well in Immulon 2 plates. Seal plates and incubate O.N. at 4°C.

5 Block plates by flicking them and blotting on paper towels. Add 200μl/well of Genetic Systems, Specimen Diluent Concentrate (1x). Incubate at least 1 hour at room temperature and then dump the contents of the plates. Wash the plates 3x in saline/Tween. Blot to dry. Allow the plates to dry at R.T. (45 min. to 1 hour). Seal and store the plates at 4°C.

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Test samples as follows. Dilute samples and standards in Specimen Diluent at 1:10. Perform serial dilutions in separate round bottom plates. Transfer 100µl/well of final dilutions to antigen coated assay plates; then incubate O.N. at 4°C. Wash plates 3x with saline/Tween.

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For conjugation add 100 µl/well of antibody-HRP conjugate in Genetic Systems Conjugate Diluent (1x). Incubate plates at Room Temp. for 60 min. Wash plates 3x in saline/Tween.

20 Add 100 µl/well of Genetic Systems EIA Chromogen Reagent (TMB) 1:100 in EIA Buffered Substrate (1x). Incubate at R.T. for 15 min. and stop with 1N H₂SO₄ 100 µl/well. Read plate at 450/630nm in EIA plate reader.

EXAMPLE 2

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Construction of CH₂ deleted BR96 molecules

Strategy for Deleting CH₂ Domains: To construct CH₂ deleted BR96 molecules, the hinge, CH₂ and CH₃ domains were removed from chimeric BR96 and humanized

BR9696-2 IgG1 molecules by an Eco47-III restriction digestion in non-coding regions. The hinge and CH₃ domains were amplified by polymerase chain reaction (PCR) from a human IgG1 (pNγ1.14) molecule lacking the CH₂ domain. Two oligonucleotides (Sense 49mer, Antisense 50mer) homologous to the sequences of IgG1 constant region at both sides preserving E.co47-III sites were synthesized. The amplified hinge and CH₃ domain PCR fragments were added into Eco47-III sites on BR96 IgG1 molecules by in vivo homologous recombination (P. Bubeck et al., Nucleic Acid Research (1993) 21:3601-3602). The new BR96 IgG1 molecules were verified by restriction mapping and sequencing.

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A sewing PCR strategy was used for the construction of CH₂ deleted human IgG1 (pNγ1.14) (Robert M. Horton, et al. (1990) Biotech 8 (5)P, 528).

The CH₁ domain was amplified as a 580 bp fragment with a sense oligonucleotide

(5' TGG CAC CGA AAG CTT TCT GGG GCA GGC CAG GCC TGA 3') (primer A) and an antisense oligonucleotide (5' TCC GAG CAT GTT GGT ACC CAC GTG GTG GTC GAC GCT GAG CCT GGC TTC GAG CAG ACA 3') (primer B) from a linearized human IgG1 constant region vector (pNγ1.7). The PCR fragment extends from the 5' end of the Hind-III site (in bold) through the Cel-II, Sal-I, Dra-III, Kpn-I, 6 bp nucleotide spacer and Mro-I sites (in bold) at the 3' end of the CH₁ domain.

The CH₃ domain was then partially amplified (to the Xba-I site) with a sense primer (5' GTC GAC CAC GTG GGT ACC AAC ATG TCC GGA GCC ACA

25 TGG ACA GAG GCC GGC T 3') (primer C) and an antisense primer (5' CTG GTT CTT GTT CAT CTC CTC TCT AGA TGG 3') (primer D) from a linearized human IgG1 constant region vector (pNγ1.7). A PCR fragment (about 150 bp) with Sal-I, Dra-III, Kpn-I, 6 nucleotide spacer and Mro-I sites (in bold) on its 5' end, extends only through the Xba-1 site (in bold) within the CH₃ domain.

The CH₁ and CH₃ partial PCR fragments were combined in a PCR without any primer. The reaction was run through two full cycles of denaturation and reannealing to allow the fragments to combine at the homologous region at the 3' ends. Primers A and D (described above) were added to the reaction and the PCR cycle was completed. The polymerase extends the DNA with primer A and primer D, yielding a full-length (660 bp) PCR fragment. The newly extended PCR fragment is arranged from the 5' end to the 3' end in the following order: Hind-III - CH₁ - Cel-II - Sal-I - Dra-III - Kpn-I - 6 bp spacer - Mro-I - CH₃ partial - Xba-1.

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The combined PCR fragment, with the CH₁ and partial CH₃ domains, was then cloned by a blunt end ligation into a Sma-I site on a pEMBL18 vector and the sequence was confirmed by dideoxy sequencing (Sanger et al. (1977) PNAS (USA) 74:5463-5466).

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To transfer the CH₁ and partial CH₃ into a mammalian expression vector, both the pEMBL18 and pNy1.7 vectors were digested with Hind-III and Xba-I. The Hind-III and Xba-I fragment was ligated into the same sites on a linearized pNy1.7 vector. The new construct, with CH₁ and a full CH₃ domain, was designated the pNy1.10 vector.

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The hinge fragment was amplified from a Hind-III digested pNγ1.7 vector with the primers designed to flank the hinge exon with a Sal-I and a Dra-III cloning site at each end. These sites also exist between the CH₁ and CH₃ domains of the pNγ1.10 construct. The sense oligonucleotide (5' ACC ATG GTC GAC CTC AGA CCT GCC AAG AGC CAT ATC 3') with a 6 bp spacer and a Sal-I cloning site (in bold) and the antisense oligonucleotide (5' CAT GGT CAC GTG GTG TGT CCC TGG ATG CAG GCT ACT CTA G 3') with a 6 bp spacer and a Dra-III cloning site (in bold) were used for the amplication of the hinge fragment (250 bp).

The hinge region PCR fragment was cloned into a Sma-I site on pEMBL18 by blunt end ligation. Both the pEMBL18 with the hinge domain and the pNy1.10 with the CH₂ and CH₃ domains were digested with Sal-1 and Dra-III. The digested hinge fragment was cloned into the Sal-1 and Dra-III linearized sites on the pNy1.10 vector. The new construct, now carrying the CH₁, hinge and CH₃ domains, was designated pNy1.11.

To make the final CH_2 deleted human IgG1 construct, both the pN γ 1.11 construct and pN γ 1.11 vector were digested with BamH1 and HindIII. A fragment containing the CH_1 , hinge and CH_3 domains was cloned into the linearized pN γ 1.11 vector. The new constant region IgG1 construct lacks the CH_2 domain and is designated pN γ 1.14 (Figure 11).

15 For digestion of BR96 IgG1 with Eco47-III, a restriction fragment with hinge, CH₂ and CH₃ domains was identified on the constant region sequence of BR96 IgG1 vector in both chimeric and humanized molecules. The 5' end of this fragment lies inside the intron between CH₁ and hinge and the 3' end is located inside the CH₃ intron of the BR96 IgG1 molecule. The hinge, CH₂ and CH₃ domains (1.368 kb fragment) were removed from BR96 IgG1 molecules by Eco47-III restriction digestion. The Eco47-III is a blunt end cutter. The BR96 IgG1 DNA digested with this enzyme does not require any pretreatment before cloning. Figure 12 is a diagrammatic representation of the pD17-hBR96-2 vector showing the Eco47-III sites used in cloning.

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The CH₂ deleted BR96 IgG1 was then constructed as follows. The hinge and CH₃ domains were amplified from a CH₂ deleted L6 IgG1 (pN γ 1.14) construct with a sense oligonucleotide (5'

CAGGGAGGGAGGTGTCTGCTGGAAGCCAGGCTCAGCGCTGACCTCAG

A 3') homologous to the constant region sequence of IgG1 at the 5' end of the Eco47-III site (in bold) and an antisense oligonucleotide (5'GGAAAGAACCATCACAGTCTCGCAGGGG CCCAGGGCAGCGCTGGGTGCTT 3') homologous to the constant region sequence of IgG1 at the 3' end of the Eco47-III site (in bold). The Eco47-III site at the 3' end of the pNγ1.14 construct is modified in the cloning process. The Eco47-III site is thus introduced into an antisense primer and used in amplification of the hinge and CH₃ domains.

- The pD17-BR96 IgG1 vector was digested with Eco47-III and the hinge, CH₂ and CH₃ domains were removed. The linearized pD17-BR96 IgG1 vector was mixed with equimolar amounts of hinge and CH₃ PCR fragments. Cotransformation of the PCR fragment with linearized DNA into E.coli DH5a competent cells resulted in a recombinant molecule, mediated by homologous recombination in bacteria. This construct lacks the CH₂ domain of BR96 IgG1 molecules, and is designated pD17-BR96-dCH2 (Figure 13).
 - 1.9 grams of CH₂-deleted chimeric BR96 was obtained as raw material from 89L of culture supernatant.

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EXAMPLE 3

Toxicity, localization and clearance of CH₂-deleted chimeric BR96 was tested in vivo as follows.

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Three dogs received 400 mg/m² of cBR96-A, the CH₂ deletion mutant of chimeric BR96, and two received chimeric BR96. Both molecules had been mildly reduced and alkylated. This is required to prevent dimerization of the deletion mutant into a tetravalent form. Both control dogs experienced the typical GI toxicity and none of

the three receiving the mutant displayed any toxicity. The control dogs and two of the test dogs were sacrificed at 1 hr to obtain duodenal tissue to measure antibody localization. Both control dogs had grossly visible GI pathology, and the test dogs had normal appearing GI tissue. The third dog has continued to show no signs of toxicity.

Results: A significant amount of localization of the CH₂ deleted cBR96 (cBR96-A) occurred to the GI tract in dogs treated with 400 mg/m², although the intact chiBR96 localized slightly better. The levels of localization indicate that roughly equivalent amounts of intact and CH₂ deleted cBR96 was delivered to the GI tract in these dogs.

Table 5. Localization of cBR96 to GI tissue.

Group	Animal	Specific	mean
		Localization	
	#271	155	
cBR96			135
	#272	114	
	#273	126	
cBR96-A			89
	#274	52	

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Using the mean level of specific localization, an amount of cBR96-A equivalent to at least 66% of the amount of cBR96 was delivered to the target organ of toxicity, the duodenum. Based on the dose ranging done with cBR96 in dogs (some clinical signs of toxicity seen at doses of 10 mg/m²), even if this difference is real, it could

not explain the difference between significant toxicity and no toxicity, evaluation to date indicated that dogs treated with cBR96-A had no toxicity, pending microscopic histopathologic examination. This evaluation was based on analysis of 2 frozen blocks per dog and 2 sections per block. Replicates were quite good. We also ran historical frozen tissues from dogs treated with native cBR96 or F(ab)2/BR96 and the levels of localization for those tissues were 110 and 0, respectively, consistent with our previous data.

Assuming that there is no toxicity at marginally higher (2X) doses of cBR96-A, these data indicate that the CH₂ domain is associated with the induction of acute gastroenteropathy, and that the removal of this domain prevents the induction of gastroenteropathy mediated by BR96.

This study confirms the results showing that F(ab')2 is not toxic in the dog model
and that the toxicity is mediated by the constant region. The CH₂ deletion mutant is
a candidate for targeting agents clinically. Because of the very long half-life of
chimeric BR96, some decrease in the mutant's half-life should be acceptable.

Figure 1 shows the measurement of the clearance of the cBR96-A in high Le^Y

expressing dogs. The study used chimeric versus constant region mutant of cBR96
2.

CBR96-2 did clear faster than the chimeric BR96. The localization of cBR96-A to the gastrointestinal epithelium is not significantly affected by this more rapid clearance. More than enough of the cBR96-A localized to have caused toxicity.

Discussion: The constant region of chimeric IgG is responsible for the GI toxicity seen in clinical trials, e.g. with chiBR96-dox. The GI toxicity seen in the dog model is very similar to the clinical toxicity. Both in man and dog, administration of the

unconjugated antibody mediates an acute GI toxicity characterized by rapid onset of vomiting, often with blood.

In man the bleeding is limited to the fundus of the stomach, causing erosion of the superficial mucosa of the stomach. Although the pathology of the wound is limited and resolves, the extreme nature of the nausea and vomiting, unrelieved by antiemetics, defines it as the dose-limiting toxicity.

This toxicity is mediated in man and dog by the antibody molecule alone. At higher doses of the antibody-dox conjugate, additional toxicity is seen in the dog model, probably due to doxorubicin. Although the intact IgG of BR96 causes toxicity in dog and man, the F(ab')2 molecule (divalent and lacking only in the constant region) is not toxic in dogs. This finding has motivated our attempts at high levels, and improves the affinity and specificity of BR96 for tumor antigen.

The CH₂ domain is known to mediate complement and FcR binding. It was not known that structural alteration of the CH₂ domain would result in immunoglobulin-induced toxicity inhibition.

20 Toxicology study of hBR96-2B

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The toxicology study of hBR96-2B in high Lewis Y expressor dogs (n=2) showed that a dose of 400 mg/m² did not cause hematemesis nor bloody stools, in contrast to BR96 which consistently causes one or both signs. A dog sacrificed at 24 hrs had normal gross appearance of the GI tract, again in marked contrast to chimeric BR96 which causes hemorrhagic lesions and mucosal erosions.

EXAMPLE 4

The polymerase chain reaction (PCR) is a widely used and versatile technique for the amplification and subsequent modification of immunoglobulin genes. The rapidity and accuracy with which antibody genes can be modified in vitro has 5 produced an assortment of novel antibody genes can be modified in vitro has produced an assortment of novel antibodies. For example, PCR methods have been used for engineering antibodies with increased affinity to antigen, for "humanizing" antibodies, and for modulating effector function (Marks, J.D., A.D. Griffiths, M. Malmqvist, T. Clackson, J.M. Bye and G. Winter. 1992. Bypassing immunization: 10 high affinity human antibodies by chain shuffling. Bio/Technology 10:779-783; Rosok, M.J., D.E. Yelton, L.J. Harris, J. Bajorath, K.-E. Hellstrom, I. Hellstrom, G.A. Cruz, K. Kristensson, H. Lin, W.D. Huse and S.M. Glaser. 1996. A combinatorial library strategy for the rapid humanization of anticarcinoma BR96 Fab. J. Biol. Chem. 271:22611-22618; Morgan, A.N., D. Jones, A.M. Nesbitt, L. 15 Chaplin, M.W. Bodmer and S. Emtage. 1995. The N-terminal end of the CH2 domain of chimeric human IgG1 anti-HLA-DR is necessary for Clq, FcyRI and FcyRIII binding. Immunology. 86:319-324).

As part of a more comprehensive study, we desired to introduce various site specific mutations in the CH₂ constant domain of human IgG₁. Six specific amino acid residues distributed throughout the CH2 domain previously identified to play a role in immune effector function were marked as targets for mutagenesis (Morgan, A.N., D. Jones, A.M. Nesbitt, L. Chaplin, M.W. Bodmer and S. Emtage. 1995. The N-terminal end of the CH2 domain of chimeric human IgG1 anti-HLA-DR is necessary for Clq, FcγRI and FcγRIII binding. Immunology. 86:319-324; Duncan, A.R. and G. Winter. 1988. The binding site for Clq on IgG. Nature 332:738-740; Tao, M.-H., R.I.F. Smith and S.L. Morrison. 1993. Structural features of human immunoglobulin G that determine isotype-specific differences in complement

activation. J.Exp.Med. 178:661-667). five of the six residues were grouped into two clusters-one cluster consisting of two residues, two amino acids apart (Location 1, or L1); and a second cluster consisting of three residues spanning a sequence of five amino acids (L2). The remaining amino acid position (L3) made for the total of six residues. We were interested in constructing a panel of mutant CH₂ domain IgGs consisting of each L mutation by itself as well as in combination with other L mutants (e.g., L1; L1; and L2; L1, L2 and L3; etc.).

Various in vitro methods have been described where PCR is used to simultaneously introduce distally located site-specific mutations within a gene sequence (Ho, S.N., H.D. Hunt, R.M. Horton, J.K. Pullen and L.R. Pease. 1989. Site-directed mutagenesis by overlap extension. Gene 77:51-59; Ge, L. and P. Rudolpf. 1996. Simultaneous introduction of multiple mutations using overlap extention PCR. BioTechniques 22:28-30). Alternatively, an in vivo procedure termed recombination PCR (RPCR) has also successfully been used for rapidly and efficiently generating 15 distally located site-specific mutations (Jones, D.H. and S.C. Winistorfer. 1993. Use of polymerase chain reaction for making recombinant constructs. p.241-250. In B.A. White (Ed.), Methods in Molecular Biology, Vol. 15. Humana Press Inc., Totowa, NJ, Jones, D.H. And B.H. Howard. 1991. A rapid method for recombination and site-specific mutagenesis by placing homologous ends on DNA 20 using polymerase chain reaction. BioTechniques 10:62-66). RPCR uses E. Coli's recombination machinery to generate intact circular recombinant plasmids from a transfected mixture of linear PCR-generated product and linearized vector. In vivo recombination is mediated through the joining of nucleotide sequences designed into the 5' ends of both PCR primers that are homologous to DNA sequences encoded by 25 the vector. In this report we describe an extension of the RPCR procedure for simultaneously introducing complex combinations of mutations into an antibody CH₂ domain.

Humanized BR96 variable region heavy and light chain genes, previously cloned and co-expressed as an assembled active Fab fragment in an M13 phage expression vector, provided the starting material (Rosok, M.J., D.E. Yelton, L.J. Harris, J. Bajorath, K.-E. Hellstrom, I. Hellstrom, G.A. Cruz, K. Kristensson, H. Lin, W.D. Huse and S.M. Glaser. 1996. A combinatorial library strategy for the rapid 5 humanization of anticarcinoma BR96 Fab. J. Biol. Chem. 271:22611-22618). The heavy and light chain V genes were amplified by PCR from a single-stranded M13 DNA template and subcloned by in vivo recombination (Jones, D.H. And B.H. Howard. 1991. A rapid method for recombination and site-specific mutagenesis by 10 placing homologous ends on DNA using polymerase chain reaction. BioTechniques 10:62-66) into vectors pD17-hGla and pD16-hCκ, to form pBR96-hGla and pBR96-hCk respectively. pD17-hG1a and pD16-hCk are eukaryotic immunoglobulin expression vectors derived from pcDNA3 (Invitrogen, San Diego, CA). The plasmid pBR96-hG1a was further modified by site-directed mutagenesis to introduce two Eco47-III restriction sites flanking the immunoglobulin hinge-CH₂-15 CH₃ domains using standard procedures. The recipient vector was then prepared by digesting pBR96-hG1a with Eco47-III, isolating the vector backbone by agarose gel electrophoresis followed by extracting the vector DNA from the excised gel slice using the Qiagen Gel Extraction kit (Qiagen, Chatsworth, CA).

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The strategy for introducing multiple mutations within the immunoglobulin CH₂ gene, shown in Figure 24, relies on the *in vivo* homologous recombination of several independently amplified PCR products with each other as well as with the pBR96-hG1a vector DNA. For introducing mutations at two distal locations two PCR products are synthesized (Figure 24B). One end of each PCR product is for recombining with an homologous end of the linear vector, and the other end, encoding the mutation(s) of interest, is for recombining with the neighboring PCR product. As shown in Figure 24B, additional distally-located mutations can be introduced into a target sequence by increasing the number of PCR products

proportionately. The recombination of neighboring PCR products always occurs across the regions containing the desired mutations, therefore the oligonucleotide primers encoding these ends (e.g., A1, A2) contain complementary mutant residues. The mutagenic PCR primers contain at least 15 nucleotides of wild-type sequence flanking each side of the mutant residues for either priming the polymerization reaction or mediating recombination. Two 49-nucleotide long PCR sense and antisense primers (Rs and Ra) contain sequences for recombining with the end regions of the Eco47-III digested pBR96-hG1a vector.

Each L mutation was amplified in a separate PCR reaction. The reaction conditions 10 were 250 ng intact pBR96-hG1a DNA template, 10 ul of 1X Pfu buffer (Stratagene, Inc. San Diego, CA), 10 nmol dNTPs, 200ng each of the appropriate PCR primers, 10% dimethysulfoxide (ATCC, Rockville, MD) and 2.5 units cloned Pfu DNA polymerase in a 100ul reaction volume. Samples were first denatured at 95° C for 5 min, cooled to 45°C for 5 min, and extended at 72°C for 1 min followed by 25 15 cycles of denaturation at 94°C for 45 sec, annealing at 45°C for 45 sec, extension at 72°C for 1 min/kb, followed by a final extension at 72°C for 7 min in a Perkin-Elmer DNA Thermal Cycler (Norwalk, CT). The amplified products were purified from a 1% agarose gel, extracted with Qiagen Gel Extraction kit and the recovered DNA quantitated. 50 ng of each PCR product was mixed with 25 ng of the Eco47-20 III digested pBR96-hG1a vector, transfected into Max competent E. coli DH5a according to the manufacturer's procedure (GIBCO BRL/Life Technologies, Gaithersburg, MD), and the entire transfection reaction plated onto selective LB agar plates containing 100 ug/ml ampicillin.

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The results of several cloning experiments are summarized in the Table that follows. Typically the transformations produced from 80 to 200 bacterial colonies. Individual colonies were selected and grown overnight in 2 ml liquid cultures for isolation of miniprep plasmid DNA (Qiagen) and analysis by Eco47-III restriction

endonuclease mapping. Among 24 independent transformants analyzed from triple homologous recombination events (two PCR products plus vector) 11 clones contained the predicted 1.4 kpb DNA insert.

Figure 25 shows a sample diagnostic restriction analysis of DNA prepared from clones derived from quadruple homologous recombination events (three PCR products plus vector). Additional sampling of clones resulting from quadruple recombination yielded a cloning efficiency of 29% (7 clones containing inserts/24 clones sampled). At this point, due to the small sampling sizes, we do not know whether the differences in the cloning efficiencies observed between the triple and quadruple recombination events are meaningful.

To evaluate the expression of Le⁷ -binding activity of the CH₂ mutant IgGs, miniprep DNAs from 6 clones derived from the triple recombination reaction and 6 clones derived from the quadruple recombination reaction exhibiting the predicted 15 diagnostic Eco47-III restriction patterns were isolated, mixed with pBR96- hCk DNA and used to co-transfect COS7 cells. 48 hour spent supernatants from 3 ml cultures were assayed for total IgG production and for Ley binding activity by enzyme-linked immunosorbent assay (EIA) as described (Yelton, D.E., M.J. Rosok, G.A. Cruz, W.L. Cosand, J. Bajorath, I. Hellstom, K.-E. Hellstorm, W.D. Huse and 20 S.M. Glaser. 1995. Affinity maturation of the BR96 anti-carcinoma antibody by codon-based mutagenesis. J.Immunol. 155:1994-2004). All twelve cultures were found to secrete approximately 2-3 ug/ml Le^γ -reactive IgG. The spectrum of Le^γ binding activities were all similar to that of native humanized BR96 IgG indicating that the homologously recombined antibodies did not acquire any gross mutations 25 that could affect antigen binding. To confirm that the desired CH₂ mutations had been incorporated, and to evaluate the recombined genes for misincorporated nucleotides, four of the clones producing functional antibody were sequenced using Sequenase Version 2 DNA Sequencing Kit (United States Biochemical). One clone

was found to contain a single nucleotide change within the forward PCR primer used for mediating recombination with vector DNA. We are uncertain whether this error occurred during chemical synthesis of the oligonucleotide primer or is a result of misincorporation during the PCR reaction, despite the fact that we used a thermostable polymerase with proofreading activity.

A RPCR procedure for homologously recombining up to three separate PCR-generated mutated antibody sequence products into a eukaryotic expression vector for the rapid construction of engineered IgG molecules is described herein. The advantage of this approach is the ability to simultaneously introduce multiple distally-located mutations with PCR products synthesized by a single round of PCR. Recombinant DNAs are produced with a reasonably high cloning efficiency and fidelity of correct nucleotide sequences. The ability to efficiently rejoin several distinct PCR products should permit combinatorial strategies for constructing complexly mutated protein domains as well as broadening the number and location of desired mutations.

Analysis of transformants generated by multiple-fragment RPCR.

Mutant IgGs	PCR	HR ^a events	Colonies	Cloning
Constructed	Fragments in		Analyzed	Efficiency ^b
	reaction			
2	2	triple	24	45%
2	3	quadruple	24	33%

^aHR-homologous recombination

^bCloning efficiency (number of clones containing 1.4kbp insert/total number of colonies

EXAMPLE 5

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This example provides two methods for introducing site specific mutations into the CH2 domain of human IgG1 constant region containing vectors.

One method involves PCR amplification of a segment or segments of the constant region, wherein mutations are introduced using appropriately constructed oligonucleotides. The vector receiving the fragment(s) is digested with a restriction enzyme to linearize the vector. PCR amplification primers are designed so that the 5' ends of the PCR fragments can hybridize to the DNA sequence of the vectors. If more than one PCR fragment is amplified, then common sequences to the two fragments are introduced by oligonucleotides. Bacteria are transfected with the PCR fragments and with the digested vector. The fragments and vector can recombine by homologous recombination using the bacteria's recombination machinery. Bacterial colonies are selected and the DNA is analyzed by size and restriction map as a preliminary determination that the vector and fragment(s) recombined correctly. Correct insertion of fragments with the mutations is confirmed by dideoxynucleotide sequence analysis. DNA is then introduced into mammalian cells as described for the CH2 deleted antibody, and the expressed antibody analyzed for binding and functional activity.

By way of example, mutations Leu to Ala at residue 235 in CH2 and Gly to Ala at residue 237 were introduced by the procedure disclosed in Example 4. The heavy chain vector used for this procedure was pD17-hG1a, similar to pD17-BR96 vector described herein except that humanized V regions (Rosok, M.J., D.E. Yelton, L.J. Harris, J. Bajorath, K-E. Hellstrom, I, Hellstrom, G.A. Cruz, K. Kristensson, H. Lin, W.D. Huse, and S.M. Glaser, 1996. J. Biol. Chem 271 37:22611-22618) with three affinity mutations (H1, H2, and H3 mutations) were substituted.

pBR96-hG1a contains two Eco47-III restriction sites flanking the Ig hinge-CH2-CH3 domains. The recipient vector was prepared by (1) digesting pBR96-hG1a with *Eco*47-III, (2) isolating the vector by agarose gel electrophoresis, and (3) extracting the vector DNA from the excised gel slice using the Qiagen Gel Extraction kit (Qiagen, Chatsworth, CA). To introduce mutations at a single location, such as for positions 235 and 237, two PCR products were synthesized.

10 herein as hBR96-2F) with mutations at 235, 237, 331, requires 3 PCR products. The recombination of neighboring PCR products occurs across the regions containing the desired mutations, therefore the oligonucleotide primers encoding these ends contain complementary mutant residues. The mutagenic PCR primers contain at least 15 nucleotides of wild-type sequence flanking each side of the mutant residues for either priming the polymerization reaction or mediating recombination. Two 49-nucleotide long PCR sense and anti-sense primers containing sequences for recombining with the end regions of the *Eco*47-III digested pBR96-hG1a vector.

PCR amplification used 250 ng intact pBR96-hG1a DNA template, 10 μl of 10X *Pfu* buffer (Stratagene, Inc., San Diego, CA), 10 nmol dNTPs, 200 ng each of the appropriate PCR primers, 10% dimethylsulfoxide (ATCC, Rockville, MD) and 2.5 units cloned *Pfu* DNA polymerase (Stratagen, Inc. San Diego, CA) in 100 μl reaction. Samples were denatured at 95°C for 5 min, annealed at 45°C for 5 min, and extended at 72°C for 1 min followed by 25 cycles of denaturation at 94°C for 45 sec, annealing at 45°C for 45 sec, extension at 72°C for 1 min/kb, and a final extension at 72°C for 7 min. The amplified products were purified from a 1% agarose gel, extracted with the Qiagen Gel Extraction kit and quantitated. 50 mg of each PCR product was mixed with 25 ng of the Eco47-III digested pBR96-hG1a vector and transfected in E.coli MAX Efficiency DH5αTM according to the

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manufacturer's instructions (GIBCO BRL/Life Technologies, Gaithersburg, MD). The entire transfection reaction was plated onto LB agar plated containing 100 µg/ml ampicillin.

- Bacterial colonies were selected and grown overnight at 37° C in 2 ml liquid 5 cultures. DNA was isolated and analyzed by Eco47-III restriction endonuclease mapping. Clones with the correct size insert were sequenced (Sequenase Version 2, U.S. Biochemical Corp., Cleveland, OH).
- The second method for introducing site specific mutations into the CH₂ domain of 10 human IgG1 involved the method of Kunkel (1987 Methods Enzymology, supra). For this procedure pD17-hG1b DNA with the F1 origin of replication was introduced into electrocompetent E. coli CJ236 dut-ung- (Bio-Rad Laboratories, Hercules, CA) by electroporation according to manufacturer's instructions. PD17hGlb is a vector having a constant region but no variable region. The F1 ori site 15 allows treatment of this vector as a phagemid.

Bacteria containing the plasmid were selected by ampicillin resistance. Single stranded uridinylated DNA was prepared using the Muta-Gene Phagemid In Vitro Mutagenesis Version 2 protocol (Bio-Rad). Mutations were introduced by sitedirected mutagenesis with the appropriate antisense oligonucleotide. For molecules with mutations at more than one location, mutations were introduced by either of the two methods discussed above. One method would be to (1) prepare one mutant, for example, mutant 2C (also referred to herein as BR96-2C) with the mutations at residues 318, 320, 322, (2) isolate ssDNA, and (3) introduce a second mutation set 25 with the appropriate anti-sense oligonucleotide. The second method would be to anneal two antisense oligonucleotides with the same uridinylated ssDNA and screen for mutants with both sets of changes. Mutant 2H (hBR96-2H) was also prepared by a combination of thse methods.

The V region of humanized BR96-2 heavy chain was introduced by the homologous recombination method described above in pD17-hJm14.H1. The pD17-hJm14.H1 plasmid contains the BR96 humanized variable region with the H1/H2/H3 mutations and the plasmid was used to transfect mutant sequences into mammalian cells. The pD17G1b vector containing the Fc mutation(s) was digested with NheI for 3 hr at 37° C and the DNA isolated by methods described above. Insertion of the V region into the vector was determined by size and restriction enzyme mapping and confirmed by sequence analysis.

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Transient expression of whole antibodies was performed by transfection of COS cells. For production of antibody, stable transfections of CHO cells were performed (see description of deleted CH2 mutant). All mutants were purified from CHO culture supernatants by protein A chromatography.

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The oligonucleotide primers homologous to the vector and used to introduce the constant regions mutations were as follows:

Oligonucleotides homologous to vector sequences:

Sens(sense)CH2 E47-3-5: CAG GGA GGG AGG GTG TCT GCT GGA AGC

20 CAG GCT CAG CGC TGA CCT CAGA

D CH2 E47-3 A (antisense): GGA AAG AAC CAT CAC AGT CTC GCA GGG GCC CAG GGC AGC GCT GGG TGC TT

Oligonucleotides to mutate Leu235 to Ala and Gly237 to Ala (underlined sequences show sites of mutation):

Antisense CH2 L235-G237/aa: GAA GAG GAA GAC TGA CGG TGC CCC

CGC GAG TTC AGG TGC TGA GG

SensCH2 L235-G237/AA: CCT CAG CAC CTG AAC TCG CGG GGG CAC

CGT CAG TCT TCC TCT TC

Oligonucleotides to mutate Glu318, Lys320, Lys322 to Ser

Antis(antisense)CH2 EKK/SSS-2: CTG GGA GGG CTT TGT TGG AGA CCG

AGC ACG ACT ACG ACT TGC CAT TCA GCC

5 Oligonucleotides to mutate Pro331 to Ala:

Antis CH2 P331/A/3: GAT GGT TTT CTC GAT GGC GGC TGG GAG GGC Sense CH2 P33/A: GCC CTC CCA GCC GCC ATC GAG AAA ACC ATC Alternative antisense oligo to introduce Ala at 331 by site-directed mutation: CH2P331A: GAT GGT TTT CTC GAT AGC GGC TGG GAG GGC TTT G

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Oligonucleotides to mutate Glu318 to Ser, Lys320 to Ser, Lys322 to Ser, and Pro331 to Ala:

Antis CH2 EKKP/SSA-6: GAT GGT TTT CTC GAT GGC GGC TGG GAG GGC TTT GTT GGA GAC CGA GCA CGA GTA CGA CTT GCC ATT CAG CCA GTC CTG GTG

Sense CH2 EKKP/SSA-6: CAC CAG GAC TGG CTG AAT GGC AAG TCG
TAC TCG TGC TCG GTC TCC AAC AAA GCC CTC CCA GCC GCC ATC
GAG AAA ACC ATC

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In vitro Assays of the Mutants

Results of the CDC demonstrate that mutant hBR96-2B has approximately 10 fold less activity than the control hBR96-1 (two affinity mutations, one in H2 and one in H3, refer to previous patent (Figure 20)). The mutants that have the least ability to kill cells in the presence of complement is hBR96-2C with the triple mutations at positions 318, 320, and 322 and the hBR96-2H mutant (least cytotoxic antibodies in the panel) which contains all six mutations at the three different locations. ADCC activity was most affected by the CH2 deleted hBR96-2 molecule (Figure 21).

hBR96-2B and -2H lost between 100 and 1000 fold activity to kill in the presence of effector cells. In the ADCC assay the hBR96-2B molecule also lost approximately 10 fold activity (Figure 21).

Figures 26-28 provide the amino acid sequences for the heavy chain variable region for both chimeric and humanized BR96 having the H1, H2, and H3 mutations. The amino acid sequence for the light chain variable region is known and methods for generating it are found in PCT Application No. 95/305444. Additionally provided is the amino acid sequence for the IgG1 constant region. Mutations in the constant region are marked.

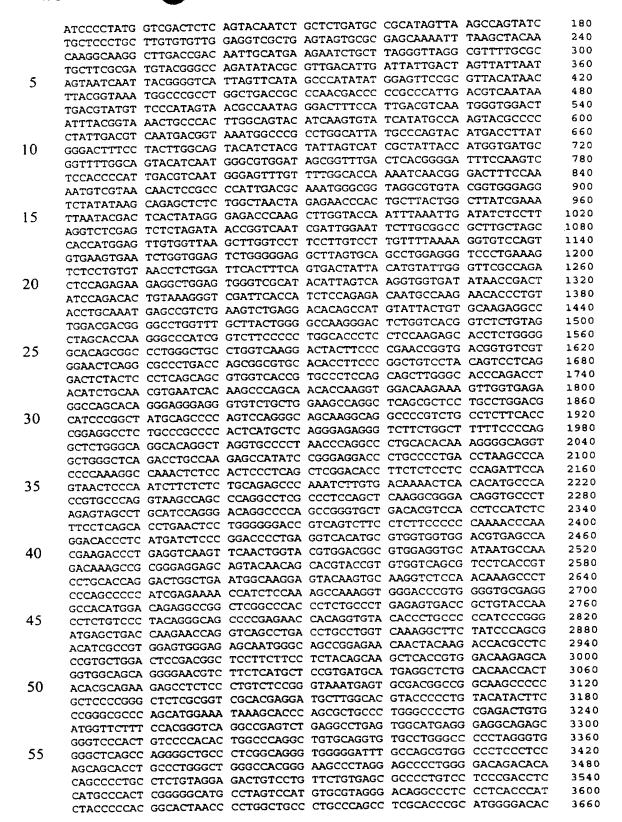
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	(iii) NUMBER OF SEQUENCES: 13
15	(iv) CORRESPONDENCE ADDRESS:(A) ADDRESSEE: Merchant & Gould(B) STREET: 11150 Santa Monica Blvd., Suite 400(C) CITY: Los Angeles
20	(D) STATE: CA (E) COUNTRY: USA (F) ZIP: 90025
25	(v) COMPUTER READABLE FORM: (A) MEDIUM TYPE: Diskette (B) COMPUTER: IBM Compatible (C) OPERATING SYSTEM: DOS (D) SOFTWARE: FastSEQ Version 2.0
30	(Vi) CURRENT APPLICATION DATA: (A) APPLICATION NUMBER: PCT/US97/ (B) FILING DATE: 01-AUG-1997 (C) CLASSIFICATION:
35	<pre>(vii) PRIOR APPLICATION DATA: (A) APPLICATION NUMBER: 60/023,033 (B) FILING DATE: 02-AUG-1996</pre>
40	<pre>(viii) ATTORNEY/AGENT INFORMATION: (A) NAME: Adriano, Sarah B (B) REGISTRATION NUMBER: 34,470 (C) REFERENCE/DOCKET NUMBER: 30436.43WOU1</pre>
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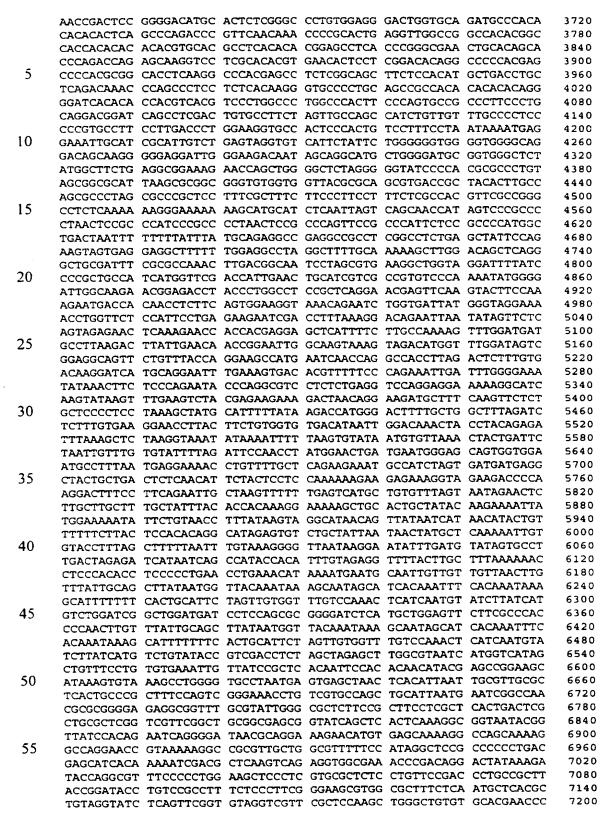
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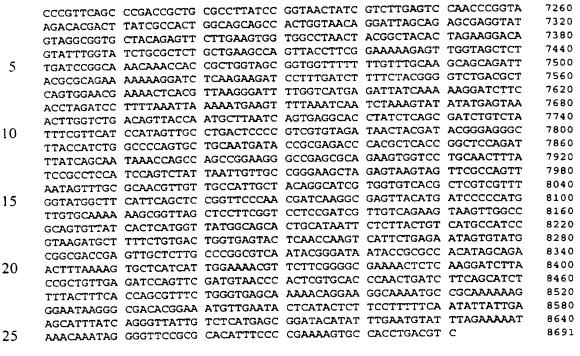
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J J	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
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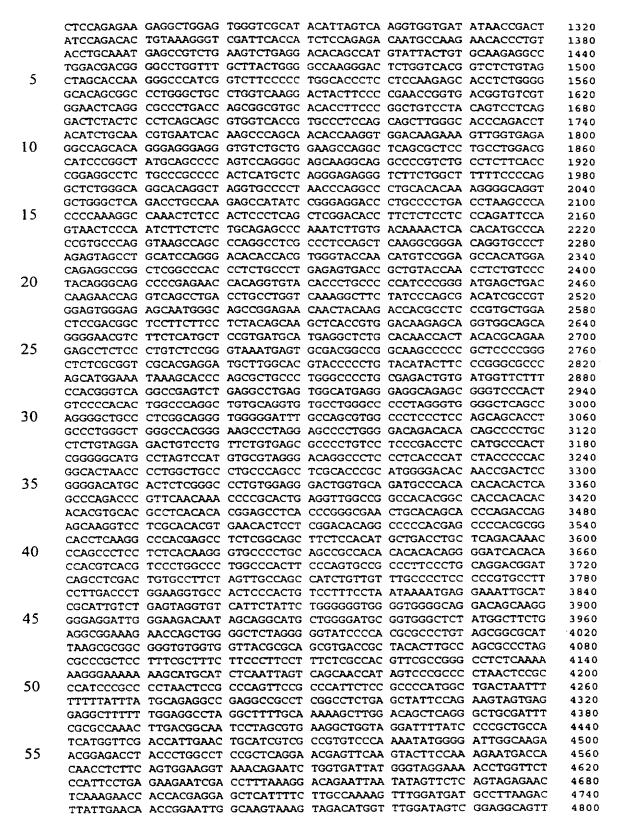
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55	TTAATACGAC	TCACTATAGG	GAGACCCAAG	CTTGGTACCA	ATTTAAATTG	ATATCTCCTT	1020
	AGGTCTCGAG	TCTCTAGATA	ACCGGTCAAT	CGATTGGAAT	TCTTGCGGCC	GCTTGCTAGC	1080
	CACCATGGAG	TTGTGGTTAA	GCTTGGTCCT	TCCTTGTCCT	TGTTTTAAAA	GGTGTCCAGT	1140
	GTGAAGTGAA	TCTGGTGGAG	TCTGGGGGAG	GCTTAGTGCA	GCCTGGAGGG	TCCCTGAAAG	1200
	TCTCCTGTGT	AACCTCTGGA	TTCACTTTCA	GTGACTATTA	CATGTATTGG	GTTCGCCAGA	1260

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13	TCATAATCAG	CCATACCACA	TTTGTAGAGG	TTTTACTTGC	TTTAAAAAAC	CTCCCACACC	5760
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20	TTATTGCAGC	TTATAATGGT	TACAAATAAA	GCAATAGCAT	CACAAATTTC	ACAAATAAAG	6060
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	TCTGTATACC	GTCGACCTCT	AGCTAGAGCT	TGGCGTAATC	ATGGTCATAG	CTGTTTCCTG	6180
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50	AAAATCGACG	CTCAAGTCAG	AGGTGGCGAA	ACCCGACAGG	ACTATAAAGA	TACCAGGCGT	6660
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(2) INFORMATION FOR SEQ ID NO:11:

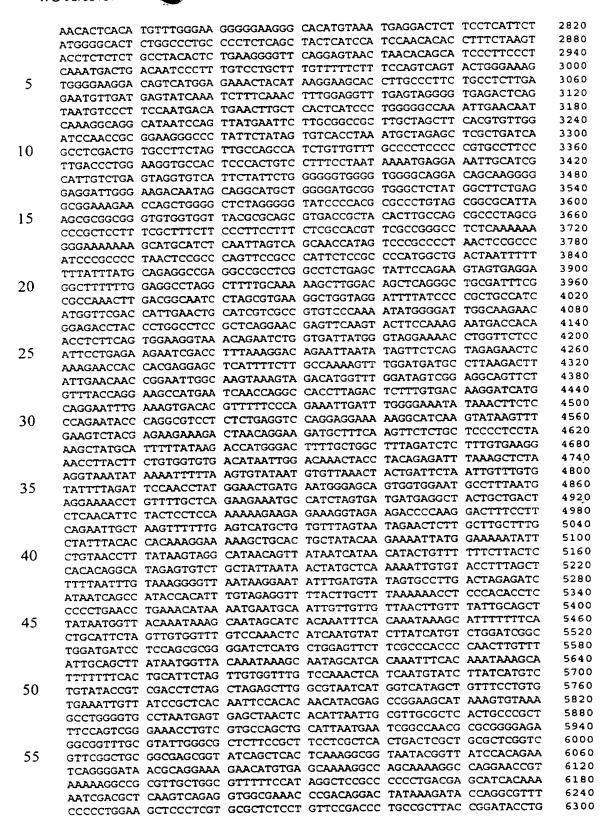
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 8897 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: cDNA

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

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45							
			**** EAR AEA	TD NO 10			

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 8321 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

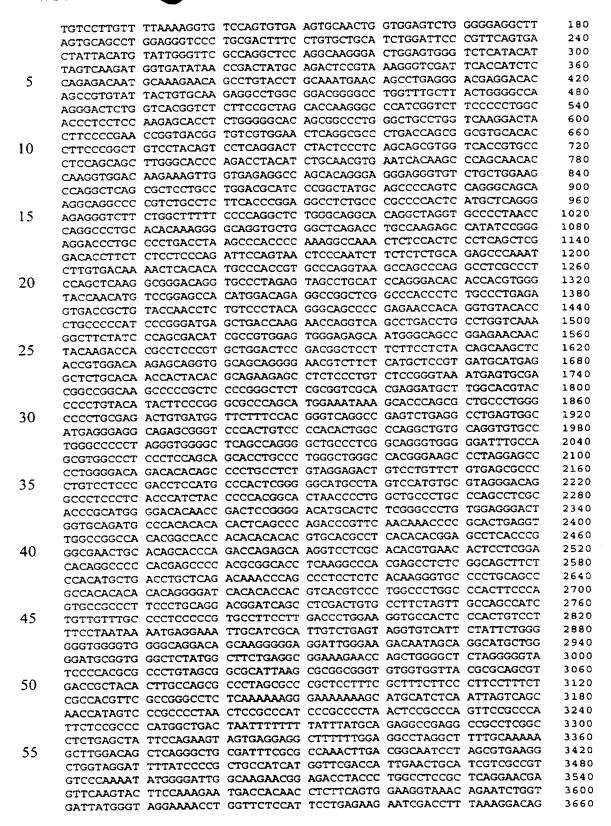
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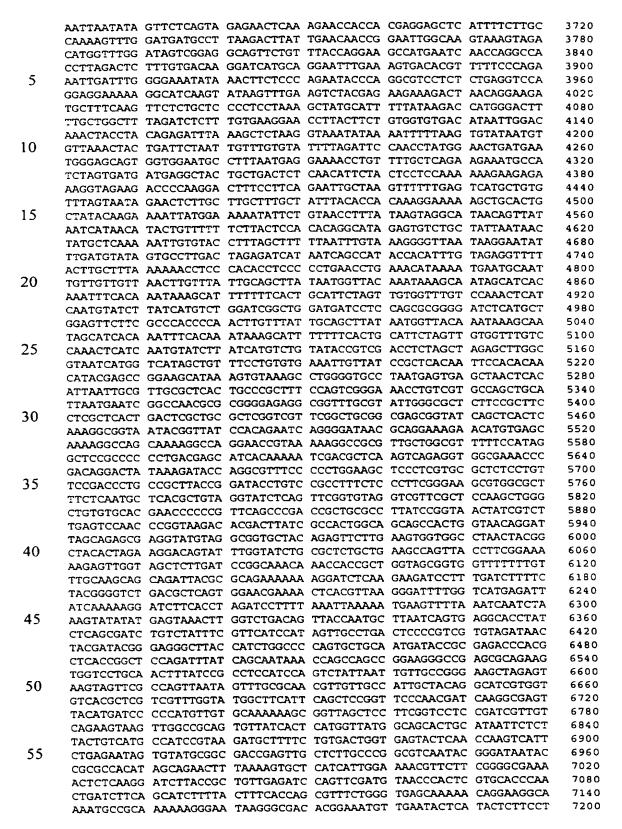
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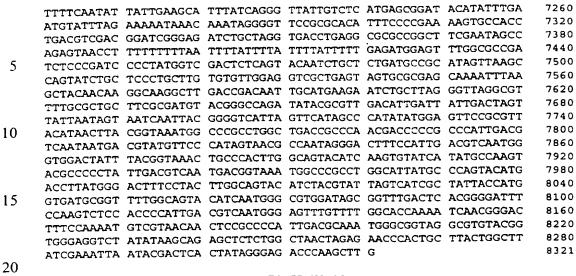
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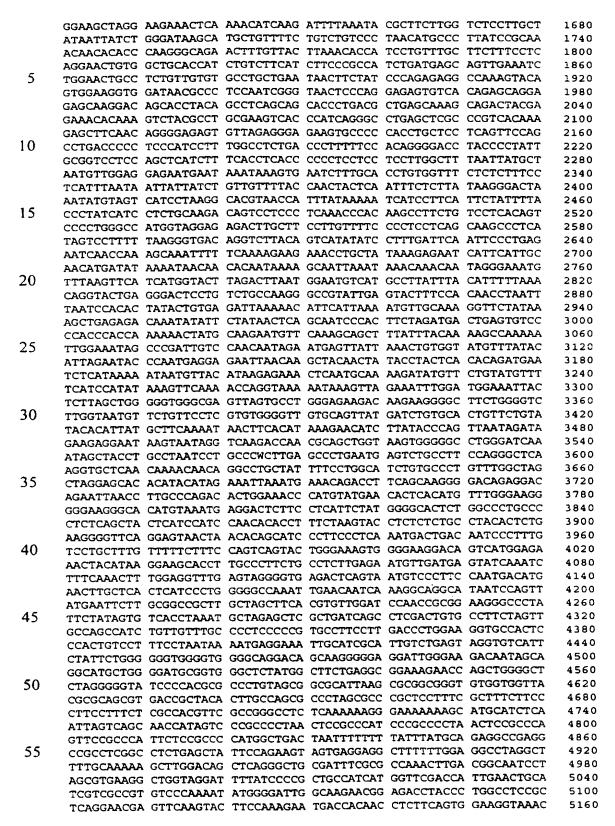
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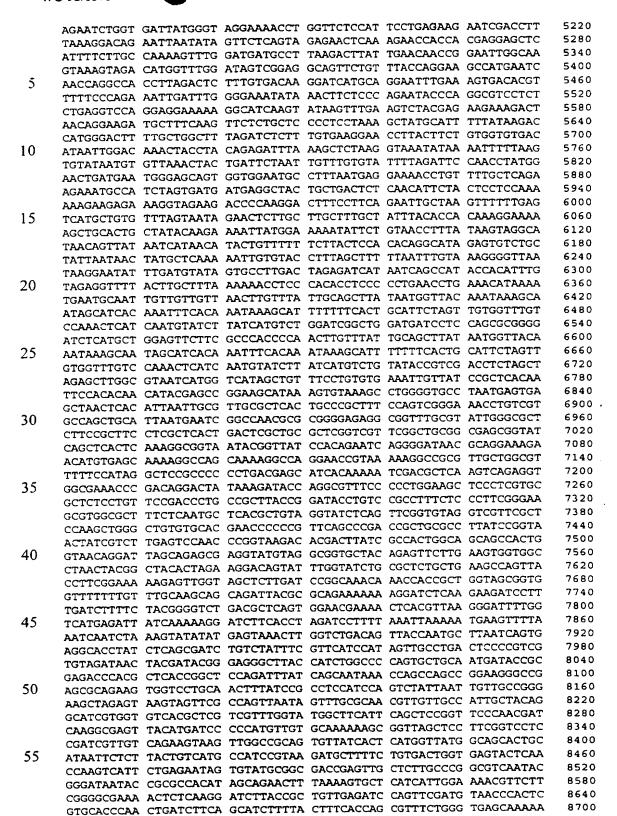


(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8897 base pairs
- 25 (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

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35	TCCCGATCCC	CTATGGTCGA	CTCTCAGTAC	AATCTGCTCT	GATGCCGCAT	AGTTAA GCCA	180
	GTATCTGCTC	CCTGCTTGTG	TGTTGGAGGT	CGCTGAGTAG	TGCGCGAGCA	AAATTTAAGC	240
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		TCAATTACGG					420
40	ATAACTTACG	GTAAATGGCC	CGCCTGGCTG	ACCGCCCAAC	GACCCCCGCC	CATTGACGTC	480
	AATAATGACG				TTCCATTGAC		540
		CGGTAAACTG					600
	GCCCCCTATT	GACGTCAATG					660
	CTTATGGGAC	TTTCCTACTT	GGCAGTACAT	CTACGTATTA	GTCATCGCTA	TTACCATGGT	720
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	AAGTCTCCAC	CCCATTGACG	TCAATGGGAG	TTTGTTTTGG	CACCAAAATC	AACGGGACTT	840
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CAGGAAGGCA	AAATGCCGCA	AAAAAGGGAA	TAAGGGCGAC	ACGGAAATGT	TGAATACTCA	8760
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What is claimed is:

A method for inhibiting immunoglobulin-induced toxicity resulting from 1. immunoglobulin immunotherapy in a subject comprising administering an 5 immunoglobulin molecule to the subject, the immunoglobulin molecule having a variable region and a constant region, the immunoglobulin molecule being modified prior to administration by structurally altering multiple toxicity associated domains in the constant region so that immunoglobulininduced toxicity is inhibited.

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A method for inhibiting immunoglobulin-induced toxicity resulting from 2. immunoglobulin immunotherapy in a subject comprising administering a structurally altered antibody to the subject, the structurally altered antibody comprising a variable region and a constant region, multiple toxicity associated domains in the constant region being modified so as to render the constant region unable to mediate an ADCC response or activate complement thereby inhibiting immunoglobulin-induced toxicity resulting from immunotherapy.

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A method for inhibiting immunoglobulin-induced toxicity resulting from 3. immunotherapy in a subject comprising administering an Ig fusion protein to the subject, the Ig fusion protein having multiple structurally altered toxicity associated domains in the constant region.

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A method for inhibiting immunoglobulin-induced toxicity resulting from 4. immunotherapy in a subject comprising administering an Ig fusion protein to the subject, the Ig fusion protein comprising a modified constant region, the

modification being a structural alteration in multiple toxicity associated regions within the CH₂ domain.

- A method for preventing immunoglobulin-induced toxicity resulting from
 immunotherapy for a disease in a subject comprising:
 - selecting an immunoglobulin which recognizes and binds a target, the target being associated with the disease;
- 10 (b) mutating the immunoglobulin so selected by structurally altering multiple toxicity associated domains in the constant region of the immunoglobulin thereby creating a structurally altered immunoglobulin;
- 15 (c) administering the structurally altered immunoglobulin of step (b) to the subject under conditions so that the structurally altered immunoglobulin recognizes and binds the target thereby alleviating symptoms associated with the disease, the structural alteration of the constant region thereby preventing immunoglobulin-induced toxicity in the subject.
 - 6. A method for preventing immunoglobulin-induced toxicity resulting from immunotherapy for a disease in a subject comprising:
- 25 (a) selecting an Ig fusion protein which recognizes and binds a target, the target being associated with the disease;
 - (b) structurally altering multiple toxicity associated domains in the CH₂ domain of the constant region of the Ig protein so selected;

(c) administering the structurally altered Ig fusion protein of step (b) to the subject under conditions so that the structurally altered Ig fusion protein recognizes and binds the target thereby alleviating symptoms associated with the disease, the structural alteration of the CH₂ domain thereby preventing immunoglobulin-induced toxicity in the subject.

- 7. The method of claim 1, 2, 3, 4, 5, or 6, wherein the portion of the constant region is the CH₂ domain.
 - 8. The method of claim 1 or 5, wherein the immunoglobulin molecule is IgG.
 - 9. The method of claim 1 or 5, wherein the immunoglobulin molecule is IgM.
 - 10. The method of claim 1 or 5, wherein the immunoglobulin molecule is IgA.
 - 11. The method of claim 2, wherein the antibody recognizes and binds Le^y.
- 20 12. The method of claim 2, wherein the antibody recognizes and binds to Lex.
 - 13. The method of claim 2, wherein the antibody is a monoclonal antibody BR96 produced by the hybridoma having the identifying characteristics of HB 10036 as deposited with the ATCC.
 - The method of claim 2, wherein the antibody is a chimeric antibody

 ChiBR96 produced by the hybridoma having the identifying characteristics

 of HB 10460 as deposited with the ATCC.

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15. The method of claim 1 or 5, wherein the immunoglobulin recognizes and binds Le^y.

- The method of claim 1 or 5, wherein the immunoglobulin recognizes and
 binds to Le^x.
 - 17. The method of claim 1 or 5, wherein the immunoglobulin is a monoclonal antibody BR96 produced by the hybridoma having the identifying characteristics of HB 10036 as deposited with the ATCC.

18. The method of claim 1 or 5, wherein the immunoglobulin is a chimeric antibody ChiBR96 produced by the hybridoma having the identifying characteristics of HB 10460 as deposited with the ATCC.

- 15 19. The method of claim 3, 4, or 6, wherein the Ig fusion protein recognizes and binds Le^y.
 - 20. The method of claim 3, 4, or 6, wherein the Ig fusion protein recognizes and binds to Le^x.
 - 21. The method of claim 3, 4, or 6, wherein the Ig fusion protein is a derivative of monoclonal antibody BR96 produced by the hybridoma having the identifying characteristics of HB 10036 as deposited with the ATCC.
- 25 22. The method of claim 3, 4, or 6, wherein the Ig fusion protein is a derivative of chimeric antibody ChiBR96 produced by the hybridoma having the identifying characteristics of HB 10460 as deposited with the ATCC.
 - 23. A pharmaceutical composition comprising a pharmaceutically effective

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amount of a structurally altered immunoglobulin, and an acceptable carrier, the structurally altered immunoglobulin (1) recognizes and binds a target, the target is associated with cancer and (2) has an inactivated CH₂ domain.

- A pharmaceutical composition comprising a pharmaceutically effective amount of structurally altered Ig fusion protein, and an acceptable carrier, the structurally altered Ig fusion protein (1) recognizes and binds a target, the target is associated with cancer and (2) has an inactivated CH₂ domain.
- A method of treating carcinomas in vivo comprising administering to a subject a pharmaceutically effective amount of the composition of claim 23 or 24.
- The method of claim 30, wherein the structurally altered immunoglobulin in the composition is labeled so as to directly or indirectly produce a detectable signal with a compound selected from the group consisting of a radiolabel, an enzyme, a chromophore, a chemiluminescer, and a fluorescer.
- The method of claim 24, wherein the Ig fusion protein in the composition is labeled so as to directly or indirectly produce a detectable signal with a compound selected from the group consisting of a radiolabel, an enzyme, a chromophore, a chemiluminescer, and a fluorescer.
- 28. The method of claim 2 or 5, wherein the antibody is conjugated to a cytotoxic agent.
 - 29. The method of claim 1, wherein the immunoglobulin is conjugated to a cytotoxic agent.

30. The method of claim 3, 4 or 6, wherein the Ig fusion protein is conjugated to a cytotoxic agent.

- The method of claim 28, 29, or 31, wherein the cytotoxic agent is selected from the group consisting of antimetabolites, alkylating agents, anthracyclines, antibiotics, anti-mitotic agents, and chemotherapeutic agents.
- 32. A method for treating a subject suffering from a cancer, the cancer being characterized as a group of cells having a tumor associated antigen on the cell surface, which method comprises administering to the subject a cancer killing amount of the composition of claim 23 or 24 joined to a cytotoxic agent under conditions which permit the molecule so joined to bind the tumor associated antigen on the cell surface so as to kill the cells so bound thereby curing the subject.
 - 33. A pharmaceutical composition comprising a pharmaceutically effective amount of a structurally altered BR96 antibody, the structurally altered antibody having an inactivated CH₂ domain.
 - 34. A method for treating a subject suffering from a proliferative type disease characterized by cells having a BR96 antigen on the cell surface which comprises administering to the subject an effective amount of the composition of claim 33 joined to doxorubicin such that the immunoconjugate binds the BR96 antigen and kills said cells thereby treating the subject.
 - 35. A method for inhibiting BR96 (ATCC: HB10036) induced toxicity resulting from immunoglobulin immunotherapy in a subject comprising administering

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BR96 to the subject, the BR96 molecule being modified prior to administration, the modification comprising the deletion or substitution of at least one amino acid residue in the toxicity associated domain localized to amino acids 310-331 and the deletion or substitution of at least one amino acid residue in the toxicity associated domain localized to amino acids 231-238 so that complement and Fc receptor mediated toxicity is inhibited.

- 36. A method for preventing BR96 (ATCC: HB10036) induced toxicity resulting from immunotherapy for cancer in a subject comprising:
 - (a) mutating the BR96 polypeptide by the deletion or substitution of at least one amino acid residue in the toxicity associated domain localized to amino acids 310-331 and the deletion or substitution of at least one amino acid residue in the toxicity associated domain localized to amino acids 231-238 so that complement and Fc receptor mediated immunoglobulin-induced toxicity is inhibited in the altered BR96 polypeptide; and
- (b) administering the structurally altered BR96 polypeptide of step (a) to the subject under conditions so that the peptide recognizes and binds cancer associated Le^y antigens, thereby alleviating symptoms associated with the cancer, the structural alteration of the toxicity associated domains thereby preventing BR96 toxicity in the subject.
- 25 37. A chimeric BR96 antibody having a structurally altered constant region having the CH1 and CH3 domains but not the CH2 domain, the antibody being designated cBR96-A.

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38. The chimeric BR96 antibody of claim 37 which is expressed by the plasmid having the sequence shown in SEQ ID NO. 10.

- 39. A BR96 antibody having humanized variable and constant regions, wherein the constant region has been structurally altered so that the CH1 and CH3 domains are present but the CH2 domain is not, the antibody being designated hBR96-2A.
- 40. The BR96 antibody of claim 39 which is expressed by the plasmid having the sequence shown in SEQ ID NO. 12.
 - 41. A BR96 antibody designated hBR96-2B having a structurally altered constant region wherein leucine at amino acid position 235 is mutated to alanine and glycine at amino acid position 237 is mutated to alanine.
- 42. A BR96 antibody designated hBR96-2C having a structurally altered constant region wherein glutamic acid at amino acid position 318 is mutated to serine; lysine at amino acid position 320 is mutated to serine; and lysine at amino acid position 322 is mutated to serine.
 - 43. A BR96 antibody designated hBR96-2D having a structurally altered constant region wherein proline at amino acid position 331 is mutated to alanine.
- A BR96 antibody designated hBR96-2E having a structurally altered constant region wherein leucine at amino acid position 235 is mutated to alanine; glycine at amino acid position 237 is mutated to alanine; glutamic acid at amino acid position 318 is mutated to serine; lysine at amino acid

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position 320 is mutated to serine; and lysine at amino acid position 322 is mutated to serine.

- A BR96 antibody designated hBR96-2F having a structurally altered constant region wherein leucine at amino acid position 235 is mutated to alanine; glycine at amino acid position 237 is mutated to alanine; and proline at amino acid position 331 is mutated to alanine.
- 46. A BR96 antibody designated hBR96-2G having a structurally altered constant region wherein glutamic acid at amino acid position 318 is mutated to serine; lysine at amino acid position 320 is mutated to serine; and lysine at amino acid position 322 is mutated to serine; and proline at amino acid position 331 is mutated to alanine.
- A BR96 antibody designated hBR96-2H having a structurally altered constant region wherein leucine at amino acid position 235 is mutated to alanine; glycine at amino acid position 237 is mutated to alanine; glutamic acid at amino acid position 318 is mutated to serine; lysine at amino acid position 320 is mutated to serine; lysine at amino acid position 322 is mutated to serine; and proline at amino acid position 331 is mutated to alanine.
 - 48. A nucleic acid molecule which encodes the BR96 antibody of claim 37, 39, and 41-47.
 - 49. A cDNA of claim 48.
 - 50. A plasmid which comprises the nucleic acid molecule of claim 48.

51. A host vector system comprising a plasmid of claim 50 in a suitable host cell.

52. A method for producing a protein comprising growing the host vector system of claim 51 so as to produce the protein in the host and recovering the protein so produced.

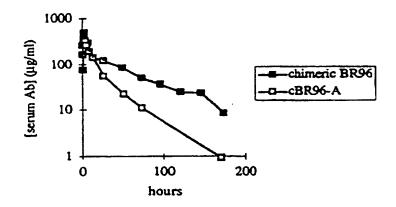


Figure 1. Plasma clearance in high LeY expressing dogs chimeric versus constant region mutant of c8R96-2.

Figure One

Figure 2

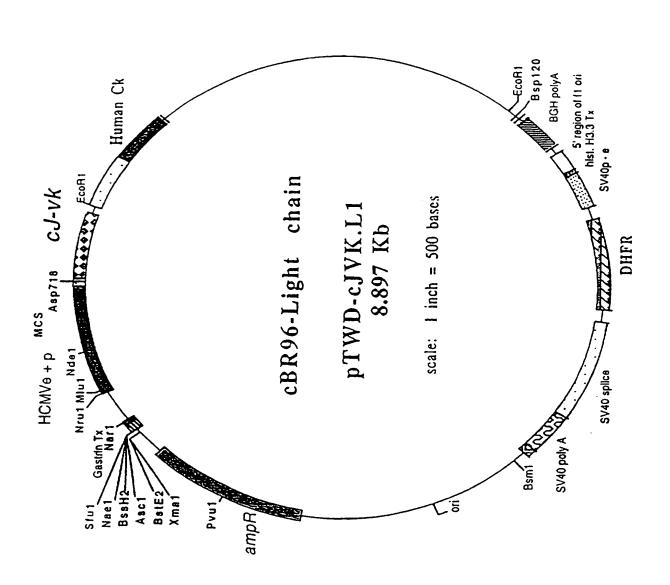


Figure 3

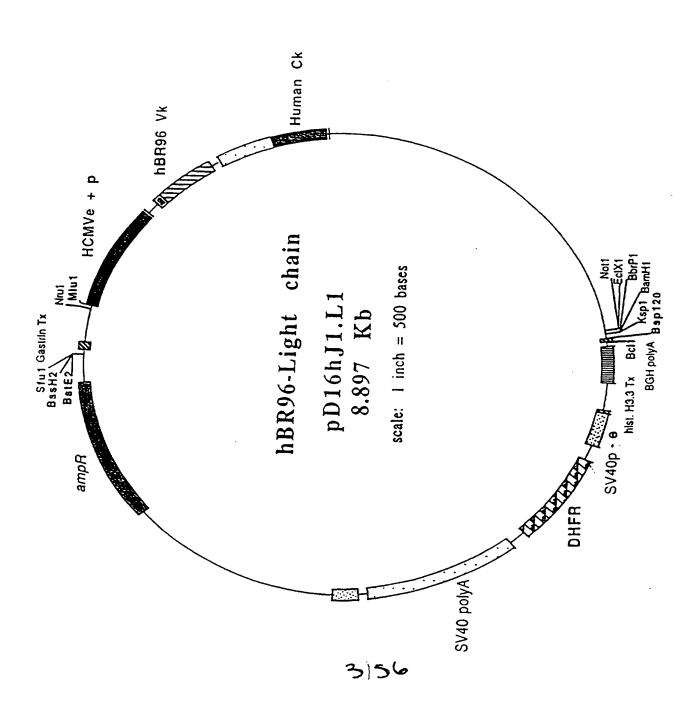


Figure 4

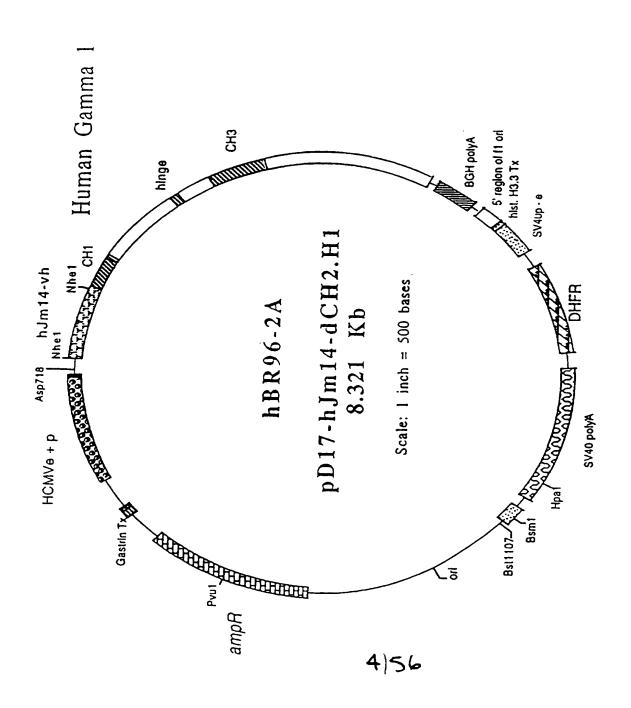
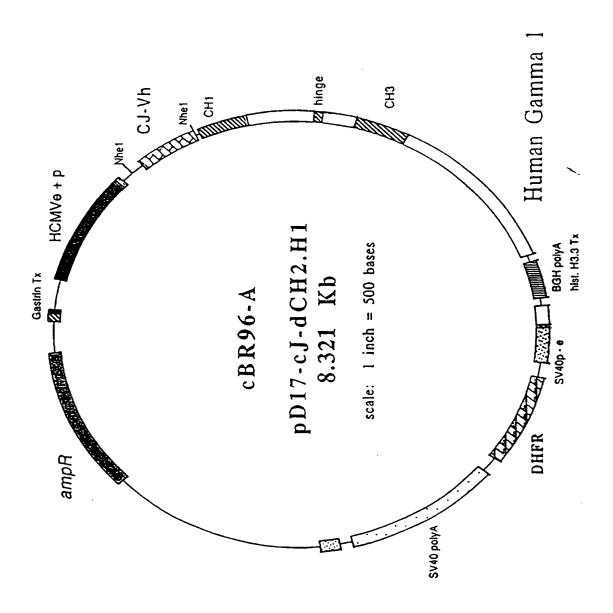
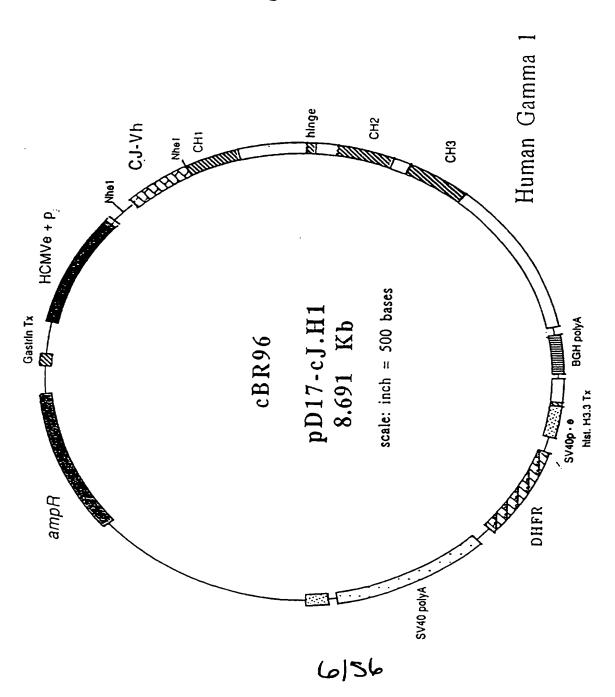


Figure 5



5)56

Figure 6



PCT/US97/13562

Figure 7

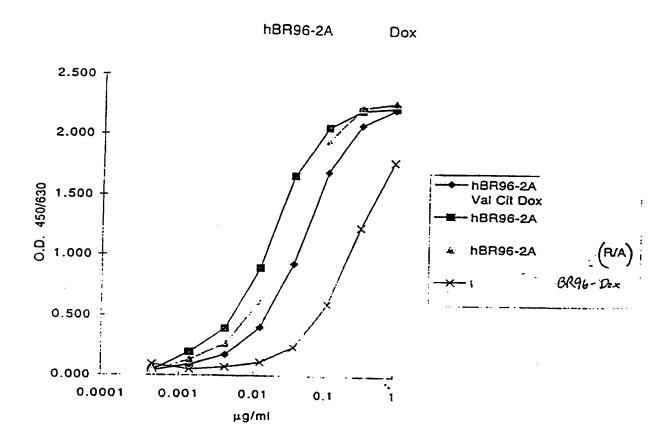
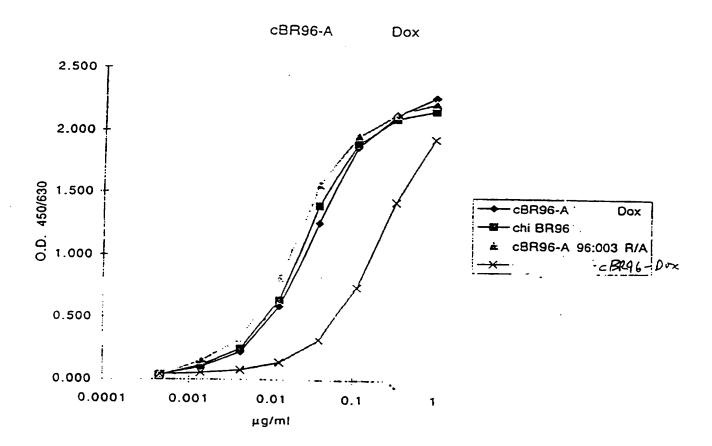
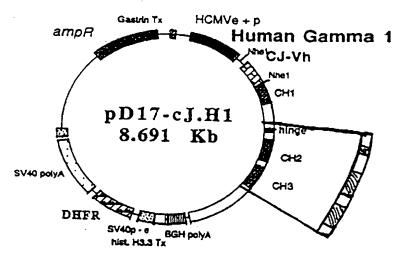


Figure 8



A- Hinge + Cl... + CH3 domains were removed from R96 IgG1 construct by E.co -III restriction digestion.



3. 2 - Hinge + CH3 domains amplified by PCR from L6 IgG1 construct lacking the CH2 domain.

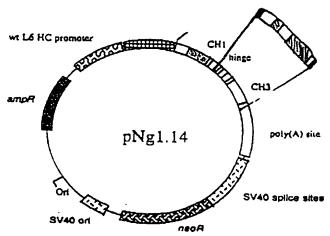


Figure 9

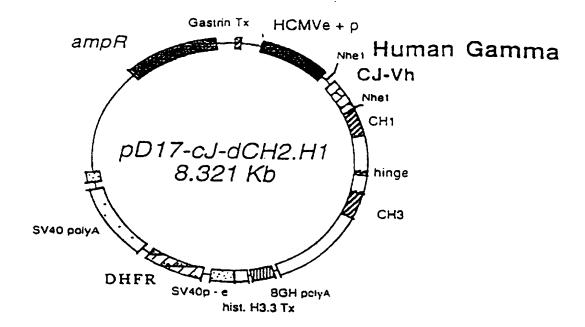
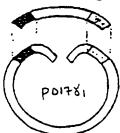


Figure 9 (CONTINUED)

- 1- Introduction of mutations by site-directed mutagenesis on double-stranded plasmid DNA.
- A- Mutations introduced into synthetic oligonucleotides used for the PCR amplification of CH2 domain.

B- Plasmid DNA linearized inside CH2 domain and cotransformed with PCR fragment into competent DH5 α .

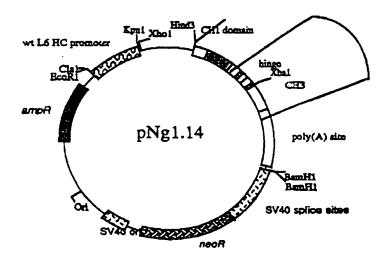


C- Cloning mediated by homologous recombination yields transformants harbouring recombinant plasmids.



Figure 10

Figure 11



PCT/US97/13562

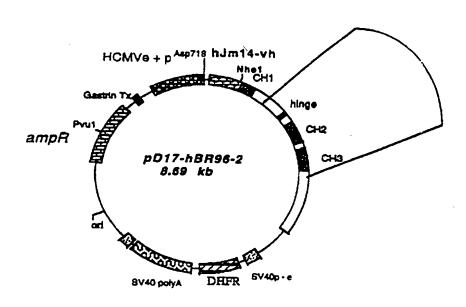


Figure 12

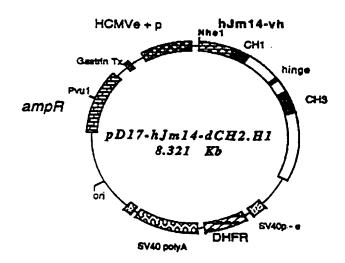


Figure 13

90	180	270	360	450	540	630	720	810	900
TTATTTTATT	AGCCAGTATC	AATTGCATGA	AGTTATTAAT	GGCTGACCGC	TGGGTGGACT	AAATGGCCCG	ATGGTGATGC	GGCAGTTTGT	CGGTGGGAGG
AATAAAATAA	TCGGTCATAG	TTAACGTACT	TCAATAATTA	CCGACTGGCG	ACCCACCTGA	TTTACCGGGC	TACCACTACG	CCCTCAAACA	GCCACCCTCC
80	170	260	350	440	530	620	710	800	890
Taattttaft	CGCATAGTTA	CTTGACCGAC	ATTATTGACT	TGGCCCGCCT	TTGACGTCAA	CAATGACGGT	CGCTATTACC	TGACGTCAAT	TAGGCGTGTA
Attaaaataa	GCGTATCAAT	GAACTGGCTG	TAATAACTGA	ACCGGGCGGA	AACTGCAGTT	GTTACTGCCA	GCGATAATGG	ACTGCAGTTA	ATCCGCACAT
70	160	250	340	430	520	610	700	790	880
CCTTTTTTTT	GCTCTGATGC	CAAGGCAAGG	GTTGACATTG	TTACGGTAAA	GGACTTTCCA	CTATTGACGT	TATTAGTCAT	TCCACCCCAT	AAATGGGCGG
GGAAAAAAA	CGAGACTACG	GTTCCGTTCC	CAACTGTAAC	AATGCCATTT	CCTGAAAGGT	GATAACTGCA	ATAATCAGTA	AGGTGGGGTA	TYTACCCGCC
60	150	240	330	420 430 440 450 GTTACATAAC TTACGGTAAA TGGCCCGCCT GGCTGACCGC CAATGTATTG AATGCCATTT ACCGGGCGGA CCGACTGGCG	510	600	690	780	870
GCCAGAGTAA	AGTACAATCT	TAAGCTACAA	AGATATACGC		ACGCCAATAG	AGTACGCCCC	TACATCTACG	TTTCCAAGTC	CCATTGACGC
CGGTCTCATT	TCATGTTAGA	ATTCGATGTT	TCTATATGCG		TGCGGTTATC	TCATGCGGGG	ATGTAGATGC	AAAGGTTCAG	GGTAACTGCG
50	140	230	320	410	500	590	680	770	860
GCTTCGAATA	GTCGACTCTC	GAGCAAAATT	TGTACGGGCC	GGAGTTCCGC	TCCCATAGTA	TCATATGCCA	TACTTGGCAG	CTCACGGGGA	CAACTCCGCC
CGAAGCTTAT	CAGCTGAGAG	CTCGTTTTAA	ACATGCCCGG	CCTCAAGGCG	AGGGTATCAT	AGTATACGGT	ATGAACCGTC	GAGTGCCCCT	GTTGAGGCGG
AGGCGCGCCG TCCGCGCGGC	CGATCTCCCG ATCCCCTATG GTCGACTCTC AGTACAATCT GCTCTGATGC CGCATAGTTA AGCCAGTATC GCTAGAGGC TAGGGGATAC CAGCTGAGAG TCATGTTAGA CGAGACTACG GCGTATCAAT TCGGTCATAG	220 AGTAGTGCGC TCATCACGCG	300 310 350 360 COTTUGUE 300 310 340 340 340 340 350 360 COTTUBUE COTTUGUE TOUTTUGUE TOUTTUGUE TOUTTUGUE TOUTTUGUE TOUTTUGUE ACATACICE ACATACICE TOUTTUGUE TAATAACIGA TEATAATTA	400 GCCCATATAT CGGGTATATA	490 TGACGTATGT ACTGCATACA	580 ATCAAGTGTA TAGTTCACAT	670 GGGACTTTCC CCCTGAAAGG	760 AGCGGTTTGA TCGCCAAACT	850 AATGTCGTAA TTACAGCATT
30	120	210	300	390	480	570	660	750	840
AGGTGACCTG	CGATCTCCCG	CAGGTCGCTG	CGTTTTGCGC	TTAGTTCATA	ACGTCAATAA	TTGGCAGTAC	ATGACCTTAT	GGCGTGGAT	GACTITICCAA
TCCACTGGAC	GCTAGAGGGC	CTCCAGCGAC	GCAAAACGCG	AATCAAGTAT	TGCAGTTATT	AACCGTCATG	TACTGGAATA	CCCGCACCTA	CTGAAAGGTT
10 20 30 80 90 50 50 50 80 GACGCGCCGCCG GCTTCGAATA GCCAGAGTAA CCTTTTTTTT TAATTTTATT TTATTTTATT	110 110 150 150 180 180 180 180 180 180 180 180 180 18	190 200 210 200 270 TGCTCCCTGC TIGIGIGIGIGG GAGGTCGCTG AGTAGTGCGC GAGCAAAATT TAAGCTACAA CAAGGCAAGG	280 280 340 350 350 350 360 300 300 320 330 340 340 350 360 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 350 360 360 360 360 360 360 360 360 360 36	370 380 440 450 420 420 430 450 450 450 450 450 450 450 450 450 45	460 470 480 490 500 500 510 520 530 540 540 500 500 510 520 530 540 540 540 540 540 540 540 540 540 54	550 550 620 630 630 630 630 630 630 630 630 630 63	640 650 710 710 720 CCTGGCATTA TGCCCAGTAC ATGACCTTACC TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC GGACCGTAAT ACGGGTCATG TACTGGAATA CCCTGAAAGG ATGAACCGTC ATGTAGATGC ATAATCAGTA GCGATAATGG TACCACTACG	710 740 800 800 810 GGTTTTGG 770 780 790 800 800 810 GGTTTTGGGGGA GTACATCACCCCAT TGACGTCAAT GGGCGTCAAT GGCGTTTGA CTCACGCCCA AAGGTTCAG AGGTGGGGTA ACTGCAGTTA CCCGCACCTA TCGCCAAACC AAAGGTTCAG AGGTGGGGTA ACTGCAGTTA CCCTCAAACA	820 880 880 890 900 TYPGGCACCA JAATCAACGG GACTTICCAA AATGTCGTAA CAACTCGGCC CCATTGACGC AAATGGGCGG TAGGCGTGTA CGGTGGGAGG AAACCGTGGT TYTAGTTGCC CTGAAAGGTT TYACAGCATT GTTGAGGCGG GGTAACTGCG TYTACCGGCC ATCGGCACAT GCCACCCTCC
10	100	190	280	370	460	550	640	730	820
GACGGATCGG	TTTGAGATGG	TGCTCCCTGC	AGAATCTGCT	AGTAATCAAT	CCAACGACCC	ATTTACGGTA	CCTGGCATTA	GGTTTTGGCA	TTTGGCACCA
CTGCCTAGCC	AAACTCTACC	ACGAGGGACG	TCTTAGACGA	TCATTAGTTA	GGTTGCTGGG	TAAATGCCAT	GGACCGTAAT	CCAAAACCGT	AAACCGTGGT

Figure 14

990	1080	1170	1260	1350	1440	1530	1620	1710	1800
GAGACCCAAG	GCTTGCTAGC	TCTCGGGGAG	GTTCGCCAGA	CGATTCACCA	GCAAGAGGCC	GTCTTCCCCC	ACGGTGTCGT	GTGGTCACCG	GTTGGTGAGA
CTCTGGGTTC	CGAACGATCG	AGACCCCCTC	CAAGCGGTCT	GCTAAGTGGT	CGTTCTCCGG	CAGAAGGGGG	TGCCACAGCA	CACCAGTGGC	CAACCACTCT
980	1060 1070	1160	1250	1340	1420 1430	1520	1610	1700	1790
TCACTATAGG	CGATTGGAAT TCTTGCGGCC	TCTGGTGGAG	CATGTATTGG	TCTAAAGGGT	ACACAGCCAT GTATTACTGT	GGGCCCATCG	CGAACCGGTG	CCTCAGCAGC	GGACAAGAAA
AGTGATATCC	GCTAACCTTA AGAACGCCGG	AGACCACCTC	GTACATAACC	ACATTTCCCA	TGTGTGGGTA CATAATGACA	CCCGGGTAGC	GCTTGGCCAC	GGAGTCGTCG	CCTGTTCTTT
970	1060	1150	1240	1330	1420	1510	1600	1690 1700	1780
TTAATACGAC	CGATTGGAAT	GTGAAGTGAA	GTGACTATTA	ATCCAGACAC	ACACAGCCAT	CTAGCACCAA	ACTACTTCCC	GACTCTACTC CCTCAGCAGC	ACACCAAGGT
AATTATGCTG	GCTAACCTTA	CACTTCACTT	CACTGATAAT	TAGGTCTGTG	TGTGTCGGTA	GATCGTGGTT	TGATGAAGGG	CTGAGATGAG GGAGTCGTCG	TGTGGTTCCA
960	1050	1140	1230	1320	1410	1500	1590	1680	1770
CTTATCGAAA	ACCGGTCAAT	GGTGTCCAGT	TTCACTTTCA	ATAACCGACT	AAGTCTGAGG	GTCTCTGTAG	CTGGTCAAGG	CAGTCCTCAG	AAGCCCAGCA
GAATAGCTTT	TGGCCAGTTA	CCACAGGTCA	AAGTGAAAGT	TATTGGCTGA	TTCAGACTCC	CAGAGACATC	GACCAGTTCC	GTCAGGAGTC	TYCGGGTCGT
950	1040	1130	1220	1310	1400	1490	1580	1670	1760
TGCTTACTGG	TCTCTAGATA	TGTTTTAAAA	AACCTCTGGA	AGGTGGTGAT	GAGCCGTCTG	TCTGGTCACG	CCTGGGCTGC	GGCTGTCCTA	CGTGAATCAC
ACGAATGACC	AGAGATCTAT	ACAAAATTTT	TTGGAGACCT	TCCACCACTA	CTCGGCAGAC	AGACCAGTGC	GGACCCGACG	CCGACAGGAT	GCACTTAGTG
CAGAGCTCTC TGGCTAACTA GAGAACCCAC TGCTTACTGG CTTATCGAAA TTAATACGAC TCACTATAGG GTCTCGAGAG ACCGATTGAT CTCTTGGGTG ACGAATGACC GAATAGGTTT AATTATGCTG AGTGATATCC	ATATCTCCTT AGGTCTCGAG TCTCTAGATA ACCGGTCAAT CGATTGGAAT TCTTGCGGCC GCTTGCTAGC TATAGAGGAA TCCAGAGCTC AGAGATCTAT TGGCCAGTTA GCTAACCTTA AGAACGCCGG CGAACGATCG	GCTTGGTCCT TCCTTGTCCT TGTTTTAAAA GGTGTCCAGT GTGAAGTGAA	1210 TCTCCTGTGT AGAGGACACA	1300 ACATTAGTCA TGTAATCAGT	1390 ACCTGCAAAT TGGACGTTTA	1480 GCCAAGOGAC CGGTYCCCTG	1570 GCACAGCGGC CGTGTCGCCG	1660 ACACCTTCCC TGTGGAAGGG	1750 ACATCTGCAA TGTAGACGTT
930 TGGCTAACTA ACCGATTGAT	1020 ATATCTCCTT TATAGAGGAA	1110 GCTTGGTCCT CGAACCAGGA	1200 TCCCTGAAAG AGGGACTTTC	1290 TGGGTCGCAT ACCCAGCGTA	1380 AACACCCTGT TTGTGGGACA	1470 GCTTACTGGG CGAATGACCC	1560 ACCTCTGGGG TGGAGACCCC	1650 AGCGGCGTGC TCGCCGCACG	ACCCAGACCT TGGGTCTGGA
	1010 ATTTAAATTG TAAATTTAAC	1090 1100 1110 1120 1130 1130 1140 1150 1160 1170 1170 CACCATGGAG TYCTGGTTGAA GCTGTGGAG TYCTGGTGGAG TYCTGGAGGTGAG TYCTGGAGGTGAG TYCTGGAGGTGAG TYCTGGAGGTGAG TYCTGGAGGGAG TYCTGGAGGAG TYCTGGAGGAGAG TYCTGGAGGAGAG TYCTGGAGGAGAG TYCTGGAGGAGAG TYCTGGAGGAGAG TYCTGGAGGAGAG TYCTGGAGGAGAGAGAGAGAGAGAGAGAGAGAGAGAGAGAGA	1180 1190 1200 1210 1220 1230 1230 1230 1240 1250 1260 CCTGGGGGG TCCCTGTGT AACCTCTGGA TTCACTTTCA GTGACTATTA CATGTATTGG GTTCGCCAGA CGAATCACGT CGGACCTCCC AGGGACTTTC AGAGGACACA TTGGAGACCT AAGTGAAAGT CACTGATAAT GTACATAACC CAAGCGGTCT	1270 1280 1290 1390 1300 1310 1320 1330 1330 1340 1350 CTCCAGAGAA GAGGCTGGAG TGGGTCGCA ACATTAGTCA AGGTGGTGAT ATAACCGACT ATCCAGACAC TGTAAAGGGT CGATTCACCA GAGGTCTCT CTCCGACCTC ACCCAGCGTA TGTAATCAGT TCCACCACTA TATTGGCTGA TAGGTCTGTG ACATTTCCCA GCTAAGTGGT	1340 1340 1410 1420 1430 1440 TCTCCAGAGA CAATGCCAAG AACACCCTGT ACTGCAAAT GAGCCGTCTG AAGTCTGAGG ACACAGCCAT GTATTACTGT GCAAGAGGCC AGAGGICTCT GTTACGGTTC TTGTGGGACA TGGACGTTTA CTCGGCAGAC TTCAGACTCC TGTGTGGGTA CATAATGACA CGTTCTCGG	1450 1510 1510 1520 1530 1530 1530 1550 1550 1550 1550 155	1540 1550 1560 1560 1570 1580 1590 1600 1600 1610 1620 1600 TOCCAAGAGC ACCTCTCC CGAACCGGTG ACGGTGTCGT ACCGTGGAAG GAGGTCTCG TGATCACG GAACCCGAC GGACCCGACG GACCAGGAG GAGGTTCTCG TGATGAAGGG GCTTGGCCAC TGCCACAGCA	1630 1640 1650 1650 1650 1670 1680 1680 1780 1790 1710 1710 1710 1710 1710 1710 171	1720 1730 1740 1750 1760 1760 1770 1780 1790 1890 1800 1800 1780 1800 1800 1800 1800 180
910	1000	1090	1180	1270	1360	1450	1540	1630	1720
TCTATATAAG	CTTGGTACCA	CACCATGGAG	GCTTAGTGCA	CTCCAGAGAA	TCTCCAGAGA	TGGACGACGG	TGGCACCCTC	GGAACTCAGG	TGCCCTCCAG
AGATATATTC	GAACCATGGT	GTGGTACCTC	CGAATCACGT	GAGGTCTCTT	AGAGGTCTCT	ACCTGCTGCC	ACCGTGGGAG	CCTTGAGTCC	ACGGGAGGTC

Figure 14 (continued)

1890	1980	2070	2160	2250	2340	2430	2520	2610	2700
AGTCCAGGGC	TTTTCCCCAG	GAGCCATATC	CCAGATTCCA	CCAGGCCTCG	GCCACATGGA	CACAGGTGTA	ACATCGCCGT	TCTACAGCAA	ACACGCAGAA
TCAGGTCCCG	AAAAGGGGTC	CTCGGTATAG	GGTCTAAGGT	GGTCCGGAGC	CGGTGTACCT	GTGTCCACAT	TGTAGCGGCA	AGATGTCGTT	TGTGCGTCTT
1850 1860 1870 1880 1890 TCAGCGCTCC TGCCTGGACG CATCCCGGCT ATGCAGCCCC AGTCCAGGGC AGTCGCGAGG ACGCACCTGC GTAGGGCCGA TACGTCGGGG TCAGGTCCCG	1970 TCTTCTGGCT AGAAGACCGA	2060 GACCTGCCAA CTGGACGGTT	2140 2150 2150 2160 CTCGGACACC TTCTCTCCTC CCAGATTCCA GAGCCTGTGG AAGAGAGGAG GGTCTAAGGT	2240 GTAAGCCAGC CATTCGGTCG	ACAGTAGCCT GCATCCAGGG ACACACCACG TGGGTACCAA CATGTCCGGA GCCACATGGA TCTCATCGGA CGTAGGTCC TGTGTGGTGC ACCCATGGTT GTACAGGCCT CGGTGTACCT	2330 2430 2400 2410 2420 2430 6CTGTACCAA CCTCTGTCCC TACAGGGCAG CCCCGAGAAC CACAGGTGTA CGACATGGTT GGAGACAGGG ATGTCCCGTC GGGGCTCTTG GTGTCCACATA	2510 2520 TATCCCAGCG ACATCGCCGT ATAGGGTCGC TGTAGCGGCA	2510 2580 2590 2600 2600 CAACTACAAG ACCACGCCTC CCGTGCTGGA CTCCGACGC TCCTTCTTCC TCTACAGCAAA GTTGATGTTC TGGTGCGGAG GGCACGACCT GAGGCTGCCG AGGAAGAAGA AGATGTCGTT	2690 2700 CACAACCACT ACACGCAGAA GTGTTGGTGA TGTGCGTCTT
1870	1960	2050	2140	2230	2320	2410	2500	2590	2680
CATCCCGGCT	AGGGAGAGGG	GCTCGGCTCA	CTCGGACACC	CCGTGCCCAG	TGGGTACCAA	TACAGGGCAG	CAAAGGCTTC	CTCCGACGCC	TGAGGCTCTG
GTAGGGCCGA	TCCCTCTCCC	CGACCCGAGT	GAGCCTGTGG	GGCACGGGTC	ACCCATGGTT	ATGTCCCGTC	GTTTCCGAAG	GAGGCTGCCG	ACTCCGAGAC
1860	1940 1950	2040	2130	2220	2310	2400	2490	2580	2670
TGCCTGGACG	TGCCGCCCC ACTCATGCTC	AGGGGCAGGT	ACTCCCTCAG	CACATGCCCA	ACACACCACG	CCTCTGTCCC	CCTGCCTGGT	CCGTGCTGGA	CCGTGATGCA
ACGGACCTGC	ACGGCGGGG TGAGTACGAG	TCCCCGTCCA	TGAGGGAGTC	GTGTACGGGT	TGTGTGGTGC	GGAGACAGGG	GGACGGACCA	GGCACGACCT	GGCACTACGT
1850	1940	2030	2120	2210	2300	2390	2480	2570	2660
TCAGCGCTCC	TGCCCGCCCC	CTGCACACAA	CAAACTCTCC	ACAAAACTCA	GCATCCAGGG	GCTGTACCAA	GTCAGCCTGA	ACCACGCCTC	TTCTCATGCT
AGTCGCGAGG	ACGGCCGGG	GACGTGTGTT	GTTTGAGAGG	TGTTTTGAGT	CGTAGGTCCC	CGACATGGTT	CAGTCGGACT	TGGTGCGGAG	AAGAGTACGA
1840	1930	2020	2110	2200	2290	2380	2470	2560	2650
GAAGCCAGGC	CGGAGGCCTC	AACCCAGGCC	CCCCAAAGGC	AAATCTTGTG	AGAGTAGCCT	GAGAGTGACC	CAAGAACCAG	CAACTACAAG	GGGGAACGTC
CTTCGGTCCG	GCCTCCGGAG	TTGGGTCCGG	GGGGTTTCCG	TTTAGAACAC	TCTCATCGGA	CTCTCACTGG	GITCTTGGTC	GTTGATGITIC	CCCCTTGCAG
GCCAGCACACACACACACACACACACACACACACACACA	1920 CCTCTTCACC GGAGAAGTGG	1990 2000 2010 2020 2030 2030 2040 2050 2060 2070 GCTCGCCTCA GACCTGCCAA GAGGGCAGGT CCTCGCCTCA GACCTGCCAA GAGCCATATC CGAGACCCGT CCGTCTCCGA TCCACGGCC GACGTCTCG GACCTCGTCTA CGACCCGTCCA CGACCCGAC CTCGACGGTT CTCGGTATAG	2100 CCTAAGCCCA GGATTCGGGT	GTAACTCCCA ATCITICITCT TGCAGAGCCC AAATCITTGTG ACAAAACTCA CACATGCCCA CGTGCCCAG GTAAGCCAGC CATTGAGGT TAGAAGAGAG ACGTCTCGGG TTTAGAACAC TGTTTTGAG GTGTTTGAGGT GGCACGGGTC CATTCGTCG	CAGGTGCCCT AGAGTAGCCT GCATCCAGGG ACACACCACG TGGGTACCAA CATGTCCGGA GCCACATGGA GTCCACGGA TCTCATCGGA TCTCATCGGA TCTCATCGGA TCTCATCGGA TCTCATCGGA CGTAGGTCCC TGTGTGGTGC ACCCATGGTT GTACAGGCCT CGGTGTACCT	2310 2380 CCTCTGCCCT GAGAGTGACC GGAGACGGGA CTCTCACTGG	CACCCTGCCC CCATCCCGGG ATGAGCTGAC CAAGAACCAG GTCAGCCTGA CCTGCCTGGT CAAAGGCTTC GTGGGACGG GGTAGGGCCT GATCCGAAG GTTCTTGGTC CAGTCGGACT GGACGGACCA GTTTCCGAAG	2550 AGCCGGAGAA TCGGCCTCTT	2620 2670 2680 2680 2680 2680 2680 2680 2680 268
1820 GGGAGGGAGG CCCTCCCTCC	1910 GCCCCGTCTG CGGGCAGAC	2000 GGCACAGGCT CCGTGTCCGA	2080 2090 CGGGAGGACC CTGCCCCTGA GCCCTCCTGG GACGGGGACT	2180 ATCTTCTCTC TAGAAGAGAG		2350 2360 CAGAGGCCGG CTCGGCCCAC GTCTCCGGCC GAGCCGGGTG	2450 CCATCCCGGG GGTAGGGCCC	2530 2540 GGAGTGGGAG AGCAATGGGC CCTCACCCTC TCGTTACCCG	2630 GACAAGAGCA CTGTTCTCGT
1810	1900	1990	2080	2170	2260 2270	2350	2440	2530 2540	2620
GGCCAGCACA	AGCAAGGCAG	GCTCTGGGCA	CGGGAGGACC	GTAACTCCCA	CCCTCCAGGT CAAGGCGGGA	CAGAGGCCGG	CACCCTGCCC	GCAGTGGGAG AGCAATGGGC	GCTCACCGTG
CCGGTCGTGT	TCGTTCCGTC	CGAGACCCGT	GCCCTCCTGG	CATTGAGGGT	GGGAGGTCGA GTTCCGCCCT	GTCTCCGGCC	GTGGGACGGG	CCTCACCCTC TCGTTACCCG	CGAGTGGCAC

Figure 14 (continued)

2790	2880	2970	3060	3150	3240	3330	3420	3510	3600
TGCTTGGCAC	ATGGTTCTTT	TCTGCAGGTG	AGCAGCACCT	TTCTGTGAGC	CTACCCCCAC	CCTGTGGAGG	CACCACACAC	GAACACTCCT	TCAGACAAAC
ACGAACCGTG	TACCAAGAAA	ACACGTCCAC	TCGTCGTGGA	AAGACACTCG	GATGGGGGTG	GGACACCTCC	GTGGTGTGTG	CTTGTGAGGA	AGTCTGTTTG
2780	2870	2960	3050	3140	3230	3320	3410	3490 3500	3590
CGCACGAGGA	CGAGACTGTG	TGCCCAGGC	CCCTCCCTCC	GACTGTCCTG	CCTCACCCAT	ACTCTCGGGC	GCCACACGGC	AGCAAGGTCC TCGCACACGT	GCTGACCTGC
GCGTGCTCCT	GCTCTGACAC	ACCGGGTCCG	GGGAGGGAGG	CTGACAGGAC	GGAGTGGGTA	TGAGAGCCCG	CGGTGTGCCG	TCGTTCCAGG AGCGTGTGCA	CGACTGGACG
2770 CTCTCGCGGT GAGAGCGCCA	2840 2850 2860 2870 TAAAGCACCC AGCGCTGCCC TGGGCCCCTG CGAGACTGTG ATTTCGTGGG TCGCGACGGG ACCCGGGGAC GCTCTGACAC	2930 2940 2950 2960 2970 CAGCCCAGGC TGGCCCAGGC TGTGCAGGTG CTCCGTCTC TGGCCTGTG TGGCGTGTG ACCGGGTCCG ACGGTCCACA	3040 GCCAGCGTGG CGGTCGCACC	3130 CTCTGTAGGA GAGACATCCT	3220 ACAGGCCCTC TGTCCGGGAG	3310 GGGGACATGC CCCCTGTACG	3400 3410 AGGTTGGCCG GCCACACGGC TCCAACCGGC CGGTGTGCCG	3490 AGCAAGGTCC TCGTTCCAGG	3580 TTCTCCACAT AAGAGGTGTA
2760	2850	2940	3030	3120	3210	3300	3390	3480	3570
GCTCCCCGGG	AGCGCTGCCC	GGGTCCCACT	TGGGGGATTT	CAGCCCCTGC	GTGCGTAGGG	AACCGACTCC	CCCCGCACTG	CCCAGACCAG	TCTCGGCAGC
CGAGGGGCCC	TCGCGACGGG	CCCAGGGTGA	ACCCCTAAA	GTCGGGGACG	CACGCATCCC	TTGGCTGAGG	GGGGCGTGAC	GGGTCTGGTC	AGAGCCGTCG
2750	2840	2930	3020	3110	3200	3290	3380	3470	3560
GCAAGCCCCC	TAAAGCACCC	GAGGCAGAGC	CTCGGCAGGG	GACAGACACA	CCTAGTCCAT	ATGGGGACAC	GTTCAACAAA	CTGCACAGCA	CCCACGAGCC
CGTTCGGGGG	ATTTCGTGGG	CTCCGTCTCG	GAGCCGTCCC	CTGTCTGTGT	GGATCAGGTA	TACCCCTGTG	CAAGTTGTTT	GACGTGTCGT	GGGTGCTCGG
2740	2820 2830 CCGGGCGCCC AGCATGGAAA GCCCGGGGG TCGTACCTTT	2920	3010	3100	3190	3280	3370	3460	3550
GCGACGGCCG		TGGCATGAGG	AGGGGCTGCC	AGCCCCTGGQ	CGGGGGCATG	TCGCACCCGC	GCCCAGACCC	CCCGGGCGAA	CACCTCAAGG
CGCTGCCGGC		ACCGTACTCC	TCCCCGACGG	TCGGGGACCC	GCCCCCOTAC	AGCGTGGGCG	CGGGTCTGGG	GGGCCCGCTT	GTGGAGTTCC
2710 2770 2780 2780 2790 2790 2790 2790 2790 2790 2790 279		2890 2950 2960 2970 2970 2970 2970 2940 2950 2960 2960 2970 CCACGGGTCA GGCCCACGC TGGCCCAGGC TGTGCAGGTG GGTGCCACT GTCCCACAC TGGCCAGGC TGTGCAGGTG GGTGCCAGT GTCCCAACAC TGGCGGTCTG ACGGGTCTCACACT CCAGGGTGT CCGGCTCTAGA CTCCGGATCTC ACCGTACTCC CTCCGTCTCG CCCAGGGTGA CAGGGGTGTG ACGGGTTCG ACGGGTTCCAACA	2990 3050 3060 TOCCTOGGCC CCCTAGGCTG GGGCTGCC AGGGGCTGCC CTCGGCAGGG TGGGGGATTT GCCAGCGTGG CCCTCCCTCC AGCAGCACCT ACGGACCCGG GGGATCCCAC CCCGAGGG TCCCCGACGG GAGCCGTCCC ACCCCCTAAA CGGTCGCACC GGGAGGGAGG TCGTCGTGGA	300 300 3000 3000 3000 3000 3000 3100 3120 3130 3140 3140 3150 3150 3140 3140 3150 3140 3140 3140 3140 3140 3140 3140 314	3160 3170 3180 3230 3200 3200 3210 3200 3230 3240 3230 3230 3230 3240 324	3250 3320 3320 3320 3320 3330 3390 3390 3300 3310 3320 3330 CTGTGGAGGCCATAACC CCTGGCTGCCCCCCCCCCCCC	3340 3350 3360 3370 3380 3380 3390 3400 3400 3400 3410 3420 GACTGGTGCA GATGCCCACA CACACACTO GCCCAGACCC GTTCAACAAA CCCCGCACTG AGGTTGGCC GCCACACGGC CACCACACAC CTGACCACGT CTACGGGTGT GTGTGTGAGT CGGGTCTGGG CAAGTTGTTT GGGGCGTGAC TCCAACCGGC CGGTGTGCCG GTGGTGTGTG	3430 3430 3440 3450 3460 3470 3480 3480 3590 3500 3510 ACACGICCAC GCCTCACACA CGGAGCCTCA CCGGGGGAA CTGCACAGGCA CCCAGACCAG AGCAAGGTCC TCGCACACGT GAACACTCCT TGTGCACGTG CGGAGTGTG GCCTCGGAGT GGGCCCGCTT GACGTGTCGT GGGTCTGGTC TCGTTCCAGG AGCGTGTGCA CTTGTGAGGA	3520 3530 3540 3550 3560 3660 COGACACAGA 3550 3500 3570 3580 3590 3600 3600 3580 3600 3600 3600 3600 3600 3600 3600 36
2720	2800 2810	2900	2990	3080	3170	3260	3350	3440	3530
CTGTCTCCGG	STACCCCCTG TACATACTTC	GGCCGAGTCT	CCCTAGGGTG	GGGCCACGGG	TCCCGACCTC	CCTGGCTGCC	GATGCCCACA	GCCTCACACA	CCCCCACGAG
GACAGAGGCC	CATGGGGGAC ATGTATGAAG	CCGGCTCAGA	GGGATCCCAC	CCCGGTGCCC	AGGCTGGAG	GGACCGACGG	CTACGGGTGT	CGGAGTGTGT	GGGGGTGCTC
2710	2800	2890	2980	3070	3160	3250	3340	3430	3520
GAGCCTCTCC	GTACCCCCTG	CCACGGGTCA	TGCCTGGGCC	GCCCTGGGCT	GCCCTGTCC	GGCACTAACC	GACTGGTGCA	ACACGTGCAC	CGGACACAGG
CTCGGAGAGG	CATGGGGGAC	GGTGCCCAGT	ACGGACCCGG	CGGGACCCGA	CGGGACAGG	CCGTGATTGG	CTGACCACGT	TGTGCACGTG	GCCTGTGTCC

Figure 14 (continued)

3690	3780	3870	3960	4050	4140	4230	4320	4410	4500
TGGCCCACTT	CCCGTGCCTT	CATTCTATTC	ATGGCTTCTG	GTTACGCGCA	CCTCTCAAAA	CCCAGTTCCG	AAGTAGTGAG	TCCTAGCGTG	ATTGGCAAGA
ACCGGGTGAA	GGGCACGGAA	GTAAGATAAG	TACCGAAGAC	CAATGCGCGT	GGAGAGTTTT	GGGTCAAGGC	TTCATCACTC	AGGATCGCAC	TAACCGTTCT
3680	3770	3860	1950	4040	4130	4220	4310	4400	4490
TCCCTGGCCC	TTGCCCCTCC	GAGTAGGTGT	GGTGGGCTCT	GGGTGTGGTG	GTTCGCCGGG	CCTAACTCCG	GCTATTCCAG	TTGACGGCAA	AAATATGGGG
AGGGACCGGG	AACGGGGAGG	CTCATCCACA	CCACCCGAGA	CCCACACCAC	CAAGCGGCCC	GGATTGAGGC	CGATAAGGTC	AACTGCCGTT	TTTATACCCC
3670	3760 3770	3850	3940	4030	4120	4210	4300	4390	4480
CCACGTCACG	CATCTGTTGT TTGCCCCTCC	CGCATTGTCT	CTGGGGATGC	TAAGCGCGGC	TTCTCGCCAC	CCATCCCGCC	CGGCCTCTGA	CGCGCCAAAC	CCGTGTCCCA
GGTGCAGTGC	GTAGACAACA AACGGGGAGG	GCGTAACAGA	GACCCCTACG	ATTCGCGCCG	AAGAGCGGTG	GGTAGGGCGG	GCCGGAGACT	GCGCGGTTTG	GGCACAGGGT
360	3750	3840	3930	4020	4110	4200	4290	4380	4470
GGATCACACA	AGTTGCCAGC	GAAATTGCAT	AGCAGGCATG	AGCGGCGCAT	TTCCCTTCCT	CTAACTCCGC	GAGGCCGCCT	GCTGCGATTT	TGCATCGTCG
CCTAGTGTGT	TCAACGGTCG	CTTTAACGTA	TCGTCCGTAC	TCGCCGCGTA	AAGGGAAGGA	GATTGAGGCG	CTCCGGCGGA	CGACGCTAAA	ACGTAGCAGC
3650	3740	3830	3920	4010	4100	4190	4280	4370	4460
CACACACAGG	TGTGCCTTCT	ATAAAATGAG	GGAAGACAAT	CGCGCCCTGT	TTTCGCTTTC	AGTCCCGCCC	TGCAGAGGCC	ACAGCTCAGG	ACCATTGAAC
GTGTGTGTCC	ACACGGAAGA	TATTTTACTC	CCTTCTGTTA	GCGCGGGACA	AAAGCGAAAG	TCAGGGCGGG	ACGTCTCCGG	TGTCGAGTCC	TGGTAACTTG
3610 3610 3620 3640 3650 3660 3660 3660 3680 3680 3680 3680 CCAGCCCCCC TCCCAGCC TGCCCCACTT GGCCCTCC TCGCCC TGCCCCACTT GGCCCACTT GGCGCACC AGGGACG ACGGGACG ACGGGGTGAA	3700 3750 3750 3770 3780 3780 3750 3750 3750 3770 3780 CCCAGIGCCG CCCTTCCTTC CCCGTGCCTT AGTTGCCAGC CATCTGTTGT TTGCCCTCC CCCGTGCCTT GGGGGGGGG GGGCAGGAGGAACGAACGGAAGGGAACGAAC	3190 3800 3800 3810 3820 3830 3830 3840 3850 3860 3860 3870 CCTTGACCCT GGAAGGGC ACTCCCACTG TCCTTTCCTA ATAAAATGAG GAAATTGCAT GGCATGTCT GAGTAGGTGT CATTCTATTC GGAACTGGGA CCTTCCACGG TGAGGGTGAC AGGAAAGGAT TATTTTACTC CTTTAACGTA GGGTAACAGA CTCATCCACA GTAAGATAAG	1980 1980 1990 1990 1970 1970 1930 1930 1950 1930 1950 1950 1960 1960 1960 1960 1960 1960 1960 196	3990 4000 4010 4020 4030 4040 4050 GGTGTGGTG GTTACGGCA CGGGGGGAT TAAGCGCGGC GGTGTGGTG GTTACGGCA CCGAGATCC CCATAGGGGT GCGCGGAACA TCGCCGCGTA ATTCGCGCCG CCCACACAC CAATGCGCGT	4060 4070 4080 4080 4090 4100 4110 4120 4130 4140 GCGTGACGC TACACTTGCC TACACTTGCC AGGCGCCCTAG CCTCTCAAAA CCGACTGGCG ATGTGAAAGG TCGCGGGATC GCGGGGAGGAAAAAAAAAA	4150 4200 4200 4220 4230 4230 A230 A230 A230 A230 A230 A230 A230 A	4240 4250 4250 4250 4270 4280 4290 4300 4310 4320 CCCATTCTC CCCCATGGC TGACTAATTT TITITATITA TGCAGAGGCC GAGGCCGCCT CGCCTCTGA GCTATTCCAG AAGTAGTGAGGGCGGAAAGAG CGGGGAACGA CGAGATTAAA AAAATAAAT ACGTCTCCGG CTCCGGCGGA GCCGGAGACT CGATAAGGTC TTCATCACCTC	4330 4350 4350 4360 4370 4380 4380 4400 4400 6AGGCTTTTT TGAGGGCTTTTGAGGGGTTGAGGGGTGAGGGGTGGGGT	4420 4430 4450 4450 4460 4470 4480 4500 4500 4500 4500 4500 4500 450
3630	3720	3810	3900	3990	4080	4170	4260	4350	4440
GTGCCCTGC	CAGGACGGAT	ACTCCCACTG	GACAGCAAGG	GGCTCTAGGG	AGCGCCCTAG	CTCAATTAGT	TGACTAATTT	GGCTTTTGCA	CCCGCTGCCA
CACGGGGACG	GTCCTGCCTA	TGAGGGTGAC	CTGTCGTTCC	CCGAGATCCC	TCGCGGGATC	GAGTTAATCA	ACTGATTAAA	CCGAAAACGT	GGGCGACGGT
3620	3710	3800	3890	3980	4070	4160	4250	4340	4430
TCTCACAAGG	CCCTTCCCTG	GGAAGGTGCC	GGTGGGGCAG	AACCAGCTGG	TACACTTGCC	AAGCATGCAT	GCCCCATGGC	TGGAGGCCTA	GGATTTTATC
AGAGTGTTCC	GGGAAGGGAC	CCTTCCACGG	CCACCCCGTC	TTGGTCGACC	ATGTGAACGG	TTCGTACGTA	CGGGGTACCG	ACCTCCGGAT	CCTAAAATAG
3610	3700	3790	3880	3970	4060	4150	4240	4330	4420
CCAGCCCTCC	CCCAGTGCCG	CCTTGACCCT	TGGGGGGTGG	AGGCGGAAAG	GCGTGACCGC	AAGGGAAAAA	CCCATTCTCC	GAGGCTTTTT	AAGGCTGGTA
GGTCGGGAGG	GGGTCACGGC	GGAACTGGGA	ACCCCCACC	TCCGCCTTTC	CGCACTGGCG	TTCCCTTTTTT	GGGTAAGAGG	CTCCGAAAAA	TTCCGACCAT

Figure 14 (continued)

4590	4680	4770	4860	4950	5040	5130	5220	5310	5400
AAACAGAATC	AGTAGAGAAC	GCAAGTAAAG	ACAAGGATCA	CTCTCTGAGG	GCTCCCCTCC	TGACATAATT	TAATTGTTTG	CAGAAGAAAT	AGGACTTTCC
TTTGTCTTAG	TCATCTTTG	CGTTCATTTC	TGTTCCTAGT	GAGAGTCC	CGAGGGGAGG	ACTGTATTAA	ATTAACAAAC	GTCTTCTTTA	TCCTGAAAGG
	4660 4670 4680 ACAGAATTAA TATAGTTCTC AGTAGAGAAC TGTCTTAATT ATATCAAGAG TCATCTTTG	4710 4720 4730 4740 4750 4760 4770 GCTCATTITC TIGCCAAAAG TITGGAIGAI GCCTIAAGAC TIAITGAACA ACCGGAATIG GCAAGIAAAG CGAGIAAAAG AACGGIITIC AAACCIACIA CGGAAITCIG AAIAACITGI TGGCCTIAAC GGITCATITC	4780 4850 4850 4860 TAGATAGTC GAGGCAGTT CTGTTTACCA GAAGCCATG AATCAACCAG GCCACCTTAG ACTCTTGTG ACAAGGATCA ACTGTTACCA ACAGGATCA ACTCTTAG CTCCGTCAA GACAAATGGT CTTTCGGTAC TTAGTTGGTC CGGTGGAATC TGAGAAACAC TGTTCCTAGT	4940 CCCAGGCGTC GGGTCCGCAG	TIGAAGICTA CGAGAAGAAA GACTAACAGG AAGATGCTTT CAAGITCTCT GCTCCCCTCC AACITCAGAI GCICITCTIT CIGAITGTCC TICIAGGAAA GTICAAGAGA CGAGGGGAGG	SOBO 5090 5110 5120 5130 ACTITIVECTG GCTITIAGATC TCTTTGTGAA GGAACCTTAC TTCTGTGGTG TGACATAATT TGAAAACGAC CGAAATCTAG AGAAACACTT CCTTGGAATG AAGACACCAC ACTGTATAAA	5200 5210 5220 TAAGTCTATA ATGTGTTAAA CTACTGATTC TAATTGTTTG ATTCACATAT TACACAATTT GATGACTAAG ATTAACAAAC	5300 CTGTTTTGCT GACAAAACGA	5340 5350 5360 5370 5380 5400 CTACTGCTG CAAAAAGAA GAGAAAGGTA GAAGACCCCA AGGACTTTCC GATGACGTGA CTCTCGAACA CTCTTTCC GATGACGAC TTTTTCTT CTCTTTCCAT CTCTTTGGGGT TCCTGAAAGG
4570	4660	4750	4840	4910	5020	\$110	5200	5290	5380
CAACCTCTTC	ACAGAATTAA	TTATTGAACA	GCCACCTTAG	TCCCAGAATA	AAGATGCTTT	TCTTTGTGAA GGAACCTTAC	ATGTGTTAAA	TGAGGAAAAC	GAGAAAGGTA
GTTGGAGAAG	TGTCTTAATT	AATAACTTGT	CGGTGGAATC	AGGGTCTTAT	TTCTACGAAA	AGAAACACTT CCTTGGAATG	TACACAATTT	ACTCCTTTTG	CTCTTTCCAT
4560	4640 4650	4740	4830	4920	5010	5100	5190	5280	5370
AGAATGACCA	GAAGAATCGA CCTTTAAAGG	GCCTTAAGAC	AATCAACCAG	TATAAACTTC	GACTAACAGG	TCTTTGTGAA	TAAGTGTATA	ATGCCTTTAA	CAAAAAAGAA
TCTTACTGGT	CTTCTTAGCT GGAAATTTCC	CGGAATTCTG	TTAGTTGGTC	ATATTTGAAG	CTGATTGTCC	AGAAACACTT	ATTCACATAT	TACGGAAATT	GITITITITITIT
4550	4640	4730	4820	4910	5000	5090	5180	5270	5360
GTACTTCCAA	GAAGAATCGA	TTTGGATGAT	GGAAGCCATG	TTTGGGGAAA	CGAGAAGAAA	GCTTTAGATC	ATAAAATTTT	CAGTGGTGGA	TCTACTCCTC
CATGAAGGTT	CTTCTTAGCT	AAACCTACTA	CCPTCGGTAC	AAACCCCTTT	GCTCTTCTTT	CGAAATCTAG	TATTTAAAA	GTCACCACCT	AGATGAGGAG
CCGCTCAGGA ACGAGTTCAA GTACTTCCAA AGAATGACCA CAACCTCTTC AGTGGAAGGT GGCGAGTCCT TGCTCAAGGTT TCTTACTGGT GTTGGAGAAG TCACCTTCCA	4630 CCATTCCTGA GGTAAGGACT	4720 4730 4740 4750 4750 4760 THICCAAAAG THICGAATIGA GCCTTAAGAC THATIGAACA ACCGGAATIGA AACGGTTITIC AAACCTACTA CGGAATICIG AATAACTIGT TGGCCTTAAC	4810 CTGTTTACCA GACAAATGGT	4810 4820 4920 4930 4930 4930 4930 4930 A920 A930 TGCAGGAATT TGAAAGTGA ACGTCTTTACC CAGAAATTGA 1TTGGGGAAA TATAAACTTC TCCAGAATA ACGTCCTTA ACTTTCACTG TGCAAAAAGG GTCTTTAACT AAACCCCTTT ATATTTGAAG AGGGTCTTAT	TTGAAGTCTA CGAGAAGAAA GACTAACAGG AACTTCAGAT GCTCTTCTTT CTGATTGTCC		5160 5170 5180 TITIAAAGCTC TAAGGTAAAT ATAAAATTT AAAITTCGAG ATTCCATTTA TAITITTAAAA	5230 5230 5240 5250. 5260 5290 5290 5290 5290 TGTATTTAG ATTCCAACCT ATGGAACTGA TGAATGGGAG CAGTGGTGGA ATGCCTTTAA TGAGGAAAAC ACATAAAAATC TAAGGTTGGA TACCTTGACT ACTTACCCTC GTCACCACCT TACGGAAATT ACTCCTTTTG	5340 5350 5350 CTACTGCTGA CTACTGCTGA CTCTCAACAT TCTACTGCTGA GATGATGTA AGATGAGAGAGAGAGAGAGAGAGAGA
4530	4600 4610 4620 4630 4630 A630 A630 A630 A630 ACCACHAATA GGCTAGGAAA ACCTGGTTCT CCATTCCTGA ACCACTAATA CCCATCCTTT TGGACCAAGA GGTAAGGACT	4710	4800	4890	4980	5070	5160	5250.	5340
CCGCTCAGGA		GCTCATTTTC	GGAGGCAGTT	ACGITITITCC	AAGTATAAGT	AGACCATGGG	TTTAAAGCTC	ATGGAACTGA	CTACTGCTGA
GGCGAGTCCT		CGAGTAAAAG	CCTCCGTCAA	TGCAAAAAGG	TYCATATYCA	TCTGGTACCC	AAATTTCGAG	TACCTTGACT	GATGACGACT
4520	4600 4610	4700	4780 4790	4810	4960	5050	5140 5150	5240	5320 5330
ACGGAGACCT ACCCTGGCCT	TGGTGATTAT GGGTAGGAAA	ACCACGAGGA	TAGACATGGT TTGGATAGTC	TCCAGGAATT TGAAAGTGAC	TCCAGGAGGA AAAAGGCATC	TAAAGCTATG CATTTTATA	GGACAACTA CCTACAGAGA	ATTCCAACCT	GCCATCTAGT GATGATGAGG
TGCCTCTGGA TGGGACCGGA	ACCACTAATA CCCATCCTTT	TGGTGCTCCT	ATCTGTACCA AACCTATCAG	ACGTCCTTAA ACTTTCACTG	AGGTCCTCT TTTTCCGTAG	ATTTCGATAC GTAAAAATAT	CCTGTTTGAT GGATGTCTCT	TAAGGTTGGA	CGGTAGATCA CTACTACTCC
4510	4600	4690	4780	4870	4960	5050	5140	5230	5320
ACGGAGACCT	TGGTGATTAT	TCAAAGAACC	TAGACATGGT	TGCAGGAATT	TCCAGGAGGA	TAAAGCTATG	GGACAAACTA	TGTATTTTAG	GCCATCTAGT
TGCCTCTGGA	ACCACTAATA	AGTTTCTTGG	ATCTGTACCA	ACGTCCTTAA	AGGTCCTCCT	ATTTCGATAC	CCTGTTTGAT	ACATAAAATC	CGGTAGATCA

Figure 14 (continued)



5490 AAAAGCTGC TTTTTCGACG	5580 TTTTTCTTAC AAAAGAATG	S670 AGGA	S760 SACC STGG	5850 NGCA NCGT	o ပ ပ	046	000	၀ ပ ပ	000
AAAAA	₹ ₹	5670 1TAATAAGGA AATTATTCCT	5760 CTCCCACACC GAGGGTGTGG	58 AGCAATAGG TCGTTATC	5940 GTCTCGATCG CAGACCTAGC	6030 TACAAATAAA ATGTTTATTT	6120 TCTTATCATG AGAATAGTAC	6210 ACAATTCCAC TGTTAAGGTG	6300 TCACTGCCCG AGTGACGGGC
5480	5570	5660	5750	5840	5930	6020	6110	6200	6290
ACCACAAAGG	AACATACTGT	TCTAAAGGGG	TTTAAAAAAC	TTACAAATAA	ATCTTATCAT	TTATAATGGT	CATCAATGTA	TTATCCGCTC	TGCGTTGCGC
TGGTGTTTCC	TTGTATGACA	ACATTTCCCC	AAATTTTTTG	AATGTTTATT	TAGAATAGTA	AATATTACCA	GTAGTTACAT	AATAGGCGAG	ACGCAACGCG
5410 5420 5430 5430 5430 5480 5480 5480 5480 5480 5480 5480 548	5500 5510 5520 5530 5540 5540 5550 5550 5510 5580 5500 5560 5570 5580 5560 5500 5500 5580 5580 5580 558	5590 5600 5610 5620 5630 5640 5650 5650 5670 TCACACACAGA CATATTAATT TGTAAAGGG TTAATAAGGA AGGTGTCCC GAATATTACCC GAATCTCCC GAATATTCCC AATTATTCCC AATTATTCCC	5680 5740 5750 5720 5730 5730 5740 5750 5740 5740 5740 5740 5740 574	5770 5780 5880 5880 5880 5880 5880 5880	5860 5810 5820 5930 5940 5900 5910 5920 5930 5940 TCACAAATIT CACTGCAITC TAGTIGIGGI TIGICCAAAC TCATCAATGI ATCTTATCAT GTCTGGATCG AGTGTITAAA GGATTITITI CACTGCAIAG ATCAACACCA AACAGGITIG AGTAGTIACA TAGAATAGTA CAGACCTAGG	5950 5950 6000 6010 6020 6030 6030 6030 6010 6030 6030 6030 603	6040 6050 6050 6060 6070 6080 6090 6100 6110 6120 GCAATAGCAT CACAAAȚITC ACAAATATAC CATTITITIC ACTGCATICT AGTICIGGIT IGICCAAACT CATCAATGIA ICITAATGAT CGȚIAICGIA GIGITIAAAG IGIITAATITIC GIAAAAAAAG IGACGIAAGA ICAACACCAA ACAGGITIGA GIAGITACAT AGAATAGIAC	6130 6140 6150 6150 6160 6170 6180 6190 6290 TCTGAAATTG TGTGAAATTG TTATCCGCTC AGACATATG CTGTATGGAAATTG TTATCCGCTC AGACATATGG CAGCAGGAGA ACCGCATTAG TACCAGGATC GACAAAGGAC ACACTTTAAC AATAGGCGAG	6270 6280 6280 6390 6300 GIGAGCIAAC TCACTGCCCG CACTCGATTG AGTGTAATTA ACGCAACGCG AGTGACGGGC
5460	5550	5640	5730	5820	5910	6000	6090	6180	6270
TIGCTIGCTT	GGCATAACAG	GTACCTTTAG	TTTGTAGAGG	TTTATTGCAG	TTGTCCAAAC	CCCAACTIGI	AGTTGTGGTT	CTGTTTCCTG	GTGAGCTAAC
AACGAACGAA	CCGTATTGTC	CATGGAAATC	AAACATCTCC	AAATAACGTC	AACAGGTTTG	GGGTIGAACA	TCAACACCAA	GACAAAGGAC	CACTCGATTG
5450	5540	5630	5720	5810	5900	5990	6080	6170	6260
AATAGAACTC	TTTATAAGTA	CAAAAATTGT	CCATACCACA	TGTTAACTTG	TAGTTGTGGT	CTTCGCCCAC	ACTGCATTCT	ATGGTCATAG	TGCCTAATGA
TTATCTTGAG	AAATATTCAT	GTTTTTAACA	GGTATGGTGT	ACAATTGAAC	ATCAACACCA	GAAGCGGGTG	TGACGTAAGA	TACCAGTATC	ACGGATTACT
5440 TGTGTTTTAGT ACACAAATCA	5530 TTCTGTAACC AAGACATTGG	5620 TAACTATGCT ATTGATACGA	5680 5720 5730 5730 5730 5730 5730 5730 5730 573	5800 CAATIGITGE GITAACAACA	5890 CACTGCATTC GTGACGTAAG	5980 TGCTGGAGTT ACGACCTCAA	6070 CATTITITIC GTAAAAAAAG	6160 TGGCGTAATC ACCGCATTAG	ACAACATACG AGCCGGAAGC ATAAAGTGTA AAGCCTGGGG TGCCTAATGA TGTTGTATGTAGACCCC ACGCATTACT
5420 5430	5520	5610	5700	5770 S780 S790 TCCCCCTGAA CCTGAAACAT AAAATGAATGAAGGGGGACTT GGACTTTOTA TTTTACTTAC	S880	5970	6040 6050 6050 6060	6150	6240
CTAAGTTTT TGAGTCATGC	TGGAAAAATA	CTGCTATTAA	TGACTAGAGA		GCATITITITI	GGGGATCTCA	GCAATAGCAT CACAAATITC ACAAATAAAG	AGCTAGAGCT	ATAAAGTGTA
GATTCAAAAA ACTCAGTACG	ACCTTTTTTAT	GACGATAATT	ACTGATCTCT		CGTAAAAAA	CCCCTAGAGT	CGITAITCGTA GTGTFTAAAG TGTFTAATITC	TCGATCTCGA	TATTTCACAT
S420	5510	5600	5690	5780	5870	5960	6050	6140	6230
CTAAGTTTTT	AAGAAAATTA	CATAGAGTGT	TATAGTGCCT	CCTGAAACAT	CACAAATAAA	CCTCCAGCGC	CACAAATTTC	GTCGACCTCT	AGCCGGAAGC
GATTCAAAAA	TICTTTTAAT	GTATCTCACA	ATATCACGGA	GGACTTTGTA	GTGTTTAATTT	GGAGGTCGCG	GTGTTTAAAG	CAGCTGGAGA	TCGGCCTTCG
5410	5500	5590	5680	5770	5860	5950	6040	6130	6220
TTCAGAATTG	ACTGCTATAC	TCCACACAGO	ATATITGATG	TCCCCCTGAA	TCACAAATTT	GCTGGATGAT	GCAATAGCAT	TCTGTATACC	ACAACATACG
AAGTCTTAAC	TGACGATATG	AGGTGTGTCC	TATAAACTAC	AGGGGACTT	AGTGTTTAAA	CGACCTACTA	CGTTATCGTA	AGACATATGG	TGTTGTATGC

Figure 14 (continued)



CCCTCTTCCG	6480 TTATCCACAG AATAGGTGTC	6570 GCGTTTTTCC CGCAAAAAGG	6660 TACCAGGCGT ATGGTCCGCA	6750 GGAAGCGTGG CCTTCGCACC	6840 CCCGTTCAGC GGCCAAGTCG	6930 ACTGGTAACA TGACCATTGT	7020 GTATTTGGTA CATAAACCAT	7110 GGTGGTTTTT CCACCAAAA	7200 CAGTGGAACG GTCACCTTGC
6370 6380 6370 6380 GAGGCGGTTTCCG CTCTTCCG CTCCTCCGCCANA CGCATAACCC GCGAGAAGGC	6460 6470 ACTCAAAGGC GGTAATACGG TGAGTTTCCG CCATTATGCC	6540 6550 6550 6560 GCCAGGAACC GTAAAAAGGC CGCGTTGCTG CGGTCCTTGG CATTTETCCG GCGCAACGAC	6640 6650 6650 ACCCGACGCGT TACCAGGCGT TGGGCTGTCC TGATATTCT ATGGTCCGCA	6700 6710 6720 6730 6750 6750 CTGTTCCGAC CCTGCCGTT ACCGGATACC TGTCCGCCTT TCTCCCTTTCG GGAAGCGTGG GACAGGGGAA TGGCCTATGG ACAGGCGGAA AGAGGGAAGC CCTTCGCACC	6800 6810 6820 6820 6830 GTAGGTCGTT CGCTCCAAGC TGGGCTGTGT GCACGAACCC CATCCAGCAA GCGAGGTTCG ACCCGACAACA CGTGCTTGGG	CAACCCGGTA AGACACGACT TATCGCCACT GGCAGCAGCC ACTGGTAACA GTTGGGCCAT TCTGTGGTGA ATAGCGGTGA CCGTCGTCGG TGACCATTGT	6980 7020 7000 7010 7020 CTTGAAGTGG TGGCCTAACT ACGCCTACAC TAGAAGGACA GTATTTGGTA GAACTTCACC ACCGATTGA TGCCGATGTG ATCTTCCTGT CATAAACCAT	7050 7100 7100 7010 7050 AACHANGGAG GGTGGTAGC GGTGGTTTTT GALANAGAG GGTGGTAGC GGTGGTTTTTT CTTTTTCTCA ACCATGGGCGGT TTGTTTGGTG GGGGCGTCG CCACCAAAAA	AAAAAGGATC TCAAGAAGAT CCTTTGATCT TTTCTACGGG GTCTGACGCT CAGTGGAACG TTTTTTCCTAG AGTTCTTCTA GGAAACTAGA AAAGATGCCC CAGACTGCGA GTCACCTTGC
	6460 ACTCAAAGGC TGAGTTTCCG	6530 6540 6550 6550 6560 CCAGCAAAAA GC CGCGTTGCTG GGTCGTTTTC CGGTCCTTGG CATTTTTCCG GCGCAACGAC		6730 TGTCCGCCTT ACAGGCGGAA	6810 6820 6830 CGCTCCAAGC TGGGCTGTGT GCACGAACCC GCGAGGTTCG ACCCGACACA CGTGCTTGGG	6900 6910 AGACACGACT TATCGCCACT TCTGTGCTGA ATAGCGGTGA	7000 ACGCCTACAC TGCCGATGTG	TOTO 7080 7080 7090 TGGTAGCTCT TGATCCGGCA AACAAACCAC ACCATCGAGA ACTAGGCCGT TTGTTTGGTG	7180 TTTCTACGGG AAAGATGCCC
6360 CGCGCGGGGA GCGCGCCCT	6450 GTATCAGCTC CATAGTCGAG	6540 GCCAGGAACC CGGTCCTTGG	6620 6630 CTCAAGTCAG AGGTGGCGAA GAGTTCAGTC TCCACCGCTT	CTGTTCCGAC CCTGCCGCTT ACCGGATACC GACAAGGCTG. GGACGGCGAA TGGCCTATGG	6810 CGCTCCAAGC GCGAGGTTCG	6900 AGACACGACT TCTGTGCTGA	6990 TGGCCTAACT ACCGGATTGA	7080 TGATCCGGCA ACTAGGCCGT	7170 CCTTTGATCT GGAAACTAGA
6350 AATCGGCCAA TTAGCCGGTT	6440 GCGGCGAGCG CGCCGCTCGC	6530 CCAGCAAAAG GGTCGTTFTTC		6710 CCTGCCGCTT GGACGCCGAA			6980 CTTGAAGTGG GAACTTCACC	7070 TGGTAGCTCT ACCATCGAGA	7160 TCAAGAAGAT AGTTCTTCTA
6340 TGCATTAATG ACGTAATTAC	6430 TCGTTCGGCT AGCAAGCCGA	6520 GAGCAAAAGG CTCGTTTTCC	6610 AAAATCGACG TTTTAGCTGC	6700 CTGTTCCGAC GACAAGGCTG.	6790 TCAGTTCGGT AGTCAAGCCA	6880 GTCTTGAGTC CAGAACTCAG	.6970 CTACAGAGIT GAIGICICAA	7060 GAAAAAGAGT CTTTTTTCTCA	7150 AAAAAGGATC TTTTTCCTAG
6330 TCGTGCCAGC AGCACGGTCG	6410 6410 6410 6420 6430 6430 6440 6450 6450 6460 6460 6470 CTYCCTCGCT CACTGACTCG CTGCGCTCGG TCGTTCGGCT GCGCGAGCG GTATCAGCTC ACTCAAAGGC GGTAATACGG	6500 6510 6520 NACGCAGGA AAGAACATGT GAGCAAAAGG FTGCGTCCT TTCTTGTACA CTCGTTTTTCC	6600 6610 GAGCATCACA AAAATCGACG CTCGTAGTGT TTTTAGCTGC	6690 OTGCGCTCTC CACGCGAGAG	6770 6780 ATGCTCACGC TGTAGGTATC TACGAGTGCG ACATCCATAG	6870 GGTAACTATC CCATTGATAG	6950 6950 AGCGAGGTAT GTAGGCGGTG TCGCTCATA CATCGCCAC	7040 7050 SCTGAAGCCA GTTACCTTCG	7140 7150 SCAGCAGATT ACGCOCAGAA AAAAAGGATC CGTCGTCTAA TGCGCGTCTT TTTTTCCTAG
6310 6320 6330 6330 CTTICCAGIC GGAAACCTIG CCTITICGAC ACCAGGICG	6410 CACTGACTCG GTGACTGAGC	6500 TAACGCAGGA ATTGCGTCCT	6590 CCCCCCTGAC GGGGGACTG						
6310 CTTTCCAGTC GAAAGGTCAG	6400 CTTCCTCGCT GAAGGAGCGA	6490 AATCAGGGGA T TTAGTCCCCT A	6580 ATAGGCTCCG TATCCGAGGC	6670 TTCCCCCTGG	6760 CGCTTTCTCA	6850 CCGACCGCTG	6940 GGATTAGCAG	7030 TCTGCGCTCT	7120 TIGITIGCAA AACAAACGITI

Figure 14 (continued)



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7290	7380	7470	7560	7650	7740	. 7830	7920	8010	8100
TYTTAAATCAA	TITICGTICAT	CCGCGAGACC	TCCGCCTCCA	ACAGGCATCG	TTGTGCAAAA	CTGCATAATT	CGGCGACCGA	TCTTCGGGGC	TYTACTTYCA
AAATYTTAGTY	AAAGCAAGTA	GGCGCTCTGG	AGGCGGAGGT	TGTCCGTAGC	AACACGTTTT	GACGTATTAA	GCCGCTGGCT	AGAAGCCCCG	AAATGAAAGT
7280 AAAATGAAGT TTTTACTTCA	7330 734CGTCTA TTTCGTTCAT CTAGACAGAT AAAGCAAGTA	7450 7450 GCCCCAGTGC TGCAATGATA CGGGGTCACG ACGTTACTAT	7550 TGCAACTTTA ACGTTGAAAT	7640 TGCCATTGCT ACGGTAACGA	7730 7730 GAGTTACATG ATCCCCCATG CTCAATGTAC TAGGGGGTAC	7820 TATGGCAGCA ATACCGTCGT	7910 ATAGTGTATG TATCACATAC	8000 TGGAAAACGT ACCTTTTGCA	8080 8090 CCAACTGATC TTCAGCATCT GGTTGACTAG AAGTCGTAGA
7270 TTTTAAATTA AAAATTTAAT	7360 CTATCTCAGC GATAGAGTCG		7540 GAAGTGGTCC CTTCACCAGG	7630 GCAACGTTGT CGTTGCAACA		7810 CACTCATGGT GTGAGTACCA	7890 7900 7910 TCAACCAAGT CATTCTGAGA ATAGTGTATG AGTTGGTTCA GTAAGACTCT TATCACATAC	7990 TGCTCATCAT ACGAGTAGTA	8080 CCAACTGATC GGTTGACTAG
7250 7260	7350	ACCCACCC TTACCATCTC TGCCCTCCCC AATGGTAGAC	7530	7620	7710	7800	7890	7980	8070
AAGGATCTTC ACCTAGATCC	AGTGAGGCAC		GCCGAGCGCA	AATAGTTYGC	CGATCAAGGC	GCAGTGTTAT	TCAACCAAGT	ACTTTAAAAG	ACTCGTGCAC
TTCCTAGAAG TGGATCTAGG	TCACTCCGTG		CGGCTCGCGT	TYATCAAACG	GCTAGTTCCG	CGTCACAATA	AGTTGGTTCA	TGAAATTTTC	TGAGCACGTG
7250	7340	7430	7520	7610	7700	7790	7880	7970	8060
AAGGATCTTC	ATGCTTAATC	ACGGGAGGGC	AGCCGGAAGG	TTCGCCAGTT	CGGTTCCCAA	TAAGTTGGCC	TGGTGAGTAC	ACATAGCAGA	GATGTAACCC
TTCCTAGAAG	TACGAATTAG	TGCCCTCCCG	TCGGCCTTCC	AAGCGGTCAA	GCCAAGGGTT	ATTCAACCGG	ACCACTCATG	TGTATCGTCT	CTACATTGGG
7240 GATTATCAAA CTAATAGTTT	ACTIGGICIG ACAGITACCA AIGCITAATC AGIGAGGCAC CTATCTCAGC TGAACCAGAC TGICAATGGT TACGAATTAG TCACTCCGTG GATAGAGTCG	7420 TAACTACGAT ATTGATGCTA	7510 TAAACCAGCC ATTTGGTCGG	7600 GAGTAAGTAG CTCATTCATC	7690 CATTCAGCTC GTAAGTCGAG	7780 TTGTCAGAAG AACAGTCTTC	7870 TTTCTGTGAC AAAGACACTG		8050 GATCCAGFIC CTAGGTCAAG
7230 TTGGTCATGA AACCAGTACT	7320 ACTTGGTCTG TGAACCAGAC	7410 7420 GTCGTGTAGA TAACTACGAT CAGCACATCT ATTGATGCTA	7490 7500 GGCTCCAGAT TTATCAGCAA CCGAGGTCTA AATAGTCGTT	7590 CGGGAAGCTA GCCCTTCGAT	GGTATGGCTT CATTCAGCTC CCATACCGAA GTAAGTCGAG	7770 CCTCCGATCG GGAGGCTAGC	7860 7870 GTAAGATGCT TTTCTGTGAC CATTCTACGA AAAGACACTG	7950 7960 ATACGGGATA ATACCGCGCC TATGCCCTAT TATGGCGGG	8040 CCGCTGTTGA GCCGACAACT
7220	7310	7400	7490	7580	7670	7760	7850	7940	8030
TTAAGGGATT	ATATGAGTAA	CTGACTCCCC	GGCTCCAGAT	TAATTGTTGC	CTCGTCGTTT	CTCCTTCGGT	CATGCCATCC	CCCGGCGTCA	AAGGATCTTA
AATTCCCTAA	TATACTCATT	GACTGAGGGG	CCGAGGTCTA	ATTAACAACG	GAGCAGCAAA	GAGGAAGCCA	GTACGGTAGG	GGGCCGCAGT	TTCCTAGAAT
7210	7300	7390	7480	7570	7660	7750	7840	7930	8020
AAAACTCACG	TCTAAAGTAT	CCATAGTTGC	CACGCTCACC	TCCAGTCTAT	TGGTGTCACG	AAGCGGTTAG	CTCTTACTGT	GTTGCTCTTG	GAAAACTCTC
TTTTGAGTGC	AGATTTCATA	GGTATCAACG	GTGCGAGTGG	AGGTCAGATA	ACCACAGTGC	TTCGCCAATC	GAGAATGACA	CAACGAGAAC	CTTTTGAGAG

Figure 14 (continued)

8280 AAACAAATAG TTTGTTTATC	
8270 TTAGAAAAT AATCTTTTTA	
8260 TTGAATGTAT AACTTACATA	
8250 GGATACATAT CCTATGTATA	
8240 TCTCATGAGC AGAGTACTCG	8330 G
8230 AGGGTTATTG TCCCAATAAC	9320 8310 8320 8320 3GGTTCCGCG CACCTGACGT C
8220 AGCATTTATC TCGTAAATAG	B310 CGAAAAGTGC CJ GCTTTTCACG GJ
8210 ATATTATTGA TATAATAACT	8300 CACATTYCCC CV GTGTAAAGGG GV
8200 TCCTTTTTCA AGGAAAAGT	8290 GGGTTCCGCG CCCAAGGCGC
	8210 8220 8270 ATAITATICA AGGATTATIG TCTCATGAGC GGATACATAT TIGAATGTAT TIAGAAAAAT AAACA TAIAAATAACT TCGTAAAATAG TCCCAATAAC AGAGTACTCG CCTATGTATA AACTTACATA AATCTITITIA TITGT

Figure 14 (continued)

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Comparison of whole chiBR96 and deleted CH2 chiBR96 on Ley/K ELISA

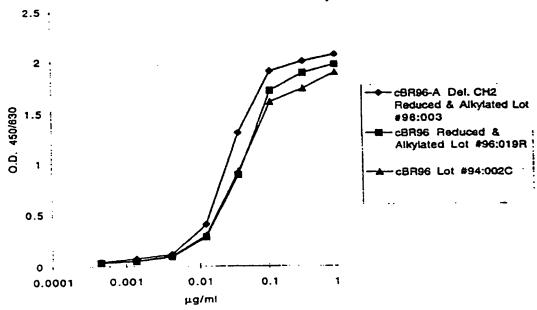


Figure 15

hBR96-2B: L235 to A235 and G237 to A237

hBR96-2C: E318 to S318, K320 to S320, and K322 to S322

hBR96-2D: P331 to A331

hBR96-2E: L235 to A235, G237 to A237, E318 to S318, K320 to S320, and

K322 to S322

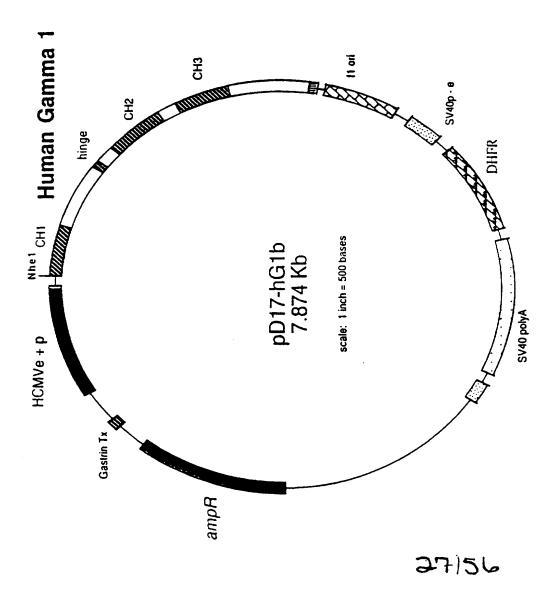
hBR96-2F: L235 to A235, G237 to A237, and P331 to A331

hBR96-2G: E318 to S318, K320 to S320, K322 to S322, and P331 to A331

hBR96-2H: L235 to A235, G237 to A237, E318 to S318, K320 to S320, K322 to

S322, and P331 to A331

Figure 16



BNSDOCID: <WO 9805787A1>

FIGURE 17

FIGURE 18A

1	GGTACCAATT	TAAATTGATA	TCTCCTTAGG	TCTCGAGTCT	CTAGATAACC
51	GGTCAATCGA	TTGGAATTCT	TGCGGCCGCT	TGCTAGCCAC	CATGGAGTTG
101	TGGTTAAGCT	TGGTCTTCCT	TGTCCTTGTT	TTAAAAGGTG	TCCAGTGTGA
151	AGTGCAACTG	GTGGAGTCTG	GGGGAGGCTT	AGTGCAGCCT	GGAGGGTCCC
201	TGCGACTTTC	CTGTGCTGCA	TCTGGATTCC	CGTTCAGTGA	CTATTACATG
251	TATTGGGTTC	GCCAGGCTCC	AGGCAAGGGA	CTGGAGTGGG	TCTCATACAT
301	TAGTCAAGAT	GGTGATATAA	CCGACTATGC	AGACTCCGTA	AAGGGTCGAT
351	TCACCATCTC	CAGAGACAAT	GCAAAGAACA	GCCTGTACCT	GCAAATGAAC
401	AGCCTGAGGG	ACGAGGACAC	AGCCGTGTAT	TACTGTGCAA	GAGGCCTGGC
451	GGACGGGGCC	TGGTTTGCTT	ACTGGGGCCA	AGGGACTCTG	GTCACGGTCT
501	CTTCCGCTAG	CACCAAGGGC	CCATCGGTCT	TCCCCCTGGC	ACCCTCCTCC
551	AAGAGCACCT	CTGGGGGCAC	AGCGGCCCTG	GGCTGCCTGG	TCAAGGACTA
601	CTTCCCCGAA	CCGGTGACGG	TGTCGTGGAA	CTCAGGCGCC	CTGACCAGCG
551	GCGTGCACAC	CTTCCCGGCT	GTCCTACAGT	CCTCAGGACT	CTACTCCCTC
701	AGCAGCGTGG	TCACCGTGCC	CTCCAGCAGC	TTGGGCACCC	AGACCTACAT
751	CTGCAACGTG	AATCACAAGC	CCAGCAACAC	CAAGGTGGAC	AAGAAAGTTG
801	GTGAGAGGCC	AGCACAGGGA	GGGAGGGTGT	CTGCTGGAAG	CCAGGCTCAG
851	CGCTCCTGCC	TGGACGCATC	CCGGCTATGC	AGCCCCAGTC	CAGGGCAGCA
901	AGGCAGGCCC	CGTCTGCCTC	TTCACCCGGA	GGCCTCTGCC	CGCCCCACTC
951	ATGCTCAGGG	AGAGGGTCTT	CTGGCTTTTT	CCCCAGGCTC	TGGGCAGGCA
1001	CAGGCTAGGT	GCCCCTAACC	CAGGCCCTGC	ACACAAAGGG	GCAGGTGCTG
1051	GGCTCAGACC	TGCCAAGAGC	CATATCCGGG	AGGACCCTGC	CCCTGACCTA
1101	AGCCCACCCC	AAAGGCCAAA	CTCTCCACTC	CCTCAGCTCG	GACACCTTCT
1151	CTCCTCCCAG	ATTCCAGTAA	CTCCCAATCT	TCTCTCTGCA	GAGCCCAAAT
1201	CTTGTGACAA	AACTCACACA	TGCCCACCGT	GCCCAGGTAA	GCCAGCCCAG
1251	GCCTCGCCCT	CCAGCTCAAG	GCGGGACAGG	TGCCCTAGAG	TAGCCTGCAT
1301	CCAGGGACAG	GCCCCAGCCG	GGTGCTGACA	CGTCCACCTC	CATCTCTTCC

1351	TCAGCACCTG	AACTOTTGG	A 3 P GGGACCGTCA	GTCTTCCTCT	TCCCCCCAAA
1401	ACCCAAGGAC	ACCCTCATGA	TCTCCCGGAC	CCCTGAGGTC	ACATGCGTGG
1451	TGGTGGACGT	GAGCCACGAA	GACCCTGAGG	TCAAGTTCAA	CTGGTACGTG
1501	GACGGCGTGG	AGGTGCATAA	TGCCAAGACA	AAGCCGCGGG	AGGAGCAGTA
1551	CAACAGCACG	TACCGTGTGG	126	CACCGTCCTG	CACCAGGACT
1601		CAAGGAGTAC	AAGTGCHAGG	TCTCCAACAA	AGCCCTCCCA
1651	GCCCCATCG	AGAAAACCAT	CTCCAAAGCC	AAAGGTGGGA	CCCGTGGGGT
1701	GCGAGGGCCA	CATGGACAGA	GGCCGGCTCG	GCCCACCCTC	TGCCCTGAGA
1751	GTGACCGCTG	TACCAACCTC	TGTCCCTACA	GGGCAGCCCC	GAGAACCACA
1801	GGTGTACACC	CTGCCCCCAT	CCCGGGATGA	GCTGACCAAG	AACCAGGTCA
1851	GCCTGACCTG	CCTGGTCAAA	GĠCTTCTATC	CCAGCGACAT	CGCCGTGGAG
1901	TGGGAGAGCA	ATGGGCAGCC	GGAGAACAAC	TACAAGACCA	CGCCTCCCGT
1951	GCTGGACTCC	GACGGCTCCT	TCTTCCTCTA	CAGCAAGCTC	ACCGTGGACA
2001	AGAGCAGGTG	GCAGCAGGGG	AACGTCTTCT	CATGCTCCGT	GATGCATGAG
2051	GCTCTGCACA	ACCACTACAC	GCAGAAGAGC	CTCTCCCTGT	CTCCGGGTAA
2101	ATGAGTGCGA	CGGCCGGCAA	GCCCCCGCTC	CCCGGGCTCT	CGCGGTCGCA
2151	CGAGGATGCT	TGGCACGTAC	CCCCTGTACA	TACTTCCCGG	GCGCCCAGCA
2201	TGGAAATAAA	GCACCCAGCG	CTGCCCTGGG	CCCCTGCGAG	ACTGTGATGG
2251	TTCTTTCCAC	GGGTCAGGCC	GAGTCTGAGG	CCTGAGTGGC	ATGAGGGAGG
2301	CAGAGCGGGT	CCCACTGTCC	CCACACTGGC	CCAGGCTGTG	CAGGTGTGCC
2351	TGGGCCCCCT	AGGGTGGGC	TCAGCCAGGG	GCTGCCCTCG	GCAGGGTGGG
2401	GGATTTGCCA	GCGTGGCCCT	CCCTCCAGCA	GCACCTGCCC	TGGGCTGGGC
2451	CACGGGAAGC	CCTAGGAGCC	CCTGGGGACA	GACACACAGC	CCCTGCCTCT
2501	GTAGGAGACT	GTCCTGTTCT	GTGAGCGCCC	CTGTCCTCCC	GACCTCCATG
2551	CCCACTCGGG	GGCATGCCTA	GTCCATGTGC	GTAGGGACAG	GCCCTCCCTC
2601	ACCCATCTAC	CCCCACGGCA	CTAACCCCTG	GCTGCCCTGC	CCAGCCTCGC
2651	ACCCGCATGG	GGACACAACC	GACTCCGGGG	ACATGCACTC	TCGGGCCCTG
2701	TGGAGGGACT	GGTGCAGATG	CCCACACACA	CACTCAGCCC	AGACCCGTTC
2751	AACAAACCCC	GCACTGAGGT	TGGCCGGCCA	CACGGCCACC	ACACACACAC
2801	GTGCACGCCT	CACACACGGA	GCCTCACCCG	GGCGAACTGC	ACAGCACCCA
		_			29156

FIGURE 18B

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2851	GACCAGAGCA	AGG : CCTCGC	ACACGTGAAC	ACTCCTCGGA	CACAGGCCCC
2901	CACGAGCCCC	ACGCGGCACC	TCAAGGCCCA	CGAGCCTCTC	GGCAGCTTCT
2951	CCACATGCTG	ACCTGCTCAG	ACAAACCCAG	CCCTCCTCTC	ACAAGGGTGC
3001	CCCTGCAGCC	GCCACACACA	CACAGGGGAT	CACACACCAC	GTCACGTCCC
3051	TGGCCCTGGC	CCACTTCCCA	GTGCCGCCCT	TCCCTGCAGG	ACGGATCAGC
3101	CTCGACTGTG	CCTTCTAGTT	GCCAGCCATC	TGTTGTTTGC	CCCTCCCCCG
3151	TGCCTTCCTT	GACCCTGGAA	GGTGCCACTC	CCACTGTCCT	TTCCTAATAA
3201	AATGAGGAAA	TTGCATCGCA	TTGTCTGAGT	AGGTGTCATT	CTATTCTGGG
3251	GGGTGGGGTG	GGGCAGGACA	GCAAGGGGGA	GGATTGGGAA	GACAATAGCA
3301	GGCATGCTGG	GGATGCGGTG	GGCTCTATGG	CTTCTGAGGC	GGAAAGAACC
3351	AGCTGGGGCT	CTAGGGGGTA	TCCCCACGCG	CCCTGTAGCG	GCGCATTAAG
3401	CGCGGCGGGT	GTGGTGGTTA	CGCGCAGCGT	GACCGCTACA	CTTGCCAGCG
3451	CCCTAGCGCC	CGCTCCTTTC	GCTTTCTTCC	CTTCCTTTCT	CGCCACGTTC
3501	GCCGGGCCTC	TCAAAAAAGG	GAAAAAAAGC	ATGCATCTCA	ATTAGTCAGC
3551	AACCATAGTC	CCGCCCTAA	CTCCGCCCAT	CCCGCCCCTA	ACTCCGCCCA
3601	GTTCCGCCCA	TTCTCCGCCC	CATGGCTGAC	TAATTTTTTT	TATTTATGCA
3651	GAGGCCGAGG	CCGCCTCGGC	CTCTGAGCTA	TTCCAGAAGT	AGTGAGGAGG
3701	CTTTTTTGGA	GGCCTAGGCT	TTTGCAAAAA	GCTTGGACAG	CTCAGGGCTG
3751	CGATTTCGCG	CCAAACTTGA	CGGCAATCCT	AGCGTGAAGG	CTGGTAGGAT
3801	TTTATCCCCG	CTGCCATCAT	GGTTCGACCA	TTGAACTGCA	TCGTCGCCGT
3851	GTCCCAAAAT	ATGGGGATTG	GCAAGAACGG	AGACCTACCC	TGGCCTCCGC
3901	TCAGGAACGA	GTTCAAGTAC	TTCCAAAGAA	TGACCACAAC	CTCTTCAGTG
3951	GAAGGTAAAC	AGAATCTGGT	GATTATGGGT	AGGAAAACCT	GGTTCTCCAT
4001	TCCTGAGAAG	AATCGACCTT	TAAAGGACAG	AATTAATATA	GTTCTCAGTA
4051	GAGAACTCAA	AGAACCACCA	CGAGGAGCTC	ATTTTCTTGC	CAAAAGTTTG
4101	GATGATGCCT	TAAGACTTAT	TGAACAACCG	GAATTGGCAA	GTAAAGTAGA
4151	CATGGTTTGG	ATAGTCGGAG	GCAGTTCTGT	TTACCAGGAA	GCCATGAATC
4201	AACCAGGCCA	CCTTAGACTC	TTTGTGACAA	GGATCATGCA	GGAATTTGAA
4251	AGTGACACGT	TTTTCCCAGA	AATTGATTTG	GGGAAATATA	AACTTCTCCC
4301	AGAATACCCA	GGCGTCCTCT	CTGAGGTCCA	GGAGGAAAA	GGCATCAAGT

FIGURE 18C

4351	ATAAGTTTGA	AGTCTACGAG	AAGAAAGACT	AACAGGAAGA	TGCTTTCAAG
4401	TTCTCTGCTC	CCCTCCTAAA	GCTATGCATT	TTTATAAGAC	CATGGGACTT
4451	TTGCTGGCTT	TAGATCTCTT	TGTGAAGGAA	CCTTACTTCT	GTGGTGTGAC
4501	ATAATTGGAC	AAACTACCTA	CAGAGATTTA	AAGCTCTAAG	GTAAATATAA
4551	AATTTTTAAG	TGTATAATGT	GTTAAACTAC	TGATTCTAAT	TGTTTGTGTA
4601	TTTTAGATTC	CAACCTATGG	AACTGATGAA	TGGGAGCAGT	GGTGGAATGC
4651	CTTTAATGAG	GAAAACCTGT	TTTGCTCAGA	AGAAATGCCA	TCTAGTGATG
4701	ATGAGGCTAC	TGCTGACTCT	CAACATTCTA	CTCCTCCAAA	AAAGAAGAGA
4751	AAGGTAGAAG	ACCCCAAGGA	CTTTCCTTCA	GAATTGCTAA	GTTTTTTGAG
4801	TCATGCTGTG	TTTAGTAATA	GAACTCTTGC	TTGCTTTGCT	ATTTACACCA
4851	CAAAGGAAAA	AGCTGCACTG	CTATACAAGA	AAATTATGGA	AAAATATTCT
4901	GTAACCTTTA	TAAGTAGGCA	TAACAGTTAT	AATCATAACA	TACTGTTTTT
4951	TCTTACTCCA	CACAGGCATA	GAGTGTCTGC	TATTAATAAC	TATGCTCAAA
5001	AATTGTGTAC	CTTTAGCTTT	TTAATTTGTA	AAGGGGTTAA	TAAGGAATAT
5051	TTGATGTATA	GTGCCTTGAC	TAGAGATCAT	AATCAGCCAT	ACCACATTTG
5101	TAGAGGTTTT	ACTTGCTTTA	AAAAACCTCC	CACACCTCCC	CCTGAACCTG
5151	AAACATAAAA	TGAATGCAAT	TGTTGTTGTT	AACTTGTTTA	TTGCAGCTTA
5201	TAATGGTTAC	AAATAAAGCA	ATAGCATCAC	AAATTTCACA	AATAAAGCAT
5251	TTTTTTCACT	GCATTCTAGT	TGTGGTTTGT	CCAAACTCAT	CAATGTATCT
5301	TATCATGTCT	GGATCGGCTG	GATGATCCTC	CAGCGCGGG	ATCTCATGCT
5351	GGAGTTCTTC	GCCCACCCCA	ACTTGTTTAT	TGCAGCTTAT	AATGGTTACA
5401	AATAAAGCAA	TAGCATCACA	AATTTCACAA	ATAAAGCATT	TTTTTCACTG
5451	CATTCTAGTT	GTGGTTTGTC	CAAACTCATC	AATGTATCTT	ATCATGTCTG
5501	TATACCGTCG	ACCTCTAGCT	AGAGCTTGGC	GTAATCATGG	TCATAGCTGT
5551	TTCCTGTGTG	AAATTGTTAT	CCGCTCACAA	TTCCACACAA	CATACGAGCC
5601	GGAAGCATAA	AGTGTAAAGC	CTGGGGTGCC	TAATGAGTGA	GCTAACTCAC
5651	ATTAATTGCG	TTGCGCTCAC	TGCCCGCTTT	CCAGTCGGGA	AACCTGTCGT
5701	GCCAGCTGCA	TTAATGAATC	GGCCAACGCG	CGGGGAGAGG	CGGTTTGCGT
5751	ATTGGGCGCT	CTTCCGCTTC	CTCGCTCACT	GACTCGCTGC	GCTCGGTCGT
5801	TCGGCTGCGG	CGAGCGGTAT	CAGCTCACTC	AAAGGCGGTA	ATACGGTTAT

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535	1	CCACAGAATC	AGGGGATAAC	GCAGGAAAGA	ACATGTGAGC	AAAAGGCCAG
590	1	CAAAAGGCCA	GGAACCGTAA	AAAGGCCGCG	TTGCTGGCGT	TTTTCCATAG
595	1	GCTCCGCCCC	CCTGACGAGC	ATCACAAAAA	TCGACGCTCA	AGTCAGAGGT
600	1	GGCGAAACCC	GACAGGACTA	TAAAGATACC	AGGCGTTTCC	CCCTGGAAGC
605	1	TCCCTCGTGC	GCTCTCCTGT	TCCGACCCTG	CCGCTTACCG	GATACCTGTC
610	1	CGCCTTTCTC	CCTTCGGGAA	GCGTGGCGCT	TTCTCAATGC	TCACGCTGTA
615	1	GGTATCTCAG	TTCGGTGTAG	GTCGTTCGCT	CCAAGCTGGG	CTGTGTGCAC
620	1	GAACCCCCCG	TTCAGCCCGA	CCGCTGCGCC	TTATCCGGTA	ACTATCGTCT
625	1	TGAGTCCAAC	CCGGTAAGAC	ACGACTTATC	GCCACTGGCA	GCAGCCACTG
630	1	GTAACAGGAT	TAGCAGAGCG	AGGTATGTAG	GCGGTGCTAC	AGAGTTCTTG
635	1	AAGTGGTGGC	CTAACTACGG	CTACACTAGA	AGGACAGTAT	TTGGTATCTG
640	1	CGCTCTGCTG	AAGCCAGTTA	CCTTCGGAAA	AAGAGTTGGT	AGCTCTTGAT
645	1	CCGGCAAACA	AACCACCGCT	GGTAGCGGTG	GTTTTTTGT	TTGCAAGCAG
650	1	CAGATTACGC	GCAGAAAAA	AGGATCTCAA	GAAGATCCTT	TGATCTTTTC
655	1	TACGGGGTCT	GACGCTCAGT	GGAACGAAAA	CTCACGTTAA	GGGATTTTGG
660	1	TCATGAGATT	ATCAAAAAGG	ATCTTCACCT	AGATCCTTTT	AAATTAAAA
665	51	TGAAGTTTTA	AATCAATCTA	AAGTATATAT	GAGTAAACTT	GGTCTGACAG
570	1	TTACCAATGC	TTAATCAGTG	AGGCACCTAT	CTCAGCGATC	TGTCTATTTC
675	51	GTTCATCCAT	AGTTGCCTGA	CTCCCCGTCG	TGTAGATAAC	TACGATACGG
680)1	GAGGGCTTAC	CATCTGGCCC	CAGTGCTGCA	ATGATACCGC	GAGACCCACG
689	51	CTCACCGGCT	CCAGATTTAT	CAGCAATAAA	CCAGCCAGCC	GGAAGGGCCG
690)1	AGCGCAGAAG	TGGTCCTGCA	ACTTTATCCG	CCTCCATCCA	GTCTATTAAT
699	51	TGTTGCCGGG	AAGCTAGAGT	AAGTAGTTCG	CCAGTTAATA	GTTTGCGCAA
700	01	CGTTGTTGCC	ATTGCTACAG	GCATCGTGGT	GTCACGCTCG	TCGTTTGGTA
70	51	TGGCTTCATT	CAGCTCCGGT	TCCCAACGAT	CAAGGCGAGT	TACATGATCC
710	01	CCCATGTTGT	GCAAAAAAGC	GGTTAGCTCC	TTCGGTCCTC	CGATCGTTGT
71	51	CAGAAGTAAG	TTGGCCGCAG	TGTTATCACT	CATGGTTATG	GCAGCACTGC
72	01	ATAATTCTCT	TACTGTCATG	CCATCCGTAA	. GATGCTTTTC	TGTGACTGGT
72	51	GAGTACTCAA	CCAAGTCATT	CTGAGAATAG	TGTATGCGGC	GACCGAGTTG
73	01	CTCTTGCCCG	GCGTTAATAC	GGGATAATAC	CGCGCCACAT	AGCAGAACTT

FIGURE 18E 32/56

7351	TAAAAGTGCT	CATCATTGGA	AAACGTTCTT	CGGGGCGAAA	ACTCTCAAGG
7401	ATCTTACCGC	TGTTGAGATC	CAGTTCGATG	TAACCCACTC	GTGCACCCAA
7451	CTGATCTTCA	. GCATCTTTTA	CTTTCACCAG	CGTTTCTGGG	TGAGCAAAAA
7501	CAGGAAGGCA	AAATGCCGCA	AAAAAGGGAA	TAAGGGCGAC	ACGGAAATGT
7551	TGAATACTCA	TACTCTTCCT	TTTTCAATAT	TATTGAAGCA	TTTATCAGGG
7601	TTATTGTCTC	ATGAGCGGAT	ACATATTTGA	ATGTATTTAG	AAAAATAAAC
7651	AAATAGGGGT	TCCGCGCACA	TTTCCCCGAA	AAGTGCCACC	TGACGTCGAC
7701	GGATCGGGAG	ATCTGCTAGG	TGACCTGAGG	CGCGCCGGCT	TCGAATAGCC
7751	AGAGTAACCT	TTTTTTTAA	TTTTATTTTA	TTTTATTTT	GAGATGGAGT
7801	TTGGCGCCGA	TCTCCCGATC	CCCTATGGTC	GACTCTCAGT	ACAATCTGCT
7851	CTGATGCCGC	ATAGTTAAGC	CAGTATCTGC	TCCCTGCTTG	TGTGTTGGAG
7901	GTCGCTGAGT	AGTGCGCGAG	CAAAATTTAA	GCTACAACAA	GGCAAGGCTT
7951	GACCGACAAT	TGCATGAAGA	ATCTGCTTAG	GGTTAGGCGT	TTTGCGCTGC
8001	TTCGCGATGT	ACGGGCCAGA	TATACGCGTT	GACATTGATT	ATTGACTAGT
8051	TATTAATAGT	AATCAATTAC	GGGGTCATTA	GTTCATAGCC	CATATATGGA
8101	GTTCCGCGTT	ACATAACTTA	CGGTAAATGG	CCCGCCTGGC	TGACCGCCCA
8151	ACGACCCCCG	CCCATTGACG	TCAATAATGA	CGTATGTTCC	CATAGTAACG
8201	CCAATAGGGA	CTTTCCATTG	ACGTCAATGG	GTGGACTATT	TACGGTAAAC
8251	TGCCCACTTG	GCAGTACATC	AAGTGTATCA	TATGCCAAGT	ACGCCCCTA
8301	TTGACGTCAA	TGACGGTAAA	TGGCCCGCCT	GGCATTATGC	CCAGTACATG
8351	ACCTTATGGG	ACTTTCCTAC	TTGGCAGTAC	ATCTACGTAT	TAGTCATCGC
8401	TATTACCATG	GTGATGCGGT	TTTGGCAGTA	CATCAATGGG	CGTGGATAGC
8451	GGTTTGACTC	ACGGGGATTT	CCAAGTCTCC	ACCCCATTGA	CGTCAATGGG
8501	AGTTTGTTTT	GGCACCAAAA	TCAACGGGAC	TTTCCAAAAT	GTCGTAACAA
8551	CTCCGCCCCA	TTGACGCAAA	TGGGCGGTAG	GCGTGTACGG	TGGGAGGTCT
8601	ATATAAGCAG	AGCTCTCTGG	CTAACTAGAG	AACCCACTGC	TTACTGGCTT
8651	ATCGAAATTA	ATACGACTCA	CTATAGGGAG	ACCCAAGCTT	

FIGURE 18F

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AGCAGCITIGG GCACCCAGAC CTACATCTGC AACGTGAATC ACAAGCCCAG CAACACCAAG TCTTCGGGTC GTTGTGGTTC CGINGCCCTNCC GCACGGGAGG GGTACCAAIT TAAATTGATA TCTCCTTAGG TCTCGAGTCT CTAGATAACC GGTCAATCGA 70 80 90 100 120 120 120 Triceantrict recedence receasing the resonance of GCCCTGGGCT GCCTGGTCAA GGACTACTTC AGGAGGTYCT CGINGGAGACC CCCGINGINGCC CGGGACCCGA CGGACCAGTY CCINGAINGAAG GCACACCTTC CGTCTGGAAG CCATGGTTAA ATTTAACTAT AGAGGAATCC AGAGCTCAGA GATCTATTGG CCAGTTAGCT AACCTTAAGA ACGCCGGCGA ACGATCGTGG TTCCCGGGTA GCCAGAAGGG GGACCGTGGG CCCGAACCGG TGACGGTGTC GTGGAACTCA GGCGCCCTGA CCAGCGGCGT GGGCTTGGCC ACTGCCACAG CACCTTGAGT CCGCGGGACT GGTCGCCGCA CCGGCTGTCC TACAGTCCTC AGGACTCTAC TCCCTCAGCA GCGTGGTCAC GGCCGACAGG AIGICAGGAG TCCTGAGATG AGGGAGTCGT CGCACCAGTG TCGTCGAACC CGTGGGTCTG GATGTAGACG TTGCACTTAG 280 400 160 TCCTCCAAGA GCACCTCTGG GGGCACAGCG 390 330 150 260 140

GYGGACAAGA AAGTTGGYGA GAGGCCAGCA CAGGGAGGGA GGGTGTCTGC TGGAAGCCAG GCTCAGCGCT CCTGCCTGGA CGCATCCCGG CTAIGCAGCC CCAGTCCAGG GCAGCAAGGC CGAGTCGCGA GGACGGACCT GCGTAGGGCC GATACGTCGG GGTCAGGTCC CGTCGTTCCG CACCTG1TCT

"FCAACCACT CICCGGTCGT GTCCCTCCCT CCCACAGACG ACCTTCGGTC

380

160

450

440

AGGCCCCGTC TGCCTCTTCA CCCGGAGGCC TCTGCCCGCC CCACTCATGC TCAGGGAGAG

TUCGGGGCAG AUGGAGAAGT GGGCCTCCGG AGAUGGGCGG GGTGAGTAUG

AGTCCCTCTC

GGTCTTCTGG CTTTTTCCCC AGGCTCTGGG CAGGCACAGG CTAGGTGCCC CTAACCCAGG CCAGAAGACE GAAAAAGGGG TCCGAGACCC GTCCGTGTCC GATCCACGGG GATTGGGTCC

FIGURE 19 A

GAGINCAAGE CYCATGETCA 1140 TECTICITICE CECAAAACEC AAGGACACEC TEATGNIETE CEGGACECET GAGGTEACAT GCGTGG'I'GGT GGACGTGAGC CACGAAGACC CTGAGGI'CAA GTTCAACTGG TACGTGGACG CGCACCACCA CCTGCACITCG GTGCTTCTGG GACTCCAGITT CAAGTTGACC AIGCACCTGC CGCGGGGGG GCAGTACAAC AGCACGTACC CGCACCICCA CGTAITACGG ITCTGTTTCG GCGCCCTCCT CGTCATGTTG ICGTGCATGG 860 870 880 890 900 GACAGGTCCC CAGCCGGGTG 235 950237 960 qetebeqeea ccercagret AGGAGAAGGG GGGTITTIGGG ITTCCTGTGGG AGIACIIAGAG GGCCTGGGGA CTCCAGTGTA CACACATGCC CACCGTGCCC AGGTAAGCCA GCCCAGGCCT CGCCCTCCAG CTGTCCACGG GATCTCATCG GACGTAGGTC CCTGTCCGGG GTCGGCCCAC CCCTGCCCCT GACCTAAGCC CACCCCAAAG GCCAAACTCT CCACTCCCTC AGCTCGGACA GGGACGGGGA CTGGATTCGG GTGGGGTTTC CGGTTTGAGA GGTGAGGGAG TCGAGCCTGT CCTTCTCTCC TCCCAGATIC CAGTAACTCC CAATCTTCTC TCTGCAGAGC CCAAATCTTG AGGGTCTAAG GTCATTGAGG GTTAGAAGAG AGACGTCTCG GGTTTAGAAC ACTGTTTTGA GTGTGTACGG GTGGCACGGG TCCATTCGGT CGGGTCCGGA GCGGGAGGTC deachecher eccaeteaga AAGAGCCATA TCCGGGAGGA GGGACCIGIG TITCCCCGIC CACGACCCGA GICTGGACGG ITCICGGIAI AGGCCCICCI CTUTGUICAG UGTCCTCACC GTCCTGCACC AGGACTGGCT GAATGGCAAG CACACCACHE GEAGGAGIGG CAGGACGTGG TCCTGACCGA CITIACCGTTC 1130 1010 940 GACTGTGCAG GTGGAGGTAG AGAAGGAGTC GTGGACTTGA CTGACACGTC CACCTCCATC TCTTCCTCAG CACCTGAACT 1120 CCCTGCACAC AAAGGGGCAG GTGCTGGGCT CAGACCTGCC 1000 820 CCGTGCAGGT GCATAATGCC AAGACAAAGC 1170 1110 990 1050 810 930 750 1160 1100 1040 800 1150 CTCAAGGCGG 1030 1090 TGACAAAACT GAGTTCCGCC GGAAGAGAGG

FIGURE 19B

1240

123033

1210

TACACCCTGC CCCCATCCCG GGATGAGCTG ACCAAGAACC AGGTCAGCCT GACCTGCCTG AAAGCCAAAG ACCCTCTGCC ACCACAGGTG TYGGAGACAG GGATGTCCCG TCGGGGCTCT TGGTGTCCAC ATCTGGGACG GGGGTAGGGC CCTACTCGAC TGGTTCTTGG TCCAGTCGGA CTGGACGGAC GCAGCCGGAG TYTYCGGTYTYC TGGGAGACGG CAGTITICCGA AGATAGGGTC GCTGTAGCGG CACCTCACCC TCTCGTTACC CGTCGGCCTC GCTCCTTCTT CCTCTACAGC TICITIGATOT TOTOGICCE AGGGCACGAC CICAGGCICC CCAGGAAGAA GGAGAIGICG AAGCTCACCG TGGACAAGAG CAGGTGGCAG CAGGGGAACG 11C1TTCTCATG CTCCGTGATG GTCCCCTTGC AGAAGAGTAC GAGGCACTAC GGGTAAATGA CCAGCGCTGC TTCTCGGAGA GGGACAGAGG CCCATTTACT GTGCGACGGC CGGCAAGCCC CCGCTCCCCG GGCTCTCGCG GTCGCACGAG GATGCTTGGC CTACGAACCG 1YCLATIGGGGG ACATISTATIGA AGGGCCCGGG GGIYCGTACCT TTATTITYCGTG GGTYGCGACG AGCCCCGAGA creceaged edaregagaa aaccareree TYCGTAGAGG GCTCGGCCC TGTACATACT TCCCGGGCGC CCAGCATGGA AATAAAGCAC 1310 CCGAGCCGGG 1370 GTCAAAGGCT TCTATCCCAG CGACATCGCC GTGGAGTGGG AGAGCAATGG CCCTGTCTCC CCGAGAGCGC CAGCGTGCTC CTGAGAGTGA CCGCTGTACC AACCTCTGTC CCTACAGGGC CAYGAGGCTC TGCACAACCA CTACACGCAG AAGAGCCYCT GAGGGTCGGG GGFAGCTCTT GTGGGACCCG TGGGGTGCGA GGGCCACATG GACAGAGGCC CCCGGTGTAC CTGTCTCCGG AACAACTACA AGACCACGCC TCCCGTGCTG GACTCCGACG 1300 1360 1540 1780 1420 1480 1660 GTCCACCGTC GTACTCCGAG ACGIGITGGT GATGTGCGTC 1350 GGCGAGGGGC 1410 1470 1530 1650 GCAAGGTCTC CAACAAAGCC correcadas smermess CACCCTGGGC ACCCCACGCT GACTCTCACT GGCGACATGG ACCTGTTCTC CACGCTGCCG GCCGTTCGGG 1340 1460 1520 1640 ACGTACCCCC 1270 1510 TYCGAGINGGC 1630 1450

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FICURE 190

CTGAGGCCTG GGACCCCGGG ACCTCTGAC ACTACCAAGA AAGGTGCCCA GTCCGGCTCA GACTCCGGAC GCTGTGCAGG GGTGGGGGAT CCTTCGGGAT 2100 CGACACGTCC CCACCCCCTA GGAAGCCCTA GGGACAGACA CACAGCCCCT GCCTCTGTAG GAGACTGTCC TGTTCTGTGA ACAAGACACT CTCGGGGGCA TGCTGGGGAT GCGGTGGGCT C'IATICOC'ITIC TCAGGCGGAA AGAACCAGCT GGGGCTC'IAG GGGGTATCCC CACGCGCCCT CATACCGAAG ACTCCGCCTT TCTTGGTCGA CCCCGAGATC CCCCATAGGG GTGCGCGGGA COCOGOGACA GGAGGCCTGG AGGTACGGGT GAGCCCCCGT ACGACCCCTA CGCCACCCGA AGTOGCATOA GCGAGGCAGA GCGGGTCCCA CTGTCCCCAC ACTGGCCCAG CCGGGAGGGA GGTCGTCGTG GACGGGACCC GACCCGGTGC CAGGCCGAGT CCCCCAGGGT GACAGGGGTG TGACCGGGTC CTGGGCCACG CTCTGACAGG CCCTCGGCAG GGTCCCCGAC GGGAGCCGTC 1970 2030 2090 2150 2210 TGCGAGACIG TGATGGTTCT TTCCACGGGT CCAGGGGCTG GGCCCTCCCT CCAGCAGCAC CTGCCCTGGG 1960 2020 2080 CCCTGTCTGT GIGTCGGGGA CGGAGACATC 2140 2200 CCCCCTAGGG 1GGGGCTCAG ACCCCGAGTC 2130 CCTCCCGACC TCCATGCCCA 1890 1950 2010 2070 2190 CCCTCCGTCT GGGGGATCCC 2120 1940 2180 TCACCGTACT TTGCCAGCGT GGAGCCCCTG GCCCCCTGT cordinate cordAACGGTCGCA ACACGGACCC TGYCCCTGGG CCTCGGGGAC 2110 2170

37/56

CGATGTGAAC

GTCGCACTGG

ACCAATGCGC

CCCCCACACC

TAATTCGCGC

CATCGCCGCG

GTAGCUGCGC ATTAAGCGCG GCGGGIGTGG TGGTTACGCG CAGCGTGACC GCTACACTTG

2250

2260

CCTYTCGCTT TCTTCCCTTC CTTTCTCGCC ACGTTCGCCG

2320

2310

2300

CCAGCGCCCT AGCGCCCGCT

2330

GGICGCGGGA ICGCGGGCGA GGAAAGCGAA AGAAGGGAAG GAAAGAGCGG IGCAAGCGGC

TCAAGCTCTA AATCGGGGCA TCCCTTTAGG GTTCCGATTT AGTGCTTTAC

2380

2370

2360

GCTTTCCCCG

CGAAAGGGGC AGTTCGAGAT TTAGCCCCGT AGGGAAATCC CAAGGCTAAA TCACGAAATG

FICURE 19D

2460	2520	2580	2640	2700	2760	2820	2880	2910 2940	3000
CCATCGCCCT	GGACTCTTGT	TAAGGGATTT	AACGCGAATT	CAGGCAGGCA	CTAACTCCGC	TGACTAATTT	AAGTAGTGAG	ACAGCTCAGG GCTGCGATTT	CCCGCTGCCA
GGTAGCGGGA	CCTGAGAACA	ATTCCCTAAA	TTGCGCTTAA	GTCCGTCCGT	GATTGAGGCG	ACTGATTAAA	TTCATCACTC	TGTCGAGTCC CGACGCTAAA	GCCCCACGCT
2450 ACGTAGTGGG TGCATCACCC	2510 CTTTAATAGT GAAATTATCA	2580 2570 2580 CGGTCTATFIC TFFTGATFITA TAAGGGATFIT GCCAGATAAG AAAACTAAAT ATFICCCTAAA	2620 2630 AGCTGATTTA ACAAAAATTT TCGACTAAAT TGTTTTTAAA	2690 CCAGGCTCCC GGTCCGAGGG	2750 AGTCCCGCCC TCAGGGCGGG	2800 2810 CCCATTCTCC GCCCCATGGC GGGTAAGAGG CGGGGTACCG	2870 GCTATTCCAG CGATAAGGTC		2990 GGATTTTATC CCI'AAAAI'AG
2440	2500	2560	2620	2680	2740		2860	2920	2980
GTGATGGTTC	AGTCCACG1T	CGGTCTATTC	AGCTGATTTA	TGGAAAGTCC	CAGCAACCAT		CGGCCTCTGA	AAAAGCT'IGG	AAGGCTGGTA
CACTACCAAG	TCAGGTGCAA	GCCAGATAAG	TCGACTAAAT	ACCTTTCAGG	GTCGTTGGTA		GCCGGAGACT	TTTT'CGAACC	TTCCGACCA'F
2430	2490	2550	2610	2670	2730	2790	2840 2850 2860	2910 2920	2950 2950 2970 CGCGCCAAAC TTGACGGCAA TCCTAGCGTG GCGCCCITITIC AACTGCCGTT AGGATCGCAC
CTTGATTAGG	TTTTCGCCCT TTGACGTTGG	AACCCTATCT	TTAAAAAATG	AGTTAGGGTG	CTCAATTAGT	CCCAGTYCCG	TGCAGAGGCC GAGGCCGCCT CGGCCTCTGA	GGCTTYTGCA AAAAGCTYGG	
GAACTAATCC	AAAAGCGGGA AACTGCAACC	TTGGGATAGA	AATTTTTTAC	TCAATCCCAC	GAGTTAATCA	GGGYCAAGGC	ACGTCTCCGG CTCCGGCGGA GCCGGAGACT	CCGAAAACGT TTTTCGAACC	
2420		2540	2600	2660	2720	2780	2840	2900	2960
CCCCAAAAA		AACAACACTC	GGCCTATTGG	AATGTGTGTC	AAGCATGCAT	CCTAACTCCG	TGCAGAGGCC	TGGAGGCCTA	TTGACGGCAA
GGGGTTTTTT		TTGTTGTGAG	CCGGATAACC	TTACACACAG	TTCGTACGTA	GGATTGAGGC	ACGTCTCCGG	ACCTCCGGAT	AACTGCCGTT
2410	2470	2530	2590	2650	2710	2770	2830	2890	2950
GGCACCTCGA	GATAGACGGT	TCCAAACTGG	TGGGGATTTC	AATTCTGTGG	GAAGTATGCA	CCATCCCGCC	TTTTTATTTA	GAGGCTTYTY	CGCGCCAAAC
CCCTGGAGCT	CTATCTGCCA	AGGTTTGACC	ACCCCTAAAG	TTAAGACACC	CTTCATACGT	GGTAGGGCGG	AAAATAAAT	CTCCGAAAAA	GCGCGCIIIIIKG
							3 8	156	

ICURE 1

CCGTCTCCCA AAATATGGGG ATTGGCAAGA 3030 3010

TAACCGTTCT TTTATACCCC GGCACAGGGT TCATGGITCG ACCATIGAAC TGCATCGTCG

TGGTAACTTG ACGTAGCAGC AGTACCAAGC

AGAATGACCA GTACTTCCAA ACCCTGGCCT CCGCTCAGGA ACGAGTTCAA 3100 3090 3080 ACGGAGACCT

TGCCTCTGGA TGGGACCGGA GGCGAGTCCT TGCTCAAGTT CATGAAGGTT TCTTACTGGT

GGGTAGGAAA ACCTGGTTCT GITGGAGAAG TCACCTTCCA TYTGTCTTAG ACCACTAATA CCCATCCTTT 3170 AGTGGAAGGT AAACAGAATC TGGTGATTAT 3160 3150 3140 CAACCTCTTC 3130

TGGACCAAGA 3210 CCATICCINGA GAAGAATCGA 3200

CCTTTAAAGG ACAGAATTAA TATAGTTCTC AGTAGAGAAC TGTCTTAATT ATATCAAGAG TCATCTTG CTTCTTAGCT GGAAATTTCC

GCTCATTTTC TTGCCAAAAG TTTGGATGAT GCCTTAAGAC 3280 3270 3260

GGTAAGGACT

3190

3250

ACITITIC TIGGIGCICCT CGAGTAAAAG AACGGITITIC AAACCTACTA CGGAATICITG 3350 3330 TCAAAGAACC ACCACGAGGA

GGAGGCAGTT CCTCCGTCAA TTATTGAACA ACCGGAATTG GCAAGTAAAG TAGACATGGT TTGGATAGTC 3320 3310

AATAACTTGT TGGCCTTAAC CGTTCATTTC ATCTGTACCA AACCTATCAG 3410 3400 3390 3380 3370

CIKITITACCA GGAAGCCAIG AAIKCAACCAG GCCACCITIAG ACTCTITIGTG ACAAGGATICA THAGITGGIC CGCTGGAATC TGAGAAACAC TGTTCCTAGT GACAAATIGGI CCTTCGGTAC

TTTGGGGAAA TATAAACTTC ACGICCITAA ACITICACIG IGCAAAAAGG GICITITAACI AAACCCCITI ATAITIGAAG TGCAGGAATT' TGAAAGTGAC ACGTTTTTTCC CAGAAATTGA

TCCCAGAATA CCCAGGCGTC CTCTCTGAGG TCCAGGAGGA AAAAGGCATC AAGTATAAGT TTTTCCGTAG AGGGICTITAT GGGTCCGCAG GAGAGACTCC AGGTCCTICCT 3520 3510 3490

ITTCATATITCA

TTGAAGTCTA CGAGAAGAAA GACTAACAGG AAGATGCTTT CAAGTTCTCT GCTCCCTCC

3560

AACTICAGAI GUTCTICIIT CTGATIGICC TICTACGAAA GITCAAGAGA CGAGGGGAGG

29156

FIGURE 19F

3650

3640

3630

TCTTTGTGAA ATTTCGATAC GTAAAAATAT TCTGGTACCC TGAAAACGAC CGAAATCTAG AGAAACACTT GGACAAACTA CCTACAGAGA 17TTAAAGCTC CCTG'ITTGAT GGATGTCTCT AAATTTCGAG TAAGGIAAAI ATAAAATITT TAAGIGIAIA ATGIGIIAAA CIACIGATIC TAATIGITIIG TACACAATI'TI GATIGACTAAG ATITAACAAAC TGAATGGGAG CAGTGGTGGA ATGCCTTTAA ACATAAAATC TAAGGTIGGA TACCTIGACT ACTTACCCTC GTCACCACCT TACGGAAATT GATGACGACT GAGAAAGGTA GAAGACCCCA AGGACTTTCC GAGAGITICTA AGATGAGGAG GITTTTTTCTT CTCTTTCCAT CTTCTGGGGT TCCTGAAAGG ACTGCTATAC AAGAAATTA TGGAAAATA TTCTTTTAAT ACCTTTTTAT GGCATAACAG TTATAATCAT AACATACTGT TTTTTCTTAC AGGTGTGTCC GTATCTCACA GACGATAATT ATTGATACGA GTTTTTAACA CATGGAAATC CTACTGCTGA MYCAGAANITG CTAAGTIYIYI TGAGTCATGC TGTGTYTAGT AATAGAACTC TTGCTTGCTY NAGTCTTYAAC GATTCAAAAA ACTCAGTACG ACACAAATCA TTATCTTGAG AACGAACGAA CCGTATTGTC AATAITTAGTA TTGTATGACA AAAAAGAATG TAACTATGCT CAAAAATTGT GTACCTTTAG GCTTTAGATC GCCATCTAGT GATGATGAGG CTACTACTCC 3770 3830 3950 3890 4070 4130 4190 4010 PAAAGCTATG CATTITIATA AGACCATGGG ACTITITGCTG CGGTAGATCA 3700 3760 3820 3940 TTTTTCGACG TGACGATATG 3880 4060 4120 4180 4000 GGAACCTTAC TTCTGTGGTG TGACATAATT CCTTGGAATG AAGACACCAC ACTGTATTAA ATTICCATITE TATTITAAAA ATTICACATAT TICCTATTTAC ACCACAAAGG AAAAAGCTGC rgtattriag attccaacct atggaactga TGAGGAAAAC CTGTTTTGCT CAGAAGAAAT TCCACACAGG CATAGAGTGT CTGCTATTAA GACAAAACGA GTCTTCTTTA CTCTCAACAT TCTACTCCTC CAAAAAGAA 4050 4110 3690 3750 3810 3870 3930 4170 3990 3740 TTTATAAGTA AAATATTCAT 3680 3800 3860 3920 TGGTGTTTCC 4160 3980 4040 4100 ACTCCTTTTC TTCTGTAACC **AAGACAT'I'GG** 3850 **ACGA'TAAATG** 40

FIGURE 190

4240

4230

4220

CTTTTTAATT TGTAAAGGGG TTAATAAGGA ATATTTGATG TATAGTGCCT TGACTAGAGA GAAAAA1TAA ACATTTCCCC AATTATTCCT TATAAACTAC ATATCACGGA ACTGATCTCT CTCCCACACC TYTATTGCAG AGGGGGACT'' GGACTTYGTA TTYTACTTAC GTYYACAACA ACAATYGAAC AAATAACGTC CGTAAAAAA TACTICION TICICCAAAC ICATCAAIGI AICTIAICAI GICTGGAICG GGGTTGAACA TIFATAATGGT TACAAATAAA GCAATAGCAT CACAAATTTC ACAAATAAAG GACGCINCTICG GTGACGTAAG ATCAACACCA AACAGGTTTG AGTAGTTACA TAGAATAGTA CAGACCTAGC CCCAACTTGT **GCATITITITI** AGTAITTAGTC GGTATGGTGT AAACATCTCC AAAAIGAACG AAATTTTTTG TCCCCCTGAA CCTGAAACAT AAAATGAATG CAATTGTTGT TGTTAACTTG TTACAAATAA AGCAATAGCA TCACAAATTT CACAAATAAA AATCITTAIT TCGITAICGI AGIGITTAAA GIGITTAITT GCTGGATGAT CCTCCAGCGC GGGGATCTCA TGCTGGAGTT CTTCGCCCAC CGACCTACTA GGAGGTCGCG CCCCTAGAGT ACGACCTCAA GAAGCGGGTG TTTAAAAAAC 4310 4430 4490 TTTTACTTGC 4360 4540 4420 4480 4600 TYTCTAGAGG 4410 4470 4530 4290 4350 4590 CCATACCACA 4340 4280 4400 4460 4520 1580 CTTATAATGG TCATAATCAG 12.10 GAAT'ATTACC CACTGCATTC TTATTGCAGC 4330 4390 4450 4510 4570

TCTTATCATG GIPAAAAAAAG TGACGTAAGA TCAACACCAA ACAGGTTTGA GTAGTTACAT AGAATAGTAC CTGTTTCCTG GACAAAGGAC CATTITITITE ACTICEATTET AGTTGTGGTT TGTCCAAACT CATCAATGTA GTCGACCTCT AGCTAGAGCT TGGCGTAATC ATGGTCATAG AGACATATES CASCTEGAGA TCGATCTCGA ACCGCATTAG TACCAGTATC 4730 4670 4660 4720 4650 4710 4640 4700 TCTGTATACC 1630 4690

TCTGAAATTG 1TATCCGCTC ACAATTCCAC ACAACATACG AGCCGGAAGC ATAAAGTGTA ACACITYIAAC AATAGGCGAG IGTTAAGGTG TGTIGTAIGC ICGGCCTTICG TAITYTCACAT

4770

4760

4740

TGTTTATTTC

GTGTTTAAAG

AATAACGICG AATATTACCA A'IGITTAITT CGTTATCGTA

41150

FIGURE 19H

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AAGCCTGGGG TGCCTAATGA GTGAGCTAAC TCACATTAAT TGCGTTGCGC TCACTGCCCG

AGTGACGGGC	4920	4980	5040	5100	5160	5220	5280	5340	5400
	CGCGCGGGGA	CTGCGCTCGG	TTATCCACAG	GCCAGGAACC	GAGCATCACA	TACCAGGCGT	ACCGGATACC	TGTAGGTATC	CCCGTTCAGC
	GCGCGCCCT	GACGCGAGCC	AATAGGTGTC	CGGTCCTTGG	CTCGTAGTGT	ATGGTCCGCA	TGGCC'FATGG	ACATCCATAG	GGGCAAGTCG
TICGBACCCC ACGGATTACT CACTCGATTG AGTGTAATTA ACGCAACGCG AGTGACGGG	4910	4970	5030	5090	5150	5210	5270	5330	5390
	AATCGGCCAA	CACTGACTCG	GGTAATACGG	CCAGCAAAAG	CCCCCCTGAC	ACTATAAAGA	CCTGCCGCTT	ATGCTCACGC	GCACGAACCC
	TTAGCCGGTT	GTGACTGAGC	CCATTATGCC	GGTCGTTTTC	GGGGGGACTG	TGA1ATTTC1	GGACGGCGAA	TACGAGTGCG	CGTGCTTGGG
AGTGTAATTA	4900	4960	5020	5080	5140	5200	5260	5320	5380
	TGCATTAATG	CTTCCTCGCT	ACTCAAAGGC	GAGCAAAAGG	ATAGGCTCCG	ACCCGACAGG	CTGTTCCGAC	CGCTTTCTCA	TGGGCTGTGT
	ACGTAATTAC	GAAGGAGCGA	TGAGTTTCCG	CTCGITTTTCC	TATCCGAGGC	TGGGCTCTCC	GACAAGGCTG	GCGAAAGAGT	ACCCGACACA
CACTCGATTG	4890	4950	5010	5070	5130	5190	5250	5310	5370
	TCGTGCCAGC	CGCTCTTTCCG	GTATCAGCTC	AAGAACATGT	GCGTTTTTCC	AGGTGGCGAA	GTGCGCTCTC	GGAAGCGTGG	CGCTCCAAGC
	AGCACGGTCG	GCGAGAAGGC	CATAGTCGAG	TTCTTGTACA	CGCAAAAAGG	TCCACCGCTT	CACGCGAGAG	CCTTCGCACC	GCGAGGTTCG
ACGGATTACT	4880	4940	5000	5060	5120	5180	5240	5300	5360
	GGGAAACCTG	GCGTATTGGG	GCGGCGAGCG	TAACGCAGGA	CGCGTTGCTG	CTCAAGTCAG	AAGCTCCCTC	TCTCCCTTCG	GTAGGTCGTT
	CCCTTTGGAC	CGCATAACCC	CGCCGCTCGC	ATTGCGTCCT	GCGCAACGAC	GAGTTCAGTC	TTCGAGGGAG	AGAGGGAAGC	CATCCAGCAA
TICGGACCCC	4870	4930	4990	5050	5110	5170	5230	5290	5350
	CTTTCCAGTC	GACGCGGTPP	TCGTTCGGCT	AATCAGGGGA	GTAAAAAGGC	AAAATCGACG	TTCCCCCTGG	TGTCCGCCTT	TCAGTTCGGT
	GAAAGGTCAG	CTCCGCCAAA	AGCAAGCCGA	1TAGTCCCCT	CAT'FTTTCCG	TTTTPAGCTGC	AAGGGGGACC	ACAGGCGGAA	AGTCAAGCCA

FIGURE 191

5450

5440

5430

5410

5640 5820 GAACTICACC ACCGGATIGA IGCCGAIGIG AICTICCIGI CAIAAACCAI GCTGAAGCCA GTTACCTTCG GAAAAAGAGT TGGTAGCTCT TGATCCGGCA CAGTGGAACG TIPITICCTAG AGTICTICIA GGAAACTAGA AAAGATGCCC CAGACTGCGA GICACCTTGC ACCTAGATCC TGGATCTAGG CAACCCGGTA AGACACGACT ACTOGTAACA GGATTAGCAG AGCGAGGTAT GTAGGCGGTG TGACCATTGT CCTAATCGTC TCGCTCCATA CATCCGCCAC CTYGAAGTGG TGGCCTAACT ACGGCTACAC TAGAAGGACA GTATTTGGTA CITITITICICA ACCATCGAGA ACTAGGCCGT ACGCGCAGAA GITGGGCCAT TCTGTGCTGA TCCCCCTCTT AAAACTCACG TTAAGGGATT TTGGTCATGA GATTATCAAA AAGGATCTTC GCAGCAGATT CCACCAAAAA AACAAACGTT CGTCGTCTAA TCAAGAAGAT CCTTTGATCT TTTCTACGGG GTCTGACGCT INTINGACTICE AATTCCCTAA AACCAGTACT CTAATAGTIT TTCCTAGAAG 5510 5570 5630 5690 5750 5810 GGTAACTATC GTCTTGAGTC CGCTGGTAGC GGTGGTTTTT TTGTTTGCAA 5740 CAGAACTCAG 5500 5560 5620 5680 5800 GCGGAATAGG CCATTGATAG 5550 CGACTTCGGT CAATGGAAGC 5670 5730 5790 5490 5610 GCGACCATCG GGCAGCAGCC CGCCTTATCC CCGTCGTCGG 5780 5480 5540 9995 5720 5600 TATCGCCACT C'TACAGAGT'T TCTGCGCTCT AACAAACCAC AAAAAGGATC GATGICTICAA AGACCCGAGA TTGTTTGGTG CCGACCGCTG GGCTGGCGAC 5470 ATAGCGGTGA 5710

CCATAGITIC CTGACTCCCC GTCGTGTAGA TAACTACGAT ACGGGAGGGC TTACCATCTG GGTPATCAACG GACTGAGGGG CAGCACATCT ATTGATGCTA TGCCCTCCCG AATGGTAGAC PGTCAATGGT TACGAATTAG TCACTCCGTG GATAGAGTCG 5980 5970 5960

TTTAAATCAA TCTAAAGTAT ATATGAGTAA ACTTGGTCTG

5860

5850

5840

TTTTAAATTA AAAATGAAGT

5830

AAATTTAAT

43150

5870

TYTTACITICA AAATTTAGIT AGAITTICATA TATACITCATT TGAACCAGAC

TTTCGTTCAT

GATCTGTCTA

CTATCTCAGC

ATGCTTAATC AGTGAGGCAC

ACAGTTACCA

2900

5920

5910

C'IAGACAGAT AAAGCAAGTA

FICURE 19J

6040

6030

6120 6180 TCCAATGATA CCGCGAGACC CACGCTCACC GCCTCCAGAT TTATCAGCAA CCGAGGTCTA AATAGTCGTT TGCAACTFTA TCCGCCTCCA TCGGCCTTCC CGCCTCGCGT CTTCACCAGG ACGTTGAAAT AGGCGGAGGT TCCACTCTAT TAATTGTTGC CGGGAAGCTA GAGTAAGTAG TTCGCCAGTT AATAGTTTGC AGGTCAGATA ATTAACAACG GCCCTTCGAT CTCATTCATC AAGCGGTCAA TTATCAAACG TGCCATTGCT ACAGGCATCG TGGTGTCACG CTCGTCGTTT GGTATGGCTT CGGTTCCCAA CGATCAAGGC GAGTTACATG ATCCCCCATG TTGTGCAAAA GCCAAGGGTT GCTAGTTCCG CTCAATGTAC TAGGGGGTAC AACACGTTTT AAGCGGTTAG CTCCTTCGGT CCTCCGATCG TYGTCAGAAG TAAGTTGGCC GCAGTGTTAT TATGGCAGCA CTGCATAATT CTCTTACTGT CATGCCATCC GTAAGATGCT CCCGGCGTCA ATACGGGATA ATACCGCGCC ACATAGCAGA ACTTTAAAAG CGTTGCAACA ACGGTAACGA TGTCCGTAGC ACCACAGTGC GAGCAGCAAA CCATACCGAA CGTCACAATA GYGAGYFACCA ATACCGTCGT GACGTATTAA GAGAATGACA GTACGGTAGG CATTCTACGA ITTCTCTGTGAC TGGTGAGTAC TCAACCAAGT CATTCTGAGA ATAGTGTATG CGGCGACCGA GCCGCTGGCT TECTCATCAT TEGAAAACGT TCTTCGGGGC GAAAACTCTC AAGGATCTTA CCGCTGTTGA NCGACTACTA ACCTYTTGCA AGAAGCCCCG CTYTTGAGAG TICCTAGAAT GCCGACAACT GGGCCGCAGT TATGCCCTAT TATGGCGCGG TGTATCGTCT TGAAATTTTC AAAGACACTG ACCACTCATG AGTTGGTTCA GTAAGACTCT TATCACATAC 6110 6290 GGAGGCTAGC AACAGTCTTC ATTCAACCGG 6230 6410 6530 6350 6470 GTGCGAGTGG GCCGAGCGCA GAAGTGGTCC 9190 6220 6340 6100 6400 6460 6280 6580 GCCCTCTGG 0609 6150 6210 6330 6390 6510 6570 6270 6450 CGGGGTCACG ACGTTACTAT AGCCGGAAGG 6200 6260 GAGGAAGCCA 6560 6080 6140 6320 6380 6440 TAAACCAGCC GCAACGTTGT CATTCAGCTC GCCCCAGINGC ATTIGGTCGG GTAAGTCGAG CACTCATGGT GTTGCTCTTG PICCCCAATC CAACGAGAAC 6190 6370 6430 6490

FICURE 19K

GGAATAAGGG CCTTATTCCC

CGCAAAAAAG

6710

6700

0699

6680

Tringiccin ccgrirmacg gcgimming

6760

CCAGCGI'ITC TGGGTGAGCA AAAACAGGAA GGCAAAATGC

GGTCGCAAAG ACCCACTCGT

TTTACTTTCA

TTCAGCATCT

CCAACTGATC

GATCCAGTTC GATGTAACCC ACTCGTGCAC

6640

6630

6620

CTAGGTCAAG CTACATTGGG TGAGCACGTG GGTTGACTAG AAGTCGTAGA AAATGAAAGT

CGAGCAAAAT TTAAGCTACA ACAAGGCAAG GCTTGACCGA CGAACTGGCT CAATIGCATG AAGAATCTGC TTAGGGTTAG GCGTTTTGCG CTGCTTCGCG ATGTACGGGC GTTAAUGTAC TICTTAGACG AATCCCAATC CGCAAAACGC GACGAAGCGC TACATGCCCG GGCTTCGAAT AGCCAGAGTA ACCTTTTTTT TTAATTTTAT CCGAAGCTTA TCGGTCTCAT TGGAAAAAA AATTAAAATA GATCCCCTAT GGTCGACTCT CAGTACAATC TGCTCTGATG CCGCATAGTT AAGCCAGTAT CTGCTCCCTG CTTGTGTGTT GAACACACAA AGGGTTATTG TCTCATGAGC GGATACATAT TTGAATGTAT TTAGAAAAAT AAACAAATAG GGAGATCTGC CCTCTAGACG AANTAAAATA AAAACTCTAC CTCAAACCGC GGCTAGAGGG CTAGGGGATA CCAGCTGAGA AGCATTTATC TCGTAAATAG TYTGTYTATC GCTCGTTTTA AATTCGATGT TGTTCCGTTC TTCGGTCATA GACGAGGGAC CGACACGGAA ATGTTGAATA CTCATACTCT TCCTTTTTCA ATATTATTGA TACAACTTAT GAGTATGAGA AGGAAAAAGT TATAATAACT TCCCAATAAC AGAGTACTCG CCTATGTATA AACTTACATA AATCTTTTTA CACCTGACGT CGACGGATCG CCCAAGGCGC GIGTAAAGGG GCTTTTCACG GTGGACTGCA GCTGCCTAGC 7010 7130 6950 6830 TITIATITIAT TITIGAGAIG GAGITITGGCG CCGAICICCC 7180 6880 6820 6940 7000 GTCATGTTAG ACGAGACTAC GGCGTATCAA 7170 CACATTTCCC CGAAAGTGC 7050 0669 7110 6930 6750 6810 6870 CTCCGCGCGG GGAGGTCGCT GAGTAGTGCG 7160 TAGGTGACCT GAGGCGCGCC 7100 CTCATCACGC 6860 6920 6980 6800 CCTCCAGCGA GGGTTCCGCG ATCCACTGGA 7030 7090 GCTGTGCCTT 6910 6970 6790 6850

FIGURE 19L

7310 7320 CTTACGGTAA ATGGCCCGCC GAATGCCATT TACCGGGCGG	7380 ATGACGTATG 1"ICCCATAGT TACTGCATAC AAGGGTATCA	7440 TATTTACGGT AAACTGCCCA ATAAATGCCA TTTGACGGGT	7500 CCTATTGACG TCAATGACGG GGATAACTGC AGTTACTGCC	7550 7560 TGGGACTTTC CTACTTGGCA ACCCTGAAAG GATGAACCGT	7620 AGTACATCAA TCATGTAGTT	7680 CTCCACCCCA TYGACGTCAA GAGGTGGGGT AACTGCAGTT	7740 AAATGTCGTA ACAACTCCGC TTTACAGCAT TGTTGAGGCG	7800
7310 CTTACGGTAA GAATGCCATT	7370 ATGACGTATG TACTGCATAC			7550 TGGGACTTTC ACCCTGAAAG	7610 CGGTTTTTGCC GCCAAAACCG	7670 CTCCACCCCA GAGGTGGGGT	7730 AAATGTCGTA TTTACAGCAT	0667
7300 CGTTACATAA GCAATGTATT	7360 GACGTCAATA CTGCAGITAAT	7420 ATGGGTGGAC TACCCACCTG	7480 AAGTACGCCC TTCATGCGGG	7540 CATGACCTTA GTACTGGAAT	7600 CATGGTGATG GTACCACTAC	7660 ATTTCCAAGT TAAAGGTTCA	7720 GGACTTTCCA CCTGAAAGGT	7780
7290 TGGAGT*PCCG ACCTCAAGGC	7350 CCCGCCCATT GGCCGGGTAA	7410 7420 ATTGACGTCA ATGGGTGGAC TAACTGCAGT TACCCACCTG	7470 ATCATATGCC TAGTATACGG	7530 ATGCCCAGTA TACGGGTCAT	7590 TCGCTATTAC AGCGATAATG	7630 7630 7650 7650 7650 7650 AGGCGTGGA TAGGGGTTTG ACTCAGGGG ATTTCCAAGT ACCCGCACC'F ATGCCCC TAAAGGTTCA	7710 AAAATCAACG TTTTAGTTGC	7770
7280 AGCCCATATA TCGGGTATAT	7340 CCCAACGACC GCGTTGCTGG	7400 GGGACTTTCC CCCTGAAAGG	7460 CATCAAGTGT GTAGTTCACA	7520 GCCTGGCATT CGGACCGTAA	7580 GTATTAGTCA CATAATCAGT	7640 TAGCGGTTTG ATCGCCAAAC	7700 TTTTGGCACC AAAACCGTGG	0911
7270 ATTAGTTCAT TAATCAAGTA	7330 TGGCTGACCG ACCGACTGGC	7390 AACGCCAATA TTGCGGTTA'F	7450 CTTGGCAGTA GAACCGTCAT	7510 TAAATGGCCC ATTTACCGGG	7570 GTACATCTAC CATGTAGATG	7630 TGGGCGTGGA ACCCGCACC'F	7690 TGGGAGTTTG ACCCTCAAAC	1750

7250

7240

7230

7220

7210

CAGATATACU CGITGACAIT GAITIAITIGAC TAGITATITAA TAGIAAITCAA ITTACGGGGTC GICTATATGC GCAACIGIAA CIAATAACIG AICAATAAIT AICAITAGIT AAIGCCCCAG

FIGURE 19M

46156

CCCATTGACG CAAATGGGCG GTAGGCGTGT ACGGTGGGAG GTCTATATAA GCAGAGCTCT GGGTAACTGC GTTTACCCGC CATCCGCACA TGCCACCCTC CAGATATAT CGTCTCGAGA

FIGURE 19N

7810 7850 7860 CTGCTTACTG CCTTATCGAA ATTAATACGA CTCACTATAG CACCGATTGA TCTCTTGGGT GACGAATGAC CGAATAGCTT TAATTATGCT GAGTGATATC

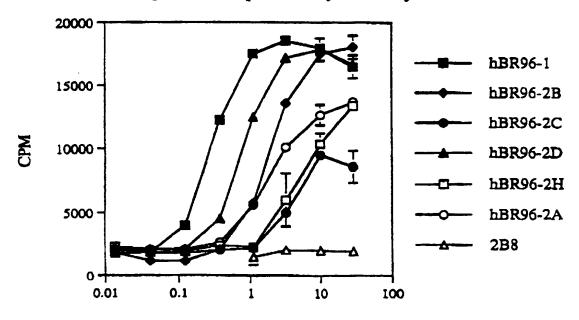
7870 GGAGACCCAA GCTT CCTCTGGGTT CGAA

7880

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FIGURE 20

Complement Dependent Cytotoxicity



Concentration IgG (µg/ml)

FIGURE 21

Antibody Dependent Cell-Mediated Cytotoxicity

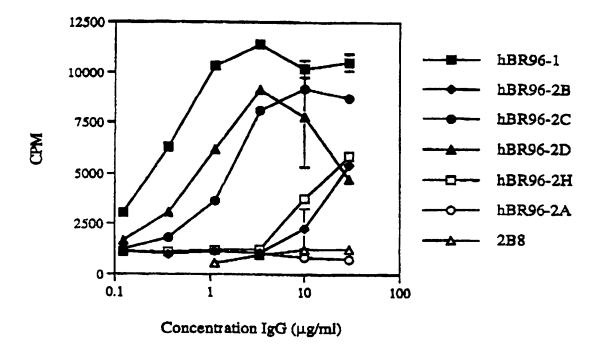
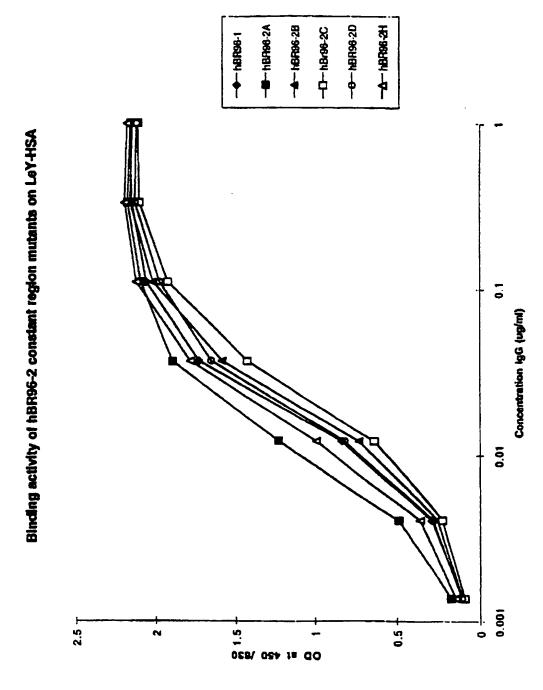
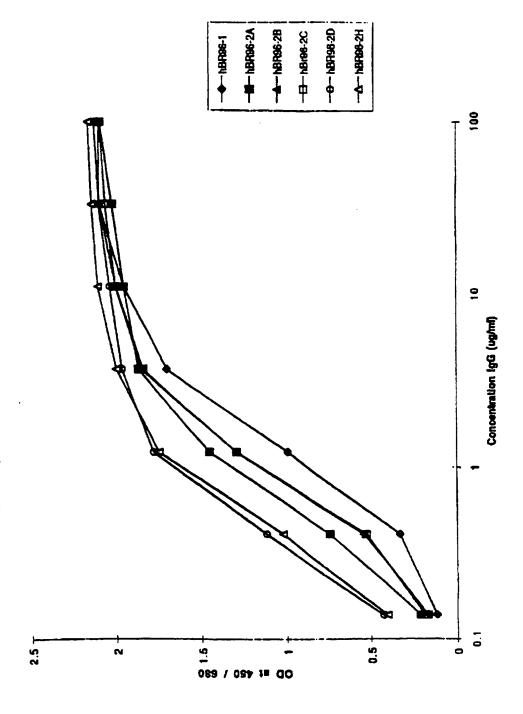


FIGURE 22

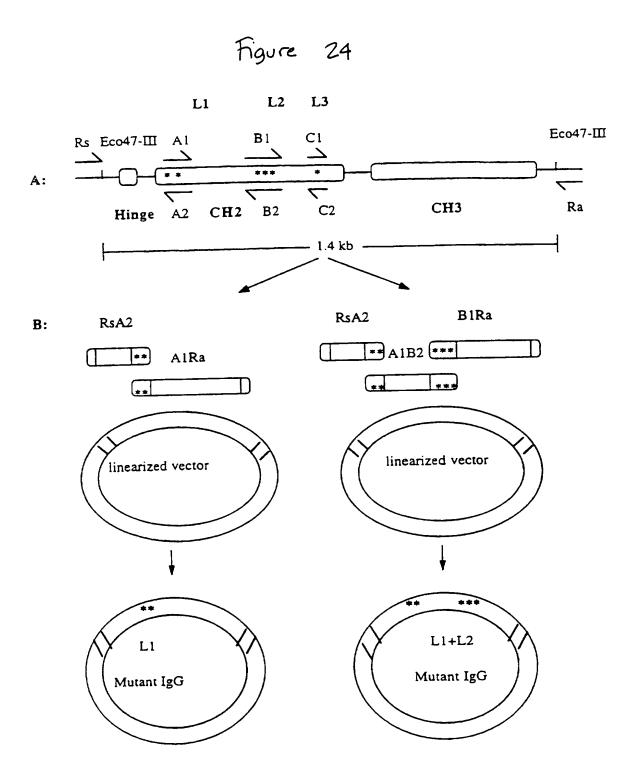


Binding activity of hBR96-2 constant region mutants on LNFPII-BSA

FIGURE 23



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Figure 25

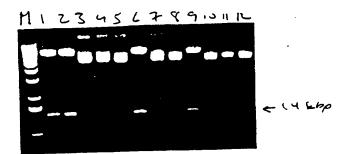


Figure 26

hBR96-2 Heavy Chain Variable Region (VH)

EVQLVESGGG LVQPGGSLRL SCAASGFPFS DYYMYWVRQA PGKGLEWVSY

51 61 71 881 91
ISQDGDITDY ADSVKGRFTI SRDNAKNSLY LQMINSLRDED TAVYYCARGL

101 ADGAWFAYWG QGTLVTVSS

human IgGI constant

YFPEPVTVSW NSGALTSGVH TFPAVLQSSG LYSLSSVVTV PSSSLCTQTY
ICNVNHKPSN TKVDKKVEPK SCDKTHTCPP CEAPEIDOOP SVFLFPPKPK
DTLMISRTPE VTCVVVDVSH EDPEVKFNWY VDGVEVHNAK TKPREEQYNS
18 10 311
TYRVVSVLTV LHQDWLNGRD YKOKVSNKAL PROPERTIESK AKCOPREPQV
YTLPPSRDEL TKNQVSLTCL VKGFYPSDIA VEWESNGQPE NNYKTTPPVL
DSDGSFFLYS KLTVDKSRWQ QGNVFSCSVM HEALHNHYTQ KSLSLSPGK

Figure 27

hBR96-2A: Heavy Chain Variable Region (VH)

EVQLVESGGG LVQPGGSLRL SCAASGFPFS DYYMYWVRQA PGKGLEWVSY

51 61 71 81 91
ISQDGDITDY ADSVKGRFTI SRDNAKNSLY LQMNSLRDED TAVYYCARGL

101 111
TADGAWFAYWG QGTLVTVSS

hBR96-2A: Human Heavy Chain IgG1 Constant Region ACH2

A STKGPSVFPL APSSKSTSGG ȚAALGCLVKD YFPEPVTVSW NSGALTSGVH

TFPAVLQSSG LYSLSSVVTV PSSSLGTQTY ICNVNHKPSN TKVDKKVEPK

SCDKTHTCPP CP GQPREPQV YTLPPSRDEL TKNQVSLTCL VKGFYPSDIA

VEWESNGQPE NNYKTTPPVL DSDGSFFLYS KLTVDKSRWQ QGNVFSCSVM

HEALHNHYTG KSLSLSPGK

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Figure 28

This sequence is the chi BR96 IgG1 with CH2 deleted.

1 EVNLVESGGG LVQPGGSLKV SCVTSGFTPS DYYMYWVRQT PEKRLEWVAY
5: ISQGGDITDY PDTVKGRFTI SRDNAKNTLY LQMSRLKSED TAMYYCARGL
101 DDGAWFAYWG QGTLVTVSVA STKGPSVFPL APSSKSTSGG TAALGCLVKD
151 YPPEPVTVSW NSGALTSGVH TFPAVLQSSG LYSLSSVVTV PSSSLGTQTY
201 ICNVNHKPSN TKVDKKVEPK SCDKTHTCPP CEGQPREPQV YTLPFSRDEL
251 TKNQVSLTCL VKGFYPSDIA VEWESNGQPE NNYKTTPPVL DSDGSFFLYS
301 KLTVDKSRWQ QGNVFSCSVM HEALHNHYTQ KSLSLSPGK

INTER TIONAL SEARCH REPORT

em: al Application No. PCT/US 97/13562

CLASSIFICATION OF SUBJECT MATTER PC 6 C12N15/62 A61 A61K51/10 A61K47/48 A61K38/17 A61K39/395 IPC 6 C12N1/21 C12N15/13 C07K16/00 C07K16/46 C07K16/30 //C07K19/00 C12N5/10 According to International Patent Classification (IPC) or to both national classification and IPC B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) C07K A61K IPC 6 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Relevant to claim No Citation of document, with indication, where appropriate, of the relevant passages S. GILLIES ET AL.: "Antigen binding and 1-8. X 23-25 biological activities of engineered mutant chimeric antibodies with human tumor specificities." HUMAN ANTIBODIES AND HYBRIDOMAS, vol. 1, no. 1, 1990, STONEHAM, MA, USA, pages 47-54, XP002050448 see the whole document -/--Patent family members are listed in annex. Further documents are listed in the continuation of box C. "T" later document published after the international filing date Special categories of cited documents or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance "X" document of particular relevance; the claimed invention "E" earlier document but published on or after the international cannot be considered novel or cannot be considered to fiting date involve an inventive step when the document is taken alone *L* document which may throw doubts on priority claim(s) or which is caled to establish the publication date of another citation or other special reason (as specified) "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such docu-*O* document referring to an oral disclosure, use, exhibition or ments, such combination being obvious to a person skilled other mean *P* document published prior to the international filing date but "&" document member of the same patent family later than the priority date claimed Date of mailing of the international search report Date of the actual completion of the international search 2 1. D1. 98 17 December 1997 Authorized officer Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Nooij, F

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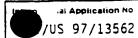
Fax: (+31-70) 340-3016

METION) DOCUMENTS CONSIDERED TO BE RELEVANT	
Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
G. SCHREIBER ET AL.: "An unmodified anticarcinoma antibody, BR96, localizes to and inhibits the outgrowth of human tumors in nude mice." CANCER RESEARCH, vol. 52, no. 12, 15 June 1992, BALTIMORE, MD, USA, pages 3262-3266, XP002050449	33,35,36
see abstract	1,2,5,7, 8,11-18, 23
A. DUNCAN ET AL.: "The binding site for Clq on IgG." NATURE, vol. 332, no. 6166, 21 April 1988, LONDON, GB, pages 738-740, XP002050450 cited in the application see the whole document	1,2,5,7,
J. LUND ET AL.: "Human FcgammaRI and FcgammaRII interact with distinct but overlapping sites on human IgG." THE JOURNAL OF IMMUNOLOGY, vol. 147, no. 8, 15 October 1991, BALTIMORE, MD, USA, pages 2657-2662, XP002050451 cited in the application see abstract	1.2,5,7,
Y. XU ET AL.: "Residue at position 331 in the IgG1 and IgG4 CH2 domains contributes to their differential ability to bind and activate complement." THE JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 269, no. 5, 4 February 1994, BALTIMORE, MD, USA, pages 3469-3474, XP002050452 cited in the application see abstract see discussion	1-8
T. MICHAELSEN ET AL.: "One disulfide bond in front of the second heavy chain constant region is necessary and sufficient for effector functions of human IgG3 without a genetic hinge." PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE USA, vol. 91, no. 20, 27 September 1994, WASHINGTON, DC, USA, pages 9243-9247, XP002050453 see the whole document	1,2,5,7,
	G. SCHREIBER ET AL.: "An unmodified anticarcinoma antibody, BR96, localizes to and inhibits the outgrowth of human tumors in nude mice." CANCER RESEARCH, vol. 52, no. 12, 15 June 1992, BALTIMORE, MD, USA, pages 3262-3266, XP002050449 see abstract A. DUNCAN ET AL.: "The binding site for C1q on IgG." NATURE, vol. 332, no. 6166, 21 April 1988, LONDON, GB, pages 738-740, XP002050450 cited in the application see the whole document J. LUND ET AL.: "Human FcgammaRI and FcgammaRII interact with distinct but overlapping sites on human IgG." THE JOURNAL OF IMMUNOLOGY, vol. 147, no. 8, 15 October 1991, BALTIMORE, MD, USA, pages 2657-2662, XP002050451 cited in the application see abstract Y. XU ET AL.: "Residue at position 331 in the IgG1 and IgG4 CH2 domains contributes to their differential ability to bind and activate complement." THE JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 269, no. 5, 4 February 1994, BALTIMORE, MD, USA, pages 3469-3474, XP002050452 cited in the application see abstract see discussion T. MICHAELSEN ET AL.: "One disulfide bond in front of the second heavy chain constant region is necessary and sufficient for effector functions of human IgG3 without a genetic hinge." PROCEEDINGS OF THE USA, vol. 91, no. 20, 27 September 1994, WASHINGTON, DC, USA, VASHINGTON, DC, USA, VASHINGTO

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INTERNATIONAL SEARCH REPORT

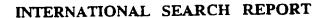


		/05 97/13502
C.(Continua	BOON) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category '	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No
A	L. TAN ET AL.: "Influence of the hinge region on complement activation, Clq binding, and segmental flexibility in chimeric human immunoglobulins." PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE USA, vol. 87, no. 1, January 1990, WASHINGTON, DC, USA, pages 162-166, XP002050454 see the whole document	1-8
A	EP 0 699 756 A (BRISTOL-MYERS SQUIBB COMPANY) 6 March 1996 cited in the application	11-18, 23,25, 28,29, 31-52
	see examples see claims	
	300 01011113	
		·

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Box i	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This Inte	ernational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: see FURTHER INFORMATION sheet PCT/ISA/210
2. X	Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically: see FURTHER INFORMATION sheet PCT/ISA/210
з. 🗀	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Inte	ernational Searching Authority found multiple inventions in this international application, as follows:
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were limely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4.	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remari	The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

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FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

Claims Nos.: 26,27

because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

Claim 26 represents a method of detection/diagnosis and refers forward to claim 30, which represents a method of treatment. Claim 27 refers to a method in claim 24; however, in claim 24 a product is claimed, not a method.

Remark: Although claims 1-22, 25, 28-32 and 34-36 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.

BNSDOCID: <WO 9805787A1>

Information on patent family members

Indian 1al Application No PCT/US 97/13562

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